



Conference on 'PUFA mediators: implications for human health' Symposium 3: Cannabinoids in human health

Elevating endocannabinoid levels: pharmacological strategies and potential therapeutic applications

Roger G. Pertwee

School of Medical Sciences, Institute of Medical Sciences, University of Aberdeen, Foresterhill, Aberdeen, AB25 2ZD, Scotland

The endocannabinoid system consists of cannabinoid CB₁ and CB₂ receptors, of endogenous agonists for these receptors known as 'endocannabinoids', and of processes responsible for endocannabinoid biosynthesis, cellular uptake and metabolism. There is strong evidence first, that this system up-regulates in certain disorders as indicated by an increased release of endocannabinoids onto their receptors and/or by increases in the expression levels or coupling efficiency of these receptors, and second, that this up-regulation often appears to reduce or abolish unwanted effects of these disorders or to slow their progression. This discovery has raised the possibility of developing a medicine that enhances up-regulation of the endocannabinoid system associated with these disorders by inhibiting the cellular uptake or intracellular metabolism of an endocannabinoid following its 'autoprotective' endogenous release. For inhibition of endocannabinoid metabolism, research has focused particularly on two highly investigated endocannabinoids, anandamide and 2-arachidonoyl glycerol, and hence on inhibitors of the main anandamide-metabolising enzyme, fatty acid amide hydrolase (FAAH), and of the main 2-arachidonoyl glycerol-metabolising enzyme, monoacylglycerol (MAG) lipase. The resulting data have provided strong preclinical evidence that selective FAAH and MAG lipase inhibitors would ameliorate the unwanted effects of several disorders, when administered alone or with a cyclooxygenase inhibitor, and that the benefit-to-risk ratio of a FAAH inhibitor would exceed that of a MAG lipase inhibitor or dual inhibitor of FAAH and MAG lipase. Promising preclinical data have also been obtained with inhibitors of endocannabinoid cellular uptake. There is now an urgent need for clinical research with these enzyme and uptake inhibitors.

**Fatty acid amide hydrolase inhibitors: Monoacylglycerol lipase inhibitors:
Endocannabinoids: Anandamide and 2-arachidonoyl glycerol:
Cannabinoid CB₁ and CB₂ receptors**

Endocannabinoids are endogenously produced compounds that can target cannabinoid CB₁ and/or CB₂ receptors, both of which are G protein-coupled⁽¹⁾. The most investigated of these endocannabinoids have been arachidonylethanolamide (anandamide) and 2-arachidonoyl glycerol, each of which can activate both CB₁ and CB₂ receptors⁽¹⁾. Endocannabinoids and

cannabinoid CB₁ and CB₂ receptors, together with the processes responsible for endocannabinoid biosynthesis, cellular uptake and metabolism, constitute the 'endocannabinoid system'⁽²⁾. Importantly, there is convincing evidence, albeit mainly from animal experiments, that there are a number of disorders in which the endocannabinoid system up-regulates in a manner that reduces or abolishes

Abbreviations: ABHD, α,β -hydrolase; COX, cyclooxygenase; FAAH, fatty acid amide hydrolase; MAG, monoacylglycerol; NSAID, non-steroidal anti-inflammatory drug; TRP, transient receptor potential.

Corresponding author: Professor R. G. Pertwee, fax 01224 437465, email rgp@abdn.ac.uk

the unwanted effects of these disorders or even slows disease progression. Signs of this 'autoprotective' up-regulation include increases in endocannabinoid release, in the expression levels of cannabinoid CB₁ or CB₂ receptors and/or in the coupling efficiency of these receptors⁽²⁾. This evidence has prompted research directed at seeking out the best strategies for exploiting the production of such autoprotection by the endocannabinoid system in the clinic^(2,3). Currently attracting particular attention are a number of 'indirect' potential strategies that involve inhibiting the cellular uptake or intracellular metabolism of an endocannabinoid following its endogenous release with the objective of increasing its concentrations at sites in the body at which it is inducing beneficial effects. The remainder of this paper focuses on these 'indirect' strategies and their therapeutic potential.

Potential therapeutic applications for inhibitors of endocannabinoid metabolism

For the inhibition of endocannabinoid metabolism, attention has so far focused largely on two enzymes: fatty acid amide hydrolase (FAAH), which is a major metabolising enzyme of anandamide but can also metabolise 2-arachidonoyl glycerol, and monoacylglycerol (MAG) lipase, which is thought to be largely but not exclusively responsible for the metabolism of 2-arachidonoyl glycerol (reviewed in^(2,10)). It is noteworthy that there are now known to be at least two forms of FAAH, FAAH-1 and FAAH-2 (which is not expressed by rodents), and also that anandamide and 2-arachidonoyl glycerol can be metabolised to varying extents by a number of enzymes in addition to FAAH or MAG lipase^(4,5,49). Thus, anandamide can be metabolised by *N*-acyl ethanolamine hydrolysing acid amidase, and 2-arachidonoyl glycerol by MAG kinase, α , β -hydrolase (ABHD)6 and ABHD12. Furthermore, both these endocannabinoids are good substrates for cytochrome P450 enzymes, lipoxygenases and cyclooxygenase (COX)-2. MAG lipase, ABHD6 and ABHD12 are all serine hydrolases. However, unlike MAG lipase, which is present on the intracellular surface of cell membranes, these other two hydrolases are both located within the membranes of the cells in which they are expressed, with the active site of ABHD12 facing the outer surface of these membranes and that of ABHD6 located on their inner, cytoplasmic surface⁽⁶⁾. MAG lipase, ABHD12, ABHD6 and other enzymes (including FAAH) are thought to account, respectively, for about 85, 9, 4 and 2% of total brain-membrane 2-arachidonoyl glycerol hydrolase activity⁽⁶⁾. Within the brain, ABHD6 seems to be expressed by neurones and often to be located postsynaptically, close to neurones that express presynaptic CB₁ receptors, whereas ABHD12 seems to be expressed by non-neuronal cells, for example microglia⁽⁷⁾.

Interestingly, there is evidence that some of the metabolites formed from endogenously produced anandamide or 2-arachidonoyl glycerol by enzymes other than FAAH and MAG lipase are pharmacologically active.

For example, one COX-2 metabolite of anandamide is PGH₂ ethanolamide, which is further metabolised to J-series PG that have been found to induce apoptosis in COX-2 overexpressing tumourigenic keratinocytes, an effect that could be enhanced by an irreversible FAAH inhibitor (URB597;⁽⁸⁾ (Fig. 1a). It has also been found that COX-2 metabolises 2-arachidonoyl glycerol to the hyperalgesic prostanoid, PGE₂ glycerol ester, an observation that has prompted the hypothesis that COX-2 could act as an enzymatic switch with the capacity to convert 2-arachidonoyl glycerol from an antinociceptive mediator to a pronociceptive prostanoid⁽⁹⁾.

