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A Review - Biology, Aquaculture and Medical Use of Seahorse, *Hippocampus* spp

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Authors' contributions

This work was carried out in collaboration between all authors. Authors YYZ and ZJQ conceived and designed the study. Author YYZ wrote the first draft of the manuscript. Author BMR managed the literature searches. Authors BMR and ZJQ reviewed the first manuscript and wrote the final manuscript. All authors read and approved the final manuscript.

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Review Article

ABSTRACT

Seahorse has been used as medicine in Asian countries such as China, Korea, Japan and Vietnam for thousands of years. However, in western countries, the ecology of seahorse has been a focus of attention of many researchers for years. The seahorse aquaculture is popular with aqua farm worldwide for the sake of the increased demand of seahorse. This review described the biology, aquaculture and medical use of seahorse. To present the ecology and highlight the role of seahorse in traditional medicine and biomedical properties, this article based on the information collected from scientific literatures from Elsevier, Wiley, Science direct and CNKI.

Keywords: Hippocampus; biology; aquaculture; medical use.

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1. INTRODUCTION

Seahorse (genus *Hippocampus*), a marine teleost fish, is in the family *Syngnathidae*, which also includes pipefish, pipehorse and seadragon [1]. In the past, people were more interested in their specialized history traits, such as male pregnancy, monogamy, and so on. However, some studies indicated social polygamy still exist in some species [2]. In China, seahorse is one of the rare traditional medicines for its medical use to cure infertility, baldness, asthma and arthritis [3].

According to the CITES Trade Database, there were 70 countries or areas as sources and/or Consumers of seahorses and the annual volume of traded seahorses averaged 7 million individuals. Hong Kong SAR, the top consumer area of seahorses, imported an average of 57% of the total traded volume [4]. A number of fishers profited from catching seahorses for use in ornamental display, curios and traditional medicine [5]. Seahorse life-history characteristics (e.g. low population densities) may tend to make them overexploitation, therefore, all of them are included on CITES Appendix II [6]. In recent years, seahorse aquaculture has been regarded as an important strategy to alleviate the pressure on the wild stock of seahorses and one supplementary solution to substitute wild-caught seahorses to satisfy global demand [7].

This review extracts and integrates available information to provide detailed and comprehensive introduction on ecology and aquaculture of seahorses and medical use of seahorse powder.

2. BIOLOGY

2.1 Morphology

All seahorses have parallel morphology: a horselike head is perpendicular to its body; a pair of medium sized-eyes is rolled independently; a long snot has no teeth; trunk is stretched over a series of bony rings. Adult seahorses have no pelvic and caudal fins, and retain one propulsive dorsal fin, two small ear-like pectoral fins and a reduced anal fin [8,9].

2.2 Distribution and Species

After the convergence of the African and Eurasian plates, which was to blame for closure

of the Tethyan seaway, the world's tropical divided into biomass can be Atlantic Ocean biome (including the Caribbean and Mediterranean) and Indo-Pacific hiome Seahorses occupy both of the temperate and tropical shallow sea waters distributing from 50 degree north to 50 degree south. So far, there have been 32 species of seahorses recognized and 70% of them are existing in Indo-Pacific [10]. It is difficult to speculate the origin time and place for seahorse accurately due to the lack of reliable seahorse fossils, but some sacholars suggest that seahorse would stem from pre-Tethyan [11]. Teske [10] used sequence data derived from four markers (the nuclear RP1 and Aldolase and the mitochondrial 16S rRNA and cytochrome genes) to determine the phylogenetic b relationships among 32 species.

2.3 Habits

2.3.1 Inhabit and movement

Seahorses are usually found in coral reefs, mangroves, macroalgaes and seaweeds, but certain are accustomed to living in sandy estuary and lagoons [12] and they tend to twist their prehensile tails round the seaweeds when they are inactive. Dorsal fin, as a propeller, plays a pivotal role in movement while their pectoral fins function as rudder, both them facilitate seahorses to move. However, the tail is inoperative actually [9].

2.3.2 Feeding

Seahorses are visual ambush predators, using a technique known as the 'pivot' feeding, which involves rapid movement to overcome prey escape capabilities [13]. They don't have any teeth so they just suck the foods up through the snouts with "cluck cluck". Researchers investigated the diet of H. guttulatus and H. hippocampus in the Aegean Sea. What they found were that the diets of these seahorses were dominated by amphipoda, anomur. decapoda together with mysidacea, and females possibly had a higher feeding activity than males. Because they are susceptible to macrobenthic fauna, seahorses are prone to forage during daytime [14].