FAAH inhibitors other than URB597 (Org231295) include both the reversible inhibitor, OL135 (Fig. 1a) and O-1887, which, like URB597, is an irreversible inhibitor⁽¹⁰⁻¹²⁾. These can all inhibit FAAH *in vitro* at concentrations that lie in the low nanomolar range and do not displace radiolabelled ligands from CB₁ receptors or (where this has been investigated) from CB₂ receptors. It is noteworthy, however, that the pharmacological profiles of most FAAH inhibitors have not been completely investigated. Just a few additional examples of the many FAAH inhibitors that have now been developed include URB532 (Fig. 1a), AM374 (palmitylsulphonyl fluoride), *N*-arachidonoyl glycine and *N*-arachidonoyl serotonin⁽²⁾, JNJ1661010 and CAY10401⁽¹³⁾, AM3506 and AM5206^(14,15), ST4070^(16,17), PF3845⁽¹⁸⁾ and PF04457845, a particularly FAAH-selective structural analogue of PF3845⁽¹⁹⁾ (Fig. 1a).

Evidence has already emerged from animal experiments that FAAH inhibitors are effective at ameliorating signs of acute, inflammatory, visceral and neuropathic pain^(2,4,13,17-24), and that URB597 can ameliorate the signs of both osteoarthritic pain⁽²⁵⁾, and referred hyperalgesia induced by bladder inflammation⁽²⁶⁾. Animal experiments have also yielded data suggesting that FAAH inhibitors might have a number of other therapeutic applications (Table 1). Importantly, there is good evidence that anandamide displays significant potency not only as a cannabinoid CB₁ and CB₂ receptor agonist, but also at activating or inhibiting certain voltage-gated ion channels, at enhancing the activation of NMDA and glycine ligand-gated ion channels, at blocking transient receptor potential (TRP) M8 cation channels and at allosterically antagonising the activation of 5-hydroxytryptamine₃ and nicotinic acetylcholine receptors⁽¹⁾. It is also well-established, first, that anandamide can activate TRPV1 and TRPA1 cation channels, albeit with less potency than it activates cannabinoid receptors⁽¹⁾, and second, that FAAH can metabolise pharmacologically active endogenous compounds other than anandamide, just one example being oleoylethanolamide which is not a cannabinoid receptor ligand but can activate both TRPV1 cation channels and the PPAR α ⁽⁴⁹⁾. It is possible too that reductions in signs of nicotine-induced reward evoked by URB597 may be PPAR α -mediated since this FAAH inhibitor seems to produce this effect mainly by elevating oleoylethanolamide and palmitoylethanolamide, which is also an endogenous PPAR α ligand⁽⁴⁰⁾. It is noteworthy, therefore, that there have



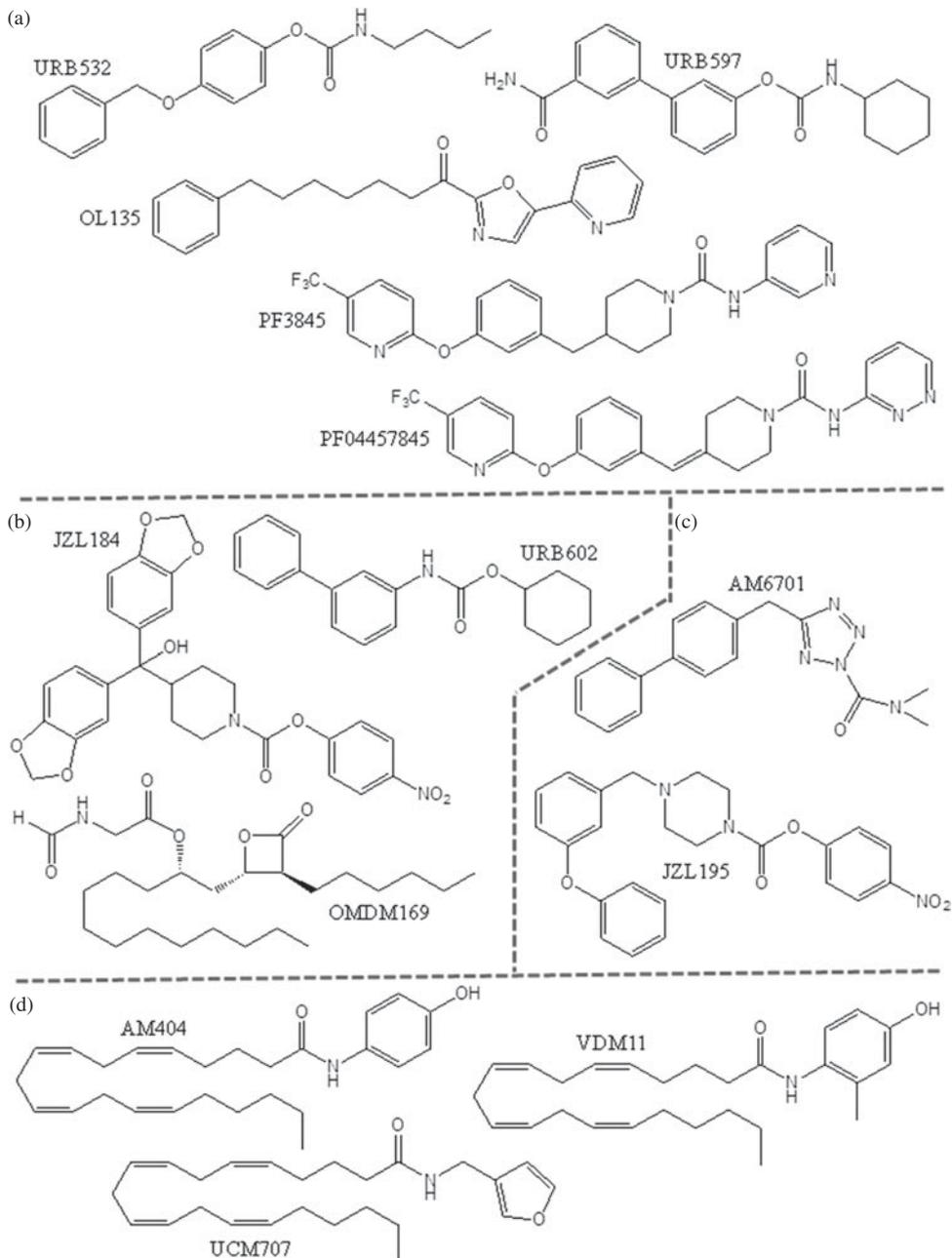


Fig. 1. Structures of a selection of inhibitors of (a) fatty acid amide hydrolase (FAAH), (b) monoacylglycerol lipase (MAG lipase), (c) dual inhibitors of FAAH and MAG lipase and (d) the cellular uptake of anandamide.

been several investigations in which a selective CB₁ or CB₂ receptor antagonist^(17,23,24,50,51), or genetic deletion of the CB₁ receptor⁽⁵²⁾, has been found to attenuate signs of antinociception induced by URB597, PF3845 or ST4070 in rodent models of visceral, inflammatory or neuropathic pain. In two of these investigations, however, the TRPV1 antagonist, 5'-iodoresiniferatoxin,⁽⁵¹⁾ or the PPAR α antagonist, GW6471^(17,24), was also found to block at least some antinociceptive effects of URB597 or ST4070, whereas in another of these investigations no such antagonism was detected⁽⁵²⁾. In addition, there have been reports that a CB₁ receptor antagonist

can block AM3506-induced inhibition of abnormal gastrointestinal motility and faecal output in mice⁽³¹⁾, and also URB597-induced reductions of vomiting in shrews and of signs of nausea in rats⁽⁴⁵⁾, and that both a CB₁ and a CB₂ receptor antagonist can block URB597-induced inhibition of scratching behaviour that had been evoked by 5-hydroxytryptamine in rats⁽³⁶⁾.