2.3.3 Reproduction

Pregnancy, a privilege for female, is defined as a period during which developing embryos are incubated in the body after egg–sperm union. But

when it comes to seahorse, pregnancy becomes different: the female injects her eggs into the male's pouch, a brooding structure, and then spermatids also enter the pouch. Such fertilization method can reduce sperm movement and shrink the lengthy zygotic migrate to the site of implantation. In fact, a zygote likes a sphere, which is beneficial to exchange of ion and gas around it. At the first few days, the pouch is small, while it will expand with the development of the embryos [15]. The male's pouch releases juveniles after 8-20 day's incubation, and juvenile seahorses are free-living and no further parental care is provided any more [16].

3. AQUACULTURE OF THE SEAHORSE

Seahorse once was regarded as a small-scale aquaculture for its low catches and economic value. A large number of seahorses were captured since 1980s as a result of excessive demand for Asian market, increased international trading volume and improvement of trawling technology, which led to the fact that all species of seahorse were listed on Appendix II of CITES or short for Convention for the International Trade in Endangered Species of Wild Fauna and Flora in November 2002 with implementation in May 2004 [17]. The efforts to culture seahorses commercially were not made until 1970s especially in southern China where the seahorses were first produced by captured-born in 1957 at Shantou. However, the farmers did not take a good knowledge of biology and breeding technology of seahorse. So these seahorse farms closed eventually [18]. Recently, an increasing number of experimental works have been conducted to investigate the impact of a series of parameters, including light, salinity, diet and temperature on survival, growth rate and reproduction. These efforts would provide theoretical supports and promote the development of commercial seahorse culturing.

3.1 Light

Light, an essential parameter in rearing seahorse, is exerting an influence on feeding, survival and growth rate, sexual maturity, spawning and endocrine hormone levels [19]. Wong [20] thought there were no significant difference among seahorses (*H. whitei* Bleeker, 1855) in various lights except Gonadosomatic index (GSI). Whereas, a study revealed that light could impact all indexes in seahorse growth markedly. Lin [21] cultured seahorses (H. erectus Perry, 1810) with different lights and he found that the juveniles cultured in 500, 1000 and 1500 lx had a high survival rates and low air-bubble disease rates. The final wet weight and standard length of the juvenile H. erectus were different at various lights. In addition, background and substrate color also have effect on seahorse skin coloration. For instance, seahorses (H. trimaculatus Leach, 1814) in blue background had the highest coloration rate compared to the others. When cultured with four different substrate colors (yellow, green, red and the mixture, respectively) for 5 days, 77.8% of the seahorses with the yellow substrate changed their skin color to yellow (P=0.007), and all seahorses at the red state turned into yellow or yellow-black [22]. Pawar [23] used tanks in green, blue, red, transparent, black and yellow to feed pelagic phase seahorses H. kuda (Bleeker 1852). Growth and survival rate were higher in the dark backgrounds than in the light; ingestion of the airbubble occurred more often in light backgrounds. Martinez [24] studied whether the tank color (transport, white, red, orange, yellow, green, blue and black) can cause difference among tested seahorses (H. abdominalis). The results indicated that all seahorses appeared to be able to survive and grow in every tanks. And the researchers needed further study to estimate the interactions of tank color and light intensity, retinal structure of the eye.

3.2 Salinity

Numerous studies reported that salinity has an effect on fish development and growth by regulating osmosensitivity which is related to survival rate, spawning and fertilizing, standard metabolic rate, food intake, food conversion efficiency and hormone stimulation, especially for some sensitive seahorses. Seahorse can live in a wide spectrum of salinity ranging from 8 to 35‰, and the most favorable salinity for juveniles is about 12‰ [25]. Cardenas [26] investigated the salinity tolerance and survival of seahorses (H. abdominalis) in different salinities through transferring seahorses from 32‰ to 5, 10, 15, 20, 25, and 32‰. As a consequence, seahorse in 5‰ sank in the tank bottom instead of swimming or feeding and the mortality would up to 100% within 72 h. The survival rates of seahorses cultured in 10 and 15‰ salinities were superior to those cultured at 25 and 32‰ salinities, which can be related to the fact that H. abdominalis is considered as pelagic during the first month of life. The optimum salinity for *H. reidi* was 10-25‰. All tested H. reidi were died in 0‰ after 10 h while those reared in 10-15‰ possessed the

maximum wet weight and survival rate [27]. Hilomen-Garcia [28] evaluated survival, growth, and total body water content after 4 and 18 days of 9-week-old H. kuda cultured in 0-85‰ transferred from salinity of 32-33‰. H. kuda could survive neither in 0% nor 55%. The upper limit of H. kuda salinity tolerance was 50%. Although 10‰ cultured seahorses had higher growth and survival compared with the control (30‰ seawater), total body water had a significant advance (10%). Therefore, like majority fish, H. kuda are accustomed to seawater salinities of 15 and 20‰. The growth of H. erectus is bound up to salinity. When the salinity was 31-33‰, H. erectus had the greatest wet weight and survival rate compared with others [21]. Salinity changes may have a significant effect on numbers of juveniles, which peaked at 10‰ while decreased at higher salinities, rather than mating behavior of *H. kuda*. In addition, the diverse salinities also induced changes of biochemical components in seahorses. For instance, the levels of SOD, CAT, MDA, AKP and ACP in H. kuda varied with salinity [29].