Moving on to MAG lipase inhibitors, these include (Fig. 1b) the non-competitive/irreversible inhibitors, URB602 and JZL184, and the reversible inhibitor, OMDM169⁽¹³⁾. Like inhibitors of FAAH, MAG lipase inhibitors display antinociceptive activity in a number

Table 1. Some potential therapeutic targets other than pain identified in preclinical investigations for fatty acid amide hydrolase inhibitors

Potential therapeutic targets	Species investigated	References
Signs and symptoms of multiple sclerosis	Mouse	2*, 13*
Signs and symptoms of Parkinson's disease	Rat	2*
Hyperactivity/dyskinesia induced by L-DOPA when it is used to treat Parkinson's disease	Monkey (marmoset)	27
Excitotoxic events, including seizures and stroke	Rat; mouse	2*, 15, 28
Hypertension	Rat	14
Inflammatory bowel diseases	Human; rat; and mouse	13*, 29* and 30
Abnormal gastrointestinal motility in acute gastroenteritis and IBS	Mouse	31
Gastric ulcers/irritation caused by NSAID	Mouse	32*, 33 and 34
Pruritus	Rat; mouse	13*, 35 and 36
Depression	Rat; mouse	13*
Anxiety-related disorders, including obsessive compulsive disorder	Mouse	13*, 20, 37 and 38
Nicotine dependence	Rat	39 and 40
Physical signs of morphine and of Δ^9 -THC withdrawal	Rat; mouse	41, 42 and 43*
Anxiety, but not other signs (physical signs), induced by nicotine withdrawal	Rat	44
Nausea and vomiting produced, for example, by chemotherapy	Rat (nausea); ferret or shrew (vomiting)	45*
Cancer	Human; rat; and mouse	2*, 13*, 46, 47* and 48

IBS, irritable bowel syndrome; L-DOPA, L-3,4-dihydroxyphenylalanine; NSAID, non-steroidal anti-inflammatory drug; Δ^9 -THC, Δ^9 -tetrahydrocannabinol.

* Review article.

of different animal models that for one or more of these compounds include models of acute, visceral, inflammatory, neuropathic and/or bone cancer pain^(4,13,20–22,43,53–57). There is evidence too, that in rats, MAG lipase inhibitors can reduce signs of inflammatory pain not only when administered systemically but also when injected directly into the spinal cord or inflamed hind paws^(58–60). Moreover, results obtained from animal experiments with JZL184 suggest that, like FAAH inhibitors, MAG lipase inhibitors have several other potential therapeutic applications (Table 2). There is evidence too that MAG lipase inhibitors might be effective against breast, ovarian, skin and prostate cancer⁽⁴³⁾. Thus, for example, it has been found that JZL184 can reduce prostate cancer (PC3) cell migration, invasion and survival *in vitro* and slow PC3 tumour xenograft growth in immune-deficient SCID mice, although interestingly, not in nude mice, probably because its inhibitory effect on MAG lipase was less in the tumours grown in nude mice than in those grown in SCID mice⁽⁶⁵⁾. Evidence has also emerged that JZL184 and URB602 can protect neurons from β -amyloid peptide-induced neurodegeneration and apoptosis, raising the possibility that Alzheimer's disease could be another suitable therapeutic target for the MAG lipase inhibitors^(43,66–68).

Like anandamide, 2-arachidonoyl glycerol, does not target only cannabinoid CB₁ and CB₂ receptors. Thus, there is evidence first, that at concentrations similar to those at which it activates these receptors, 2-arachidonoyl glycerol can inhibit K_v4.3 voltage-gated ion channels and allosterically antagonise the activation of nicotinic acetylcholine receptors, and second, that at higher concentrations, it can also target a number of other receptors or ion channels, including glycine, γ -aminobutyric acid_A and adenosine A₃ receptors and certain voltage-gated calcium channels^(1,69). Even so, at least some of the effects that the MAG lipase inhibitors

Table 2. Some potential therapeutic targets other than pain identified in preclinical investigations for monoacylglycerol lipase inhibitors

Potential therapeutic targets	Species investigated	References
Inflammatory bowel diseases	Mouse	29*, 61
Gastric ulcers/irritation caused by NSAID	Mouse	62
Pruritus	Rat	36
Anxiety-related disorders	Rat; mouse	20, 43*, 55*, 63
Parkinson's disease	Mouse	43*
Traumatic brain injury	Mouse	2*
Physical signs of morphine and of Δ^9 -THC withdrawal	Mouse	41, 43*
Vomiting produced, for example, by chemotherapy	Ferret; shrew	45*, 64

NSAID, non-steroidal anti-inflammatory drugs; Δ^9 -THC, Δ^9 -tetrahydrocannabinol.

* Review article.

have been reported to produce *in vivo*, appear to have been partly or wholly cannabinoid-receptor mediated. More specifically, there have been reports that the CB₁-selective antagonist, AM251 reduces the ability of JZL184 to attenuate vomiting in shrews and signs of nausea in rats⁽⁶⁴⁾, that AM251 and the CB₂-selective antagonist, AM630, both prevent JZL184-induced reductions of 5-hydroxytryptamine-evoked scratch bouts in rats⁽³⁶⁾, and that these two antagonists can also prevent JZL184 from reducing signs of colitis in mice⁽⁶¹⁾. Evidence has also been obtained from experiments performed with CB₁ and CB₂ receptor-deficient mice, that reductions in signs of neuropathic pain induced by JZL184 were mediated by CB₁ but not CB₂ receptors⁽²¹⁾. In contrast, the production of this effect by the FAAH inhibitor, PF3845, seemed to depend on the activation of both the CB₁ and CB₂ receptors. It is noteworthy too, that

Busquets-Garcia *et al.*⁽²⁰⁾ have found that an anxiolytic-like effect produced by JZL184 in mice seemed to be CB₂-receptor mediated since it could be abolished firstly, by the CB₂-selective antagonists, SR144528 and AM630, but not by the CB₁-selective antagonist, SR141716A (rimonabant), and secondly, by genetic deletion of the CB₂ receptor but not of the CB₁ receptor. On the other hand, the FAAH inhibitor, URB597, produced an anxiolytic-like effect in mice that appeared to be CB₁ but not CB₂ receptor-mediated.

The evidence that some of the apparently beneficial effects of FAAH and MAG lipase inhibitors appear to be partly or wholly cannabinoid CB₁ receptor-mediated, prompts the question of whether these inhibitors also produce some or all of the unwanted effects that can result from direct CB₁ receptor activation. It is noteworthy, therefore, that there have been reports that unlike direct CB₁ agonists, it is possible for FAAH inhibitors to produce antinociception in mice at doses that do not also induce hypomotility, hypothermia, catalepsy or hyperphagia⁽⁷⁰⁻⁷²⁾. It is also unlikely that a FAAH inhibitor would share the ability of a direct CB₁ receptor agonist to induce physical or psychological dependence. Thus, repeated administration of one FAAH inhibitor, PF3845, has been found not to produce signs of physical dependence in mice⁽⁷³⁾, and the results obtained with another FAAH inhibitor, URB597, in experiments with squirrel monkeys, suggest that such inhibitors do not possess reinforcing properties and would not trigger relapse to drug abuse⁽⁷⁴⁾. Since tolerance can develop to antinociceptive effects of direct cannabinoid receptor agonists⁽⁷³⁾, it is noteworthy too, however, that there has been a report that tolerance appeared to develop to the ability of URB597 to attenuate inflammatory pain behaviour in rats, although not to its ability to inhibit FAAH, when this compound was administered repeatedly⁽²⁴⁾.

With regard to MAG lipase inhibitors, there has been a report that JZL184 shares the ability of direct CB₁ receptor agonists to induce both hypothermia and hypomotility, although not catalepsy, at an antinociceptive dose⁽⁵⁷⁾. Moreover, when it was administered repeatedly to mice, this MAG lipase inhibitor was found to produce physical dependence, as indicated by SR141716A-precipitated paw fluttering, and to cause tolerance to develop both to its own antinociceptive effect, unless it was administered at a low antinociceptive dose, and to that of the direct CB₁/CB₂ receptor agonists, Δ⁹-tetrahydrocannabinol and *R*-(+)-WIN55212^(56,73). In line with some of these observations is the finding that both JZL184 and the direct CB₁/CB₂ receptor partial agonist, Δ⁹-tetrahydrocannabinol, could reduce four naloxone precipitated physical withdrawal signs in morphine-dependent mice⁽⁴¹⁾. In contrast, only two of these withdrawal signs could be attenuated by the FAAH inhibitor, PF3845. Finally, the data obtained by Busquets-Garcia *et al.*⁽²⁰⁾ from mouse experiments suggest that a dose of JZL184 that induced apparent CB₂ receptor-mediated reductions in anxiety, did not affect memory, whereas memory was impaired by URB597 at a dose that induced apparent CB₁

receptor-mediated anxiolysis. No tolerance developed to the anxiolytic-like effect of JZL184 or URB597, or to the memory-impairing effect of URB597, when either of these two compounds was administered repeatedly.

Clearly, further research is required to establish whether a FAAH-inhibitor is likely to have a significantly higher or lower benefit-to-risk ratio than a MAG lipase inhibitor for each of the potential therapeutic applications that they have in common. It is noteworthy, however, that there is already strong evidence that in order to optimise the benefit-to-risk ratio of any new medicine that acts by inhibiting endocannabinoid metabolism, it will be important to ensure that this medicine does not inhibit both these enzymes at the same time since it is likely that the therapeutically beneficial and adverse effects produced by dual inhibitors of FAAH and MAG lipase would closely resemble those that are produced by direct cannabinoid receptor agonists. Thus, signs of central CB₁ receptor activation that were very similar to those produced by a direct CB₁ receptor agonist and included hypomotility, antinociception and catalepsy, have been found to be displayed by mice in response to such dual inhibition, by certain organophosphorus nerve agents⁽⁷⁵⁾, and by JZL195 (Fig. 1c), a compound that inactivates both FAAH and MAG lipase *in vivo* with high efficacy, and is also an inhibitor of ABHD6^(4,76). More importantly, these dual inhibitor-induced effects were blocked by pretreatment with a CB₁-selective antagonist: either AM251 or SR141716A. It has also been found that mice treated with JZL195 displayed signs of impaired short-term spatial memory that were similar in magnitude to those produced by the direct CB₁/CB₂ receptor agonist, Δ⁹-tetrahydrocannabinol, and could be prevented by SR141716A⁽⁷⁷⁾. A similar effect on memory was induced by the MAG lipase inhibitor, JZL184, in mice from which FAAH had been genetically deleted⁽⁷⁷⁾. Interestingly, however, another dual inhibitor of FAAH and MAG lipase, AM6701 (Fig. 1c), has been reported to reduce both severity of seizures and impairment of rotarod and balance beam performance induced in rats by kainic acid with significantly greater efficacy than AM6702, a compound that is significantly more potent at inhibiting FAAH than MAG lipase⁽⁷⁸⁾.

A dual inhibitor of FAAH and MAG lipase might of course have an acceptable benefit-to-risk ratio if, for example, its distribution is restricted to one or more particular regions of the body. Indeed, there is already evidence that one possible strategy would be to administer a dual inhibitor to the skin for the treatment of cancer pain⁽⁷⁹⁾. An alternative possibility would be to administer a peripherally-restricted dual inhibitor of FAAH and MAG lipase, or indeed to co-administer a FAAH inhibitor and a MAG lipase inhibitor, either or both of which are peripherally restricted. One example of a peripherally-restricted FAAH inhibitor that has already been developed is URB937⁽⁸⁰⁾, a compound that has been reported to exert profound antinociceptive effects in rodent models of visceral, inflammatory and

neuropathic pain⁽⁸¹⁾. Since the hydrolases, MAG lipase, ABHD6 and ABHD12, most probably metabolise distinct pools of 2-arachidonoyl glycerol *in vivo*⁽⁶⁾, further research that explores the potential therapeutic advantages of selectively inhibiting just one or other of these enzymes, or indeed of simultaneously inhibiting two or even all three of these enzymes, is also clearly warranted. It is noteworthy, therefore, that a selective inhibitor of ABHD6 was developed in the same year that this enzyme was found to metabolise 2-arachidonoyl glycerol^(4,6,82).

Evidence has also emerged from animal experiments not only that administration of a FAAH inhibitor in combination with a COX inhibitor/non-steroidal anti-inflammatory drug (NSAID) would be effective for the amelioration of inflammatory and neuropathic pain⁽³⁴⁾ and visceral pain^(32,33), but also, as already mentioned, that FAAH inhibition lowers the unwanted ulcerogenic activity of COX inhibitors/NSAID^(32–34,83). It is noteworthy, therefore, that carprofen, a compound that has been reported to produce fewer ulcerogenic side effects in human subjects than other NSAID, has been found to be a multi-inhibitor of COX-1, COX-2 and FAAH⁽⁸³⁾. It is noteworthy, too, first that the peripherally restricted FAAH inhibitor, URB937, has been found in mice both to interact synergistically with the NSAID, indomethacin, to reduce signs of inflammatory and neuropathic pain and to lower the number and severity of indomethacin-induced gastric lesions⁽³⁴⁾, and second, that the MAG lipase inhibitor, JZL184, has also been found to protect against NSAID-induced gastric damage in mice⁽⁶²⁾. Other potential adjunctive therapeutic strategies include the combined administration of a FAAH inhibitor, with a COX-2 inhibitor and a low dose of a glucocorticoid for rheumatoid arthritis⁽⁸⁴⁾, with palmitoylethanolamide for the treatment of melanoma⁽⁸⁵⁾, with morphine for relieving visceral pain⁽²³⁾, and with an inhibitor of the metabolism of endogenous opioids (enkephalins) for the alleviation of inflammatory and neuropathic pain⁽⁸⁶⁾.