3.3 Diet

Diet difference may influence the juvenile seahorses. Therefore, it is a necessary that the diet should be changed with seahorse's growth. The new-born seahorses, having a small-scale snot with an average of 0.56 mm ranging from 0.45-0.68 mm, onlv can seize some microplanktons such as rotifer, microalgae and etc. while senior seahorses would be fond of Cladocera and Copepods or something larger. Secade [30] observed the same trend in fish survival, growth, protein, ash or humidity content, except for color changes when they bred the seahorses (H. hippocampus) with Artemia and frozen Mysis. According to Segade [30], Woods [31] thought there was no significant difference among tested seahorses cultured with Artemia sp. enriched with Algamac-3050 and frozen mysids Amblyops kempi. Enriched with chlorella could be conducive to seahorse's feeding and could expand the biodiversity of breeding grounds, mediate the water quality and optimize the survival circumstances [32]. Several studies showed that diet also had a pivotal effect on seahorse's reproduction. For instances, seahorses cultured with Artemia combined with tilapia or frozen mysids had a higher frequency of delivery and had more broods. Were they cultured with fresh-mysid rather than frozenmysid, seahorses (H. erectus) had a superior

nutritional status, which would improve their survival and growth [33].

Artemia density is a key factor to breed seahorses. A high density could cause improvement of corruption and organic matters, and aggravate the deterioration of water qualitylow pH and high ammonia nitrogen and nitrite nitrogen content, which were favorable for Vibrio growth instead of seahorses. However, low density could not supply sufficient nutrition for seahorses [34]. Therefore, a proper Artemia density could both maintain water quality and cut down the cost without impacting survival and growth. Yin [35] investigated the effect of various Artemia densities on juvenile seahorse (H. erectus). What he found was that 1-10 day seahorses should be cultured with Artemia with 10 individual/mL. After a month, the density of 10 individual/mL was appropriate.

3.4 Temperature

Temperature, a primary parameter during fish life, seems to be related to feeding, metabolism, immune function, the incubation rate and the gender of fishes [19], so do seahorses. Temperature mutations influenced the growth, biochemical composition and enzyme activity of seahorse (H. kuda). Sun [36] put the seahorses survived in 23°C to 15, 28 and 33°C respectively and he thought a proper mutation could facilitate the growth, biochemical composition and enzyme activity of seahorses. When the mutation was excessive, it led to poor health and death. Gonadosomatic index (GSI), fecundity and spawning, fertilization and hatching rate were all in the charge of temperature. Lin [20,37] demonstrated that the higher the temperature, the higher fecundity and spawning and fertilization rate, and the GSI was peaked at 28°C-the optimal temperature for reproduction. The sum of effective temperature was 14066.9°C h^{-1} . Furthermore, hatching behaviors of juvenile seahorses (5-10 day) were subject to temperature. At 18°C, seahorses sank into bottom without hatching or swimming, and they began to hatch with a low-rate at 22°C. But the maximum hatching rate reached at 26°C. A 107 day experiment conducted by Wong [20] showed that the growth rate of H. whitei were 0.14 mm/d and 0.26 mm/d at 17°C and 26°C, respectively. H. trimaculatus accommodated themselves to the water at 26°C [38]. Planas [39] described the survival and wet weight of H. guttulatus cultured in 15, 18 and 21°C. Those in 15°C died earlier than the other with a short swimming time. The

survival and wet weight were 21.1%, 85.9% and 81.1%, and 14.7 mg, 51.9 mg and 106.43 mg, eventually.

3.5 Disease

Seahorse aquaculture has encountered many bottlenecks including diseases. A study conducted by researchers from four countries showed that inflammatory processes caused by bacterial, parasitic and virus dominated the pathological progress. Neoplasms or neoplasticlike lesions, environment and congenital factors were the secondary cause [40]. Some researchers [41,42] compared the intestinal bacterial flora in health and intestinal-diseased seahorse intestinal and the microbiota differences between wild and cultured seahorses. What they found were that Vibrionaceae commonly represented the dominant population in the intestinal tract of all seahorses. In addition, Enterobacteriacea, Rhodobacteraceae, Sphingomonadaceae, Flavobacteriaceae, and Alcaligenaceae were also identified in seahorse intestinal tract. In another experiment, a new pathogenic bacteria or V. fortis which could cause enteritis in cultured seahorses [43]. V. infected parahaemolyticus isolated from seahorses was sensitive to Streptomyces Rifampicin, kanamycin, ciprofloxacin, norfloxacin, tetracycline, compound Daxin, bacteria must rule, Ned acid [44]. Declercq [45] thought that the white necrotic tail not only associated with Vibrio sp. and Mycobacterium sp., but also influenced by Uronema-like parasite.