Finally, there has been one recent published report about a clinical trial performed with the FAAH inhibitor, PF04457845⁽⁸⁷⁾. This was a multicentre, randomised, double-blind, placebo-controlled trial in which patients experiencing pain due to osteoarthritis of the knee received repeated doses of PF04457845, which has a good human safety profile^(87,88), of naproxen which served as a positive control, or of placebo. Unexpectedly, it was found that although PF04457845 did seem to inhibit FAAH, as indicated by its ability to elevate the plasma levels of anandamide, palmitoylethanolamide, oleoylethanolamide and linoleoylethanolamide in patients, it did not share the ability of naproxen to reduce osteoarthritic pain in this trial. Given the many positive results that have been obtained with FAAH inhibitors in animal models of pain, including rat and guinea-pig models of osteoarthritic pain⁽²⁵⁾, the inability of PF04457845 to relieve human osteoarthritic pain at a dose that does seem to inhibit FAAH prompts an urgent need for further research.

Potential therapeutic applications for inhibitors of endocannabinoid cellular uptake

A number of compounds that can inhibit the cellular uptake of anandamide have been developed. These include AM404, VDM11 and UCM707 (Fig. 1d), as well as OMDM1, OMDM2 and LY2183240^(2,10), one or more of which has been found to decrease cancer cell proliferation and tumour growth and to be effective in animal models against signs of pain, multiple sclerosis, excitotoxicity, Parkinson's disease, Huntington's disease, anxiety, cholera toxin-induced diarrhoea and colitis^(2,10,23). In addition, evidence has recently emerged that reinstatement of nicotine-seeking behaviour can be inhibited by both VDM11 and AM404 in rats, and that the latter compound can prevent the development of nicotine dependence in rats as well^(89,90). Evidence has also been obtained, from mouse experiments, that AM404 might be effective against obsessive compulsive disorder⁽³⁷⁾. In addition to inhibiting the cellular uptake of anandamide, AM404 can activate and desensitise TRPV1 receptors, and inhibit COX-1, COX-2 and PG synthesis^(49,91). Even so, this compound has been found to reduce signs of visceral pain in mice in a manner that seems to be CB₁ receptor-mediated⁽²³⁾. Importantly, LY2183240 is an inhibitor not only of endocannabinoid cellular uptake but also of FAAH and MAG lipase, and VDM11, UCM707 and AM404, which is a FAAH substrate, also have the ability to reduce the metabolism of anandamide and other acylethanolamides by FAAH^(10,90).

Although endocannabinoids clearly do pass through cell membranes, the question of whether or not this process is mediated by a transporter has been a topic of debate for many years⁽⁴⁹⁾. It is noteworthy, therefore, that there is now evidence for the existence of an anandamide-selective intracellular binding protein that may facilitate first, the cellular uptake of anandamide by reducing the free concentration of anandamide within cells; second, the translocation of anandamide to intracellular organelles where FAAH-1 is located, and third, the release of anandamide from cells by increasing its intracellular transfer from its sites of biosynthesis within cell membranes^(49,92,93). This binding protein has been named FAAH-like anandamide transporter. ARN272, a compound that inhibits both the binding of anandamide to FAAH-like anandamide transporter in a competitive manner, and the intracellular accumulation of this endocannabinoid, has been found to attenuate signs of inflammatory pain in mice⁽⁹²⁾. There has been a report too that SB-FI-26, an inhibitor of other intracellular transporters of anandamide that are known as fatty acid binding proteins, also displays antinociceptive activity in a mouse model of inflammatory pain⁽⁹⁴⁾.

Conclusions and future directions

The present paper has reviewed evidence published up to March 2013, that signs and symptoms of a number of disorders could be ameliorated by administering an



inhibitor of the anandamide-metabolising enzyme, FAAH, of the 2-arachidonoyl glycerol-metabolising enzyme, MAG lipase or of endocannabinoid cellular uptake. Although the published findings obtained with these inhibitors are very promising, they have come almost entirely from experiments with animals, prompting an urgent need for clinical trials that build on these findings by seeking out disorders in which significant beneficial effects could be achieved by (i) administering selective inhibitors of FAAH, MAG lipase or endocannabinoid cellular uptake; (ii) simultaneously inhibiting FAAH and MAG lipase, for example with a dual inhibitor; (iii) targeting particular regions of the body, for example, by direct application of one or more of these inhibitors to the skin, or by administering peripherally restricted inhibitors; and (iv) inhibiting FAAH or MAG lipase simultaneously with either or both COX-1 and COX-2. The need for such research with patients is strengthened by the findings already obtained in clinical trials, first, that the FAAH inhibitor, PF04457845, did not reduce osteoarthritic pain, and second, that concomitant inhibition of COX-1, COX-2 and FAAH produces fewer ulcerogenic side effects in human subjects than inhibition of just COX-1 and COX-2 by an NSAID. In addition, it will be important to search for potential therapeutic strategies that exploit: (i) the existence of at least two subtypes of FAAH; FAAH-1 and FAAH-2, and (ii) the recently discovered presence in the brain of a new set of endocannabinoids: lipoxin A₄, which behaves as an allosteric enhancer of anandamide-induced activation of the CB₁ receptor^(95,96), and an endogenous peptide, named Pepcan-12, that can weaken CB₁ receptor activation by direct agonists, seemingly by acting as a negative allosteric modulator at these receptors⁽⁹⁷⁾.

Acknowledgements

R. G. P. is grateful for financial support from both GW Pharmaceuticals and the National Institute on Drug Abuse.

Financial Support

The work was supported by Research grants from the National Institute on Drug Abuse (grant numbers DA-009789 and DA-03672) and funding from GW Pharmaceuticals.

Conflicts of Interest

None.