4. MEDICAL USE

Seahorses have been used widely as a traditional medicine and invigorant from time immemorial, whose edible and medicinal effects have been seen in Compendium of Materia Medica (《本草纲目》) and Shennong Materia Medica(《神农本草》). Traditional medicinal uses and local names of some prescription authorized by CFDA are listed in Table 1. During the past decades, considerable research efforts have been directed to reveal their therapeutic potentials, such as anti-fatigue, anti-inflammatory, anti-tumoral activities [3].

4.1 Hormone-like Effect (Table 2)

A group of researchers [46,47] worked with Sun Yat-sen University investigated the Invigorating Yang and tonifying kidney function of Haima capsule (Qinyangchun, Wutongtang) bv intragastric administration to castrate mice. After 20 days, levels of load swimming time, serum testosterone and weight of the reproductive organs had improved. Another study feeding the kidney deficiency mouse with Haima capsule for 12 days found that capsule might possess androgenic hormone-like physiological function which enhanced the sexual response and mating ability and shortened the ejaculation latency time [48]. Meng [49] verified the Haima capsule could also treat benign prostatic hyperplasia (BPH). Similarly, Lu [50] employed Kidney Yangdeficiency mouse to illuminate the effect of Haima tonify Yang capsule, and they confirmed that this kind of capsule could alleviate impotence and enhance the mating ability.

Researchers contended that the acetic ether extract of *H. trimaculatus* had therapeutical effect on castration and testosterone-induced benign prostatic hyperplasia and oligospermatism explicated by decreasing serum ACP activity and reducing the expression of PCNA and bFGF in the prostate. Compared to Finasteride, a drug used to reduce symptoms of prostate enlargement, the extract did not induce finasteride-related adverse event [51].

Zhang [52] mixed the powder of *H. japonicas* with CMC-Na (0.5%) to prepare the suspension (0.59 g/ml). After the intragastric administration of the suspension to Hydrocortisone-treated mice, levels of peripheral serum hemolysin and cAMP were increased and the weight of prostate and testes were rising.

4.2 Antioxidant Activity (Table 3)

Wang [53] investigated the antioxidant activity of the water, methanol, n-butanol and ethyl acetate and chloroform extracts of seahorse (H. japonicus Kaup). The results showed that water extract possessed the highest antioxidant activity measured by DPPH radical scavenging activity with an equation of linear regression: Y=99.998-0.698X, $r^2=0.960$. The dry-seahorse was ground into powder and extracted with methanol within 30 min, then filtered it with ultrasonic. The authors studied antioxidant activity of seahorse by measuring the oxidation time required for the reduction of potassium permanganate, and the clearance rate of superoxide anion and the total and the clearance rate of superoxide anion and the total reducing power compared with ascorbic acid, which were 21.28 s, 31.98% and 0.48 of seahorse extract and 15.8 s, 42.42% and 0.686

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of ascorbic acid. Therefore, the researcher believed that the seahorse filtrate exhibited excellent antioxidant activity [54]. Using Sephadex G-10 gel to separate the water extract of H. trimaculatus could obtain 6 kinds of fractions containing various amounts of phenolics and amino acids among which FrIV had the highest antioxidant activity with IC_{50} value of 0.25 mg/mL. Chen [55] thought the antioxidant activity was related to types and contents of the amino acids, small molecular peptide and phenolic compounds derivated from the extract. In another study, the water extract of H. trimaculatus could reduce the peroxide value of oils, and it had the highest anti-oxidative activity when the extract gained in the optimum condition (pH=3.3, 40°C, 40 min) [56]. Qian [57] extracted H. kuda with water, methanol and ethanol and studied the free-radical and ROS scavenging activities of these different extracts which were measured with various antioxidant assay in vitro (e.g. reducing power, total antioxidant, DPPH radical scavenging, hydroxyl radical scavenging). Moreover, the inhibitory effects on intracellular ROS on RAW264.7 and myeloperoxidase (MPO) activity in HL60 cells were also evaluated. In 2012, two new phthalate derivatives from the methanol extract of *H. kuda* were isolated. Smilar to the former, both of them possessed antioxidant activities and had no cytotoxicity on RAW264.7 and MRC-5 cells [58].

Chen [59] employed the alkali protease to hydrolyze *H. trimaculatus* and studied the antioxidant activity of this product. As a result, the antioxidant activity of this protein hydrolysate was 4.73 times higher than that of ascorbic acid on the same condition. Papain enzyme,

Drug Name	Main Composition	Function
Haimasanshen	Hippocampus, Epimedium, Morinda	Invigorating Yang,
Pellet	officinalis How, Velvet antler, donkey kidney, etc.	Tonifying kidney
Haimawanying	Hippocampus, Saussurea costus,	Accelerating circulation of
Paste	notopterygium, Angelica dahurica,	blood, Acesodyne,
	Angelica sinensis, etc.	Bruise
Haimiaduobian	Hippocampus, gecko, cynomorium	Invigorating Yang,
Pellet	songaricum, Semen Cuscutae, etc.	Beneficial to the sperm
Haimabaji	Hippocampus, Morinda officinalis How,	Invigorating Yang,
Capsule	pilose antler, Cnidium monnieri, etc.	Tonifying kidney, Anti-
		fatigue, Impotence
Jingzhihaimazhuifen Paste	Hippocampus, Unprocessed,	Accelerating circulation of
	Achyranthes, parsnip, eucommia, cinnamon, saffron, etc.	blood, Acesodyne
Shexianghiamazhui	Hippocampus, Unprocessed,	Accelerating circulation of
feng Paste	Achyranthes, parsnip, eucommia, cinnamon, saffron, etc.	blood, Acesodyne
Shenronghaima	Hippocampus, dog kidney, pilose antler,	Invigorating Yang,
Pellet	ginseng, Epimedium, etc.	Tonifying kidney, Anti- fatigue
Haimaqiangshen Pellet	Hippocampus, Radix Rehmanniae	Spermatorrhea, Asynodia,
	Preparata, Fructus Corni, pilose antler, deer kidney, etc.	Tonifying kidney
Haimashuhuo	Hippocampus, panax notoginseng,	Bruise,
Pellet	daemonorops draco, red flower,	Reducing swelling,
	Ligusticum wallichii, etc.	Alleviating pain
Haimabushen	Hippocampus, rehmannia, donkey kidney,	Nourishing yin, Tonifying
Pellet	dog kidney, etc.	kidney, Strong brain
Haimaxiangcao	Hippocampus, Ginseng, Cistanche,	Invigorating Yang,
Wine	Epimedium, etc.	Tonifying kidney, Fatigue, Impotence

Table 1. Traditional medicine containing Hippocampus in China

Subject	Processing method	Experimental model	Reference
Haima capsule		Castration mouse	[46, 47]
Haima capsule		Kidney deficiency mouse	[48]
Haima capsule		Castration rats	[49]
Haima tonify Yang capsule		Kidney Yang-deficiency mouse	[50]
H. trimaculatus	Acetic ether extract	Castration rats	[51]
H. japonicus	Powder	Hydrocortisone-treated mice	[52]

Table 2. The hormone-like effect of seahorse

Subject	Processing method	Experimental model	Reference
H. japonicus	Ethanol extract	DPPH radical scavenging activity	[53]
Seahorse	Ethanol extract	Scavenging activity on superoxide anion	[54]
		Total antioxidant activity	
H. trimaculatus	Water extract	DPPH free radical scavenging Hydroxyl radical scavenging Total reducing power	[55]
		Ferrous ion chelating ability	
H. trimaculatus	Water extract	Antioxidative activities on fat and oil	[56]
H. kuda	Ethanol extract Water extract Methanol extract	Free radical scavenging activities Reactive oxygen species	[57]
H. kuda	Methanol extract		[58]
H. trimaculatus	Enzymatic hydrolysis	DPPH radical scavenging activity	[59]
H. kuda	Enzýmatic hýdrolýsis	DPPH free radical scavenging Hydroxyl radical scavenging Ferrous ion chelating ability	[60]

Table 3. The antioxidant effect of seahorse

selected out from 5 kinds of proteases, was used to hydrolyze seahorse protein. After the filtration and freeze drying, researchers [60] found 5 kinds of fractions of which component 3, 4 and 5 had excellent activity with IC_{50} of 0.896, 0.982, and 0.814 mg/mL respectively. All of the fractions could both significantly eliminate DPPH radical and hydroxyl radical and had iron-chelating ability. ACE inhibitory peptides obtained from the hydrolysate exhibits antioxidant as well as hypotensive function. Similarly, Liu [61] thought that the hydrolysate prepared by neutral protease could scavenge DPPH radical and superoxide anion radical.

4.3 Anti-Fatigue Activity (Table 4)

To explore the anti-fatigue activity of seahorse, neutral protease was availed to hydrate seahorse ponder. The researchers reported that the enzymatic extract of *H. kelloggi* had anti-fatigue activity evaluated by the activity of SOD and MAD, as well as the levels of blood lactic acid and serum urea, besides, the extracts could inhabit Escherichia coli, Bacillus gasoformans and Proteusbacillus vulgaris [61]. Suspending liquid of eight kinds of marine creatures-Syngnathus sp., H. trimaculatus, oyster, tortoise plastron, Oplopanax elatus Nakai, Asteria, Spirulina and seatangle, had different anti-fatigue activity on tested mice after 14 days [62].

Zhu [63] reported that ethanolic extract of *H. trimaculatus* Leach played a significant part in anti-fatigue and enhancing hypoxia tolerance. In another experiment, the result showed that the enzymatic hydrolysis of *H. kuda* prolonged the swimming time, cold resistance and hypoxia tolerance time, as well as decreased serum lactic acid and serum urea nitrogen content after swimming [64].