References

1. Pertwee RG, Howlett AC, Abood ME *et al.* (2010) International union of basic and clinical pharmacology. LXXIX. Cannabinoid receptors and their ligands: beyond CB₁ and CB₂. *Pharmacol Rev* **62**, 588–631.
2. Pertwee RG (2005) The therapeutic potential of drugs that target cannabinoid receptors or modulate the tissue levels or actions of endocannabinoids. *AAPS J* **7**, E625–E654.
3. Pertwee RG (2012) Targeting the endocannabinoid system with cannabinoid receptor agonists: pharmacological strategies and therapeutic possibilities. *Phil Trans R Soc B* **367**, 3353–3363.
4. Alvarez-Jaimes LJ & Palmer JA (2011) The role of endocannabinoids in pain modulation and the therapeutic potential of inhibiting their enzymatic degradation. *Curr Pharm Biotechnol* **12**, 1644–1659.
5. Zhao P & Abood ME (2013) GPR55 and GPR35 and their relationship to cannabinoid and lysophospholipid receptors. *Life Sci* **92**, 453–457.
6. Blankman JL, Simon GM & Cravatt BF (2007) A comprehensive profile of brain enzymes that hydrolyze the endocannabinoid 2-arachidonoylglycerol. *Chem Biol* **14**, 1347–1356.
7. Savinainen JR, Saario SM & Laitinen JT (2012) The serine hydrolases MAGL, ABHD6 and ABHD12 as guardians of 2-arachidonoylglycerol signalling through cannabinoid receptors. *Acta Physiol* **204**, 267–276.
8. Kuc C, Jenkins A & Van Dross RT (2012) Arachidonoyl ethanolamide (AEA)-induced apoptosis is mediated by J-series prostaglandins and is enhanced by fatty acid amide hydrolase (FAAH) blockade. *Mol Carcinog* **51**, 139–149.
9. Hu SS-J, Bradshaw HB, Chen JS-C *et al.* (2008) Prostaglandin E₂ glycerol ester, an endogenous COX-2 metabolite of 2-arachidonoylglycerol, induces hyperalgesia and modulates NFκB activity. *Br J Pharmacol* **153**, 1538–1549.
10. Di Marzo V (2008) Targeting the endocannabinoid system: to enhance or reduce? *Nat Rev Drug Discov* **7**, 438–455.
11. Martin BR, Beletskaya I, Patrick G *et al.* (2000) Cannabinoid properties of methylfluorophosphonate analogs. *J Pharmacol Exp Ther* **294**, 1209–1218.
12. Pertwee RG (2008) Ligands that target cannabinoid receptors in the brain: from THC to anandamide and beyond. *Addict Biol* **13**, 147–159.
13. Petrosino S & Di Marzo V (2010) FAAH and MAGL inhibitors: therapeutic opportunities from regulating endocannabinoid levels. *Curr Opin Investig Drugs* **11**, 51–62.
14. Godlewski G, Alapafuja SO, Bátkai S *et al.* (2010) Inhibitor of fatty acid amide hydrolase normalizes cardiovascular function in hypertension without adverse metabolic effects. *Chem Biol* **17**, 1256–1266.
15. Naidoo V, Nikas SP, Karanian DA *et al.* (2011) A new generation fatty acid amide hydrolase inhibitor protects against kainate-induced excitotoxicity. *J Mol Neurosci* **43**, 493–502.
16. Gattinoni S, De Simone C, Dallavalle S *et al.* (2010) Enol carbamates as inhibitors of fatty acid amide hydrolase (FAAH) endowed with high selectivity for FAAH over the other targets of the endocannabinoid system. *ChemMedChem* **5**, 357–360.
17. Caprioli A, Coccorello R, Rapino C *et al.* (2012) The novel reversible fatty acid amide hydrolase inhibitor ST4070 increases endocannabinoid brain levels and counteracts neuropathic pain in different animal models. *J Pharmacol Exp Ther* **342**, 188–195.
18. Ahn K, Johnson DS, Mileni M *et al.* (2009) Discovery and characterization of a highly selective FAAH inhibitor that reduces inflammatory pain. *Chem Biol* **16**, 411–420.
19. Johnson DS, Stiff C, Lazerwith SE *et al.* (2011) Discovery of PF-04457845: a highly potent, orally bioavailable, and



- selective urea FAAH inhibitor. *ACS Med Chem Lett* **2**, 91–96.
20. Busquets-Garcia A, Puighermanal E, Pastor A *et al.* (2011) Differential role of anandamide and 2-arachidonoylglycerol in memory and anxiety-like responses. *Biol Psychiatry* **70**, 479–486.
 21. Kinsey SG, Long JZ, Cravatt BF *et al.* (2010) Fatty acid amide hydrolase and monoacylglycerol lipase inhibitors produce anti-allodynic effects in mice through distinct cannabinoid receptor mechanisms. *J Pain* **11**, 1420–1428.
 22. Kinsey SG, Long JZ, O'Neal ST *et al.* (2009) Blockade of endocannabinoid-degrading enzymes attenuates neuropathic pain. *J Pharmacol Exp Ther* **330**, 902–910.
 23. Miller LL, Picker MJ, Umberger MD *et al.* (2012) Effects of alterations in cannabinoid signaling, alone and in combination with morphine, on pain-elicited and pain-suppressed behavior in mice. *J Pharmacol Exp Ther* **342**, 177–187.
 24. Okine BN, Norris LM, Woodhams S *et al.* (2012) Lack of effect of chronic pre-treatment with the FAAH inhibitor URB597 on inflammatory pain behaviour: evidence for plastic changes in the endocannabinoid system. *Br J Pharmacol* **167**, 627–640.
 25. Schuelert N, Johnson MP, Oskins JL *et al.* (2011) Local application of the endocannabinoid hydrolysis inhibitor URB597 reduces nociception in spontaneous and chemically induced models of osteoarthritis. *Pain* **152**, 975–981.
 26. Merriam FV, Wang Z-Y, Hillard CJ *et al.* (2011) Inhibition of fatty acid amide hydrolase suppresses referred hyperalgesia induced by bladder inflammation. *BJU Int* **108**, 1145–1149.
 27. Johnston TH, Huot P, Fox SH *et al.* (2011) Fatty acid amide hydrolase (FAAH) inhibition reduces L-3,4-dihydroxyphenylalanine-induced hyperactivity in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-lesioned non-human primate model of Parkinson's disease. *J Pharmacol Exp Ther* **336**, 423–430.
 28. Naderi N, Ahmad-Molaei L, Mazar-Atabaki A *et al.* (2012) L-type calcium channel mediates anticonvulsant effect of cannabinoids in acute and chronic murine models of seizure. *Neurochem Res* **37**, 279–287.
 29. Alhouayek M & Muccioli GG (2012) The endocannabinoid system in inflammatory bowel diseases: from pathophysiology to therapeutic opportunity. *Trends Mol Med* **18**, 615–625.
 30. Andrzejak V, Muccioli GG, Body-Malapel M *et al.* (2011) New FAAH inhibitors based on 3-carboxamido-5-aryl-isoxazole scaffold that protect against experimental colitis. *Bioorg Med Chem* **19**, 3777–3786.
 31. Bashashati M, Storr MA, Nikas SP *et al.* (2012) Inhibiting fatty acid amide hydrolase normalizes endotoxin-induced enhanced gastrointestinal motility in mice. *Br J Pharmacol* **165**, 1556–1571.
 32. Fowler CJ (2012) NSAIDs: eNdocannabinoid stimulating anti-inflammatory drugs? *Trends Pharmacol Sci* **33**, 468–473.
 33. Naidu PS, Booker L, Cravatt BF *et al.* (2009) Synergy between enzyme inhibitors of fatty acid amide hydrolase and cyclooxygenase in visceral nociception. *J Pharmacol Exp Ther* **329**, 48–56.
 34. Sasso O, Bertorelli R, Bandiera T *et al.* (2012) Peripheral FAAH inhibition causes profound antinociception and protects against indomethacin-induced gastric lesions. *Pharmacol Res* **65**, 553–563.
 35. Schlosburg JE, Boger DL, Cravatt BF *et al.* (2009) Endocannabinoid modulation of scratching response in an acute allergic model: a new prospective neural therapeutic target for pruritus. *J Pharmacol Exp Ther* **329**, 314–323.
 36. Spradley JM, Davoodi A, Gee LB *et al.* (2012) Differences in peripheral endocannabinoid modulation of scratching behavior in facial vs. spinally-innervated skin. *Neuropharmacology* **63**, 743–749.
 37. Gomes FV, Casarotto PC, Resstel LBM *et al.* (2011) Facilitation of CB1 receptor-mediated neurotransmission decreases marble burying behavior in mice. *Prog Neuro-Psychopharmacol Biol Psychiatry* **35**, 434–438.
 38. Kinsey SG, O'Neal ST, Long JZ *et al.* (2011) Inhibition of endocannabinoid catabolic enzymes elicits anxiolytic-like effects in the marble burying assay. *Pharmacol Biochem Behav* **98**, 21–27.
 39. Forget B, Coen KM & Le Foll B (2009) Inhibition of fatty acid amide hydrolase reduces reinstatement of nicotine seeking but not break point for nicotine self-administration – comparison with CB₁ receptor blockade. *Psychopharmacology* **205**, 613–624.
 40. Scherma M, Panlilio LV, Fadda P *et al.* (2008) Inhibition of anandamide hydrolysis by cyclohexyl carbamic acid 3'-carbamoyl-3-yl ester (URB597) reverses abuse-related behavioral and neurochemical effects of nicotine in rats. *J Pharmacol Exp Ther* **327**, 482–490.
 41. Ramesh D, Ross GR, Schlosburg JE *et al.* (2011) Blockade of endocannabinoid hydrolytic enzymes attenuates precipitated opioid withdrawal symptoms in mice. *J Pharmacol Exp Ther* **339**, 173–185.
 42. Shahidi S & Hasanein P (2011) Behavioral effects of fatty acid amide hydrolase inhibition on morphine withdrawal symptoms. *Brain Res Bull* **86**, 118–122.
 43. Mulvihill MM & Nomura DK (2013) Therapeutic potential of monoacylglycerol lipase inhibitors. *Life Sci* **92**, 492–497.
 44. Cippitelli A, Astarita G, Duranti A *et al.* (2011) Endocannabinoid regulation of acute and protracted nicotine withdrawal: effect of FAAH inhibition. *PLoS ONE* **6**, e28142.
 45. Parker LA, Rock EM & Limebeer CL (2011) Regulation of nausea and vomiting by cannabinoids. *Br J Pharmacol* **163**, 1411–1422.
 46. Endsley MP, Thill R, Choudhry I *et al.* (2008) Expression and function of fatty acid amide hydrolase in prostate cancer. *Int J Cancer* **123**, 1318–1326.
 47. Malfitano AM, Ciaglia E, Gangemi G *et al.* (2011) Update on the endocannabinoid system as an anticancer target. *Expert Opin Ther Targets* **15**, 297–308.
 48. Proto MC, Gazerro P, Di Croce L *et al.* (2012) Interaction of endocannabinoid system and steroid hormones in the control of colon cancer cell growth. *J Cell Physiol* **227**, 250–258.
 49. Piscitelli F & Di Marzo V (2012) “Redundancy” of endocannabinoid inactivation: new challenges and opportunities for pain control. *ACS Chem Neurosci* **3**, 356–363.
 50. Khasabova IA, Khasabov S, Paz J *et al.* (2012) Cannabinoid type-1 receptor reduces pain and neurotoxicity produced by chemotherapy. *J Neurosci* **32**, 7091–7101.
 51. Starowicz K, Makuch W, Osikowicz M *et al.* (2012) Spinal anandamide produces analgesia in neuropathic rats: possible CB₁- and TRPV1-mediated mechanisms. *Neuropharmacology* **62**, 1746–1755.
 52. Booker L, Kinsey SG, Abdullah RA *et al.* (2012) The fatty acid amide hydrolase (FAAH) inhibitor PF-3845 acts in the nervous system to reverse LPS-induced tactile allodynia in mice. *Br J Pharmacol* **165**, 2485–2496.
 53. Bisogno T, Ortar G, Petrosino S *et al.* (2009) Development of a potent inhibitor of 2-arachidonoylglycerol hydrolysis