4.4 Anti-inflammatory, Anti-tumoral and Immune-Modulating Activities (Table 5)

Chen [65] proved that the petroleum ether extract of seahorse was composed of phospholipids, unsaturated fatty acid and phenolic compounds,

Subject	Processing method	Experimental model	Reference
H. kelloggi	Enzymatic hydrolysis	Mice forced swimming test	[61]
H. trimaculatus	Water extract	Mice forced swimming test	[62]
H. kuda	Ethanol extract	Mice forced swimming test	[63]
H. kelloggi	Enzymatic hydrolysis	Mice forced swimming test	[64]

Table 4. Anti-fatigue activity of seahorse

Table 5. Anti-inflammatory, anti-tumoral and immune-modulating activities of se	eahorse
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Subject	Processing method	Experimental model	Reference
H. kuda	Petroleum ether extract	RAW264.7 cell	[65]
H. kuda	Enzymatic hydrolysis	MG-63 and SW-1353 cell	[67]
H. erectus	Powder	⁶⁰ Co-y treated mice	[68]
H. kuda	Ethanol extract	Intragastic administration	[63]

which suppressed the production of NO in LPSinduced RAW264.7 cell. To manifest the antineuro inflammatory function of paeonol or 1-(2hydroxy-4-methoxyphenyl) ethanone, isolated from H. kuda Bleeler, Himaya [66] studied the inflammatory response in LPS-induced BV-2 microglial and RAW264.7 macrophage cells, and they found that the paenonl could inhibit the proinflammatory cytokines in both activated BV-2 and RAW264.7 cells in a dose-dependent manner. Moreover, MAPK and NF-KB signal pathway were inhibited as well. Ryu [67] optimized the enzymatic hydrolysis of H. kuda protein and investigated the anti-inflammatory activity of the peptide DPFDKDDWDNWK (1821 Da) purified from the hydrolysis. As expected, this peptide promoted the formation of collagen and mineralization by reducing the expression of MMP-13, iNOS and COX-2 in TPA-induced MG-63 and SW-1353 cells.

The effect of suspend of *H. erectus* powder was studied by intragastic administration to ⁶⁰Co- γ treated mice. Compared with control group (⁶⁰Co- γ treated only), the levels of peripheral white blood cells and platelets were higher for 3 days and were significantly higher for 7 days. When it lasted for 30 days, the survival rates of experimental groups (2 groups) and control group were 33.3% (20 mg/kg), 53.3% (40 mg/kg) and 6.7% respectively [68]. Zhu [63] concluded that ethanolic extract of seahorse possessed immune-modulating and anti-allergy properties by enhancing the phagocytosis of macrophage and reducing the swelling degree caused by 2, 4-Dinitrochlorobenzene.

4.5 Other Activities

Xu [69] used methanol to extract the supernatant of the raw powder (*H. kuda*) twice and then obtained the jasmine crystal followed by lyophilization of the extract. They measured the inhibitory rate of the extract on carotid artery thrombosis and cerebral thrombosis with Aspirin as a positive control group, and the results showed that rate were 24.2%, 40.5%, and 46.8% at does of 50 mg/kg, 100 mg/kg and 200 mg/kg while the Aspirin was 44.7% at 10 mg/kg. It was unsaturated fatty acids that formed the main constitutes of the extract, which could reduce blood viscosity and raise blood flow velocity.

In traditional Chinese medicine, seahorse is used for tranquilizer and sedative. To explore this medicinal effect, Zhang [70] determined them by the calcium mobilization assay. The result showed that the water and ethanol extract of the *H. kelloggi, H. trimaculatus. H. japonicas,* and *H. kuda* could significantly inhabit the calcium influx in rat neuron, which underlied this medical mechanism.

The middle-cerebral-artery-occlusion-induced focal ischemia/reperfusion (I/R) rat brain damage model similar to human ischemic stroke was used to study the effects of seahorse extraction on neurological deficit score (NDS), cerebral infarction volume and cerebral water content after I/R. In briefly, the seahorse powder was suspended into 0.3% CMC-Na solution. The suspension was given by intra gastric administration to mice at a dose of 10 ml/kg an hour before I/R. The results showed that the extract lowered the NDS significantly and reduced cerebral infarct volume and brain edema reaction at the interval of 1 h, 24 h and 72 h after I/R [71].

5. CONCLUSION

As described above, seahorses have been shown to played diverse roles in tradition medicine and pharmacologic properties. The majority of the trade was for TCM and the demand continued to rise. Although the seahorse aquaculture has been developed for several years combined with updated technical, there are still many difficulties in this industry. Therefore, it is imperative for us to encourage technical advance to alleviate the pressure on wild population.