- with antinociceptive activity *in vivo*. *Biochim Biophys Acta* **1791**, 53–60.
54. Comelli F, Giagnoni G, Bettoni I *et al.* (2007) The inhibition of monoacylglycerol lipase by URB602 showed an anti-inflammatory and anti-nociceptive effect in a murine model of acute inflammation. *Br J Pharmacol* **152**, 787–794.
 55. Fowler CJ (2012) Monoacylglycerol lipase – a target for drug development? *Br J Pharmacol* **166**, 1568–1585.
 56. Ghosh S, Wise LE, Chen Y *et al.* (2013) The monoacylglycerol lipase inhibitor JZL184 suppresses inflammatory pain in the mouse carrageenan model. *Life Sci* **92**, 498–505.
 57. Long JZ, Li W, Booker L *et al.* (2009) Selective blockade of 2-arachidonoylglycerol hydrolysis produces cannabinoid behavioral effects. *Nat Chem Biol* **5**, 37–44.
 58. Guindon J, Desroches J & Beaulieu P (2007) The antinociceptive effects of intraplantar injections of 2-arachidonoyl glycerol are mediated by cannabinoid CB₂ receptors. *Br J Pharmacol* **150**, 693–701.
 59. Woodhams SG, Wong A, Barrett DA *et al.* (2012) Spinal administration of the monoacylglycerol lipase inhibitor JZL184 produces robust inhibitory effects on nociceptive processing and the development of central sensitization in the rat. *Br J Pharmacol* **167**, 1609–1619.
 60. Guindon J, Guijarro A, Piomelli D *et al.* (2011) Peripheral antinociceptive effects of inhibitors of monoacylglycerol lipase in a rat model of inflammatory pain. *Br J Pharmacol* **163**, 1464–1478.
 61. Alhouayek M, Lambert DM, Delzenne NM *et al.* (2011) Increasing endogenous 2-arachidonoylglycerol levels counteracts colitis and related systemic inflammation. *FASEB J* **25**, 2711–2721.
 62. Kinsey SG, Nomura DK, O'Neal ST *et al.* (2011) Inhibition of monoacylglycerol lipase attenuates nonsteroidal anti-inflammatory drug-induced gastric hemorrhages in mice. *J Pharmacol Exp Ther* **338**, 795–802.
 63. Aliczki M, Balogh Z, Tulogdi A *et al.* (2012) The temporal dynamics of the effects of monoacylglycerol lipase blockade on locomotion, anxiety, and body temperature. *Behav Pharmacol* **23**, 348–357.
 64. Sticht MA, Long JZ, Rock EM *et al.* (2012) Inhibition of monoacylglycerol lipase attenuates vomiting in *Suncus murinus* and 2-arachidonoyl glycerol attenuates nausea in rats. *Br J Pharmacol* **165**, 2425–2435.
 65. Nomura DK, Lombardi DP, Chang JW *et al.* (2011) Monoacylglycerol lipase exerts dual control over endocannabinoid and fatty acid pathways to support prostate cancer. *Chem Biol* **18**, 846–856.
 66. Chen X, Zhang J & Chen C (2011) Endocannabinoid 2-arachidonoylglycerol protects neurons against β -amyloid insults. *Neuroscience* **178**, 159–168.
 67. Chen R, Zhang J, Wu Y *et al.* (2012) Monoacylglycerol lipase is a therapeutic target for Alzheimer's disease. *Cell Rep* **2**, 1329–1339.
 68. Piro JR, Benjamin DI, Duerr JM *et al.* (2012) A dysregulated endocannabinoid-eicosanoid network supports pathogenesis in a mouse model of Alzheimer's disease. *Cell Rep* **1**, 617–623.
 69. Sigel E, Baur R, Rácz I *et al.* (2011) The major central endocannabinoid directly acts at GABA_A receptors. *Proc Natl Acad Sci USA* **108**, 18150–18155.
 70. Cravatt BF & Lichtman AH (2004) The endogenous cannabinoid system and its role in nociceptive behavior. *J Neurobiol* **61**, 149–160.
 71. Kathuria S, Gaetani S, Fegley D *et al.* (2003) Modulation of anxiety through blockade of anandamide hydrolysis. *Nat Med* **9**, 76–81.
 72. Lichtman AH, Leung D, Shelton CC *et al.* (2004) Reversible inhibitors of fatty acid amide hydrolase that promote analgesia: evidence for an unprecedented combination of potency and selectivity. *J Pharmacol Exp Ther* **311**, 441–448.
 73. Schlosburg JE, Blankman JL, Long JZ *et al.* (2010) Chronic monoacylglycerol lipase blockade causes functional antagonism of the endocannabinoid system. *Nat Neurosci* **13**, 1113–1121.
 74. Justinova Z, Mangieri RA, Bortolato M *et al.* (2008) Fatty acid amide hydrolase inhibition heightens anandamide signaling without producing reinforcing effects in primates. *Biol Psychiatry* **64**, 930–937.
 75. Nomura DK, Blankman JL, Simon GM *et al.* (2008) Activation of the endocannabinoid system by organophosphorus nerve agents. *Nat Chem Biol* **4**, 373–378.
 76. Long JZ, Nomura DK, Vann RE *et al.* (2009) Dual blockade of FAAH and MAGL identifies behavioral processes regulated by endocannabinoid crosstalk *in vivo*. *Proc Natl Acad Sci USA* **106**, 20270–20275.
 77. Wise LE, Long KA, Abdullah RA *et al.* (2012) Dual fatty acid amide hydrolase and monoacylglycerol lipase blockade produces THC-like Morris water maze deficits in mice. *ACS Chem Neurosci* **3**, 369–378.
 78. Naidoo V, Karanian DA, Vadivel SK *et al.* (2012) Equipotent inhibition of fatty acid amide hydrolase and monoacylglycerol lipase – dual targets of the endocannabinoid system to protect against seizure pathology. *Neurotherapeutics* **9**, 801–813.
 79. Khasabova IA, Chandiramani A, Harding-Rose C *et al.* (2011) Increasing 2-arachidonoyl glycerol signaling in the periphery attenuates mechanical hyperalgesia in a model of bone cancer pain. *Pharmacol Res* **64**, 60–67.
 80. Moreno-Sanz G, Barrera B, Guijarro A *et al.* (2011) The ABC membrane transporter ABCG2 prevents access of FAAH inhibitor URB937 to the central nervous system. *Pharmacol Res* **64**, 359–363.
 81. Clapper JR, Moreno-Sanz G, Russo R *et al.* (2010) Anandamide suppresses pain initiation through a peripheral endocannabinoid mechanism. *Nat Neurosci* **13**, 1265–1270.
 82. Li WW, Blankman JL & Cravatt BF (2007) A functional proteomic strategy to discover inhibitors for uncharacterized hydrolases. *J Am Chem Soc* **129**, 9594–9595.
 83. Favia AD, Habrant D, Scarpelli R *et al.* (2012) Identification and characterization of carprofen as a multi-target fatty acid amide hydrolase/cyclooxygenase inhibitor. *J Med Chem* **55**, 8807–8826.
 84. Lowin T, Zhu W, Dettmer-Wilde K *et al.* (2012) Cortisol-mediated adhesion of synovial fibroblasts is dependent on the degradation of anandamide and activation of the endocannabinoid system. *Arthritis Rheum* **64**, 3867–3876.
 85. Hamtiaux L, Masquelier J, Muccioli GG *et al.* (2012) The association of *N*-palmitoylethanolamine with the FAAH inhibitor URB597 impairs melanoma growth through a supra-additive action. *BMC Cancer* **12**, 1–14.
 86. Roques BP, Fournié-Zaluski M-C & Wurm M (2012) Inhibiting the breakdown of endogenous opioids and cannabinoids to alleviate pain. *Nat Rev Drug Discov* **11**, 292–310.
 87. Huggins JP, Smart TS, Langman S *et al.* (2012) An efficient randomised, placebo-controlled clinical trial with the irreversible fatty acid amide hydrolase-1 inhibitor PF-04457845, which modulates endocannabinoids but fails to induce effective analgesia in patients with pain due to osteoarthritis of the knee. *Pain* **153**, 1837–1846.



88. Li GL, Winter H, Arends R *et al.* (2011) Assessment of the pharmacology and tolerability of PF-04457845, an irreversible inhibitor of fatty acid amide hydrolase-1, in healthy subjects. *Br J Clin Pharmacol* **73**, 706–716.
89. Gamaledin I, Guranda M, Goldberg SR *et al.* (2011) The selective anandamide transport inhibitor VDM11 attenuates reinstatement of nicotine seeking behaviour, but does not affect nicotine intake. *Br J Pharmacol* **164**, 1652–1660.
90. Scherma M, Justinová Z, Zanettini C *et al.* (2012) The anandamide transport inhibitor AM404 reduces the rewarding effects of nicotine and nicotine-induced dopamine elevations in the nucleus accumbens shell in rats. *Br J Pharmacol* **165**, 2539–2548.
91. Reyes-Cabello C, Alen F, Gómez R *et al.* (2012) Effects of the anandamide uptake blocker AM404 on food intake depend on feeding status and route of administration. *Pharmacol Biochem Behav* **101**, 1–7.
92. Fu J, Bottegoni G, Sasso O *et al.* (2012) A catalytically silent FAAH-1 variant drives anandamide transport in neurons. *Nat Neurosci* **15**, 64–69.
93. Marsicano G & Chaouloff F (2012) Moving bliss: a new anandamide transporter. *Nat Neurosci* **15**, 5–6.
94. Berger WT, Ralph BP, Kaczocha M *et al.* (2012) Targeting fatty acid binding protein (FABP) anandamide transporters – a novel strategy for development of anti-inflammatory and anti-nociceptive drugs. *PLoS ONE* **7**, 1–12.
95. Pamplona FA, Ferreira J, Menezes-de-Lima O *et al.* (2012) Anti-inflammatory lipoxin A₄ is an endogenous allosteric enhancer of CB₁ cannabinoid receptor. *Proc Natl Acad Sci USA* **109**, 21134–21139.
96. Pertwee RG (2012) Lipoxin A₄ is an allosteric endocannabinoid that strengthens anandamide-induced CB₁ receptor activation. *Proc Natl Acad Sci USA* **109**, 20781–20782.
97. Bauer M, Chicca A, Tamborrini M *et al.* (2012) Identification and quantification of a new family of peptide endocannabinoids (pepcans) showing negative allosteric modulation at CB₁ receptors. *J Biol Chem* **287**, 36944–36967.