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. Kleiber DL, Blight LK, Caldwell IR, Vincent, ACJ. The importance of seahorses and pipefishes in the diet of marine animals. Rev Fish Biol Fish. 2011;21:205-223.
- Woodall LC, Koldewey HJ, Shaw PW. Serial monogamy in the European longsnouted seahorse *Hippocampus guttulatus*. Conserv. Genet. 2011;12:1645-1649.
- Kumaravel K, Ravichandran S, Balasubramanian T, Sonneschein L. Seahorses-A source of traditional medicine. Nat Prod Res. 2012;26:2330-2334.
- Evanson M, Foster S, Wiswedel S, Vincent A. Tracking the international trade of Seahorses (*Hippocampus* Species). Fisheries Centre Research Reports. 2011; 19:1-92.
- Vincent ACJ, Foster SJ, Koldewey HJ. Conservation and management of seahorses and other Syngnathidae. J Fish Biol. 2011;78:1681-1724.
- Foster S, Wiswedel S, Vincent A. Opportunities and challenges for analysis of wildlife trade using CITES dataseahorses as a case study. Aquat Conserv Mar Freshw Ecosyst. 2016;26:154-172.
- 7. Koldewey HJ, Martin-Smith KM. A global review of sea horse aquaculture. Aquaculture. 2010;302:131-152.
- 8. Foster SJ, Vincent ACJ. Life history and ecology of seahorses: Implications for

conservation and management. J Fish Biol. 2004;65:1-61.

- Lv JY, Sun YY, Wang XH, Yang DW, Li BJ, Chen L. The Biological Characteristics and Ecological Habits of *Hippocampus* spp. Fisheries science & Technology. 2001;3: 32-33.
- Teske PR, Cherry MI, Matthee CA. The evolutionary history of seahorses (*Syngnathidae*: *Hippocampus*): Molecular data suggest a West Pacific origin and two invasions of the Atlantic Ocean. Mol Phylogenet Evol. 2004;30:273–286.
- Casey SP, Hall HJ, Stanley HF, Vincent ACJ. The origin and evolution of seahorses (Genus *Hippocampus*): A phylogenetic study using the cytochrome b gene of mitochondrial DNA. Mol Phylogenet. Evol. 2004;30:261-272.
- 12. Lourie SA, Foster SJ, Cooper EWT, Vicent ACJ. A Guide to the identification of seahorses. College Park; 2004.
- Gemmell BJ, Sheng J, Buskey EJ. Morphology of seahorse head hydrodynamically aids in capture of evasive prey. Nat. Commun. 2013;4:2840.
- Kitsos MS, Tzomos T, Anagnostopoulou L, Koukouras A. Diet composition of the seahorses, *Hippocampus guttulatus* Cuvier 1829 and *Hippocampus hippocampus* (L. 1758) (*Teleostei, Syngnathidae*) in the Aegean Sea. J Fish Biol. 2008;2:1259-1267.
- 15. The reproduction habits of hippocampus. China Fisheries News. 2013;3:1.
- 16. Stölting KN, Wilson AB. Male pregnancy in seahorses and pipefish: Beyond the mammalian model. Bioassays. 2007;29: 884-896.
- 17. Lam JTL, Koldewey HJ, Yasué M, Vincent ACJ. Comparing interview and trade data in assessing changes in the seahorse *Hippocampus* spp. trade following CITES listing. Oryx. 2014;50:1-11.
- Xi YF, Yin F. Advances in artificial culture of *Hippocampus*. Modern Fisheries Information. 2011;26:9-15.
- 19. Sun DW, Zhan Y, Xu ZR. The important role of temperature in fishery culture. Chinese Feed. 2003;4:20-21.
- 20. Wong JM, Benzie JAH. The effects of temperature, Artemia enrichment, stocking density and light on the growth of juvenile seahorses, *Hippocampus whitei* (Bleeker, 1855), from Australia. Aquaculture. 2003; 228:107-121.

- 21. Lin Q, Lin J, Huang L. Effects of light intensity, stocking density and temperature on the air-bubble disease, survivorship and growth of early juvenile seahorse *Hippocampus erectus* Perry, 1810. Aquac Res. 2010;42:91-98.
- Qin G, Lin Q, Gu N, Lin J, Huang L. Effect of broodstock origin, background and substrate color on skin coloration of threespotted seahorses *Hippocampus trimaculatus* Leach, 1814. J Exp Mar Biol Ecol. 2012;15:129-134.
- Pawar HB, Sanaye SV, Murugan A, Sreepada RA. Effect of background color of tanks on growth and survival of Juvenile Yellow seahorse, *Hippocampus kuda* (Bleeker 1852), in the Pelagic Phase. Isr J Aquacult-Bamid. 2011;621:1-6.
- 24. Martinez-Cardenas L, Purser GJ. Effect of tank color on Artemia ingestion, growth and survival in cultured early juvenile potbellied seahorses (*Hippocampus abdominalis*). Aquaculture. 2007;264:92-100.
- 25. Zhu LH. The key technical points of *Hippocampus* breeding. Hebei Fisheries. 2005;5:42.
- Martinez-Cardenas L, Purser JG. Effect of Direct transfer to different salinities on early juvenile pot-bellied seahorse, *Hippocampus abdominalis*. Survival in Culture Conditions. J World Aquacult Soc. 2016;46:201-206.
- 27. Hora MDSCD, Joyeux JC, Rodrigues RV, Sousa-Santos LPD, Gomes LC, Tsuzuki MY. Tolerance and growth of the longsnout seahorse *Hippocampus reidi*, at different salinities. Aquaculture. 2016;463:1-6.
- Hilomen-Garcia GV, Delos RR, Garcia CMH. Tolerance of seahorse *Hippocampus kuda* (Bleeker) juveniles to various salinities. J Appl Chthyol. 2003;19:94-98.
- 29. Xu YJ, Sun B. Effects of salinity stress on the growth, components and enzyme activity of juvenile *Hippocampus kuda*. Oceanologia et Limnologia Sinica. 2016;6: 1279-1285.
- Segade á, Robaina L, Otero-Ferrer F, García Romero J, Molina DL. Effects of the diet on seahorse (*Hippocampus hippocampus*) growth, body colour and biochemical composition. Aquac. Nutr. 2015;21:807-813.
- 31. Woods CMC, Valentino F. Frozen mysids as an alternative to live Artemia in culturing

seahorses *Hippocampus abdominalis*. Aquac Res. 2003;34:757-763.

- 32. Wei XD, Ye CM, Chen DH. Effects of diets on growth of juvenile seahorses. Acta Scientiarum Naturalium Universitatis Sunyatseni. 2003;42:60-63.
- Wang HX, Liu XD, Zhang D, Zhang GY, Shi JH. Effects of frozen mysis and fresh mysis on the reproduction of *Hippocampus erectus*. Fisheries Science & Technology Information. 2012;39:6-9.
- Lv JY. Effects of animal diet on bacteria and physicochemical factors in aquaculture water. Acta Scientiarum Naturalium Universitatis Sunyatseni. 2007;46:65-65.
- 35. Yin F, Tang BJ, Zhang D, Wang JG. Growth and Survival of Juvenile Lined Seahorse (*Hippocampus erectus*) at Different Food Densities. Chin J Appl Environ Biol. 2012;18:617-622.
- 36. Sun B, Chen S, Xu YJ, Dai GP. Effects of temperature mutation on the growth, components and enzyme activity of *Hippocampus kuda*. Oceanologia et Limnologia Sinica. 2012;42:67-72.
- Lin Q, Gao Y, Sheng J, Chen Q, Zhang B, Lu J. The effects of food and the sum of effective temperature on the embryonic development of the seahorse, *Hippocampus kuda* Bleeker. Aquaculture. 2007;262:481-492.
- Sheng J, Lin Q, Chen Q, Gao Y, Chen L, Lu J. Effects of food, temperature and light intensity on the feeding behavior of threespot juvenile seahorses, *Hippocampus trimaculatus* Leach. Aquaculture. 2006; 256:596-607.
- Planas M, Blanco A, Chamorro A, Valladares S, Pintado J. Temperatureinduced changes of growth and survival in the early development of the seahorse *Hippocampus guttulatus*. J. Exp Mar Biol. Ecol. 2012;438:154-162.
- 40. LePage V, Young J, Dutton CJ, Crawshaw G, Paré JA, Kummrow M, McLelland DJ, Huber P, Young K, Russell S, Al-Hussinee L, Lumsden JS. Diseases of captive yellow seahorse *Hippocampus kuda* Bleeker, potbellied seahorse *Hippocampus abdominalis* Lesson and weedy seadragon Phyllopteryx taeniolatus (Lacépède). J Fish Dis. 2015;38:439-450.
- 41. Li F, Wang K, Luo W, Huang L, Lin Q. Comparison of the intestinal bacterial flora in healthy and intestinal-diseased seahorses *Hippocampus trimaculatus*, *Hippocampus erectus*, and *Hippocampus*

spinosissimus. J World Aquac. Soc. 2015; 46:263-272.

- Wang K, Zhang Y, Li F, Lin Q. Comparison of intestinal microbiota in intestinal tracts of the wild and cultured seahorses, *Hippocampus trimaculatus* (Leach): Intestinal bacteria of seahorse. J. World Aquac. Soc. 2016;47:123-128.
- 43. Wang X, Zhang Y, Qin G, Luo W, Lin Q. A novel pathogenic bacteria (*Vibrio fortis*) causing enteritis in cultured seahorses, *Hippocampus erectus* Perry, 1810. J. Fish Dis. 2016;39:765-769.
- 44. Yang HY, Yang WD, Heng G, Zhao W, Zhang GH. Study on isolation, identification and sensitivity to antibioties of seahorse *H. Kelloggi*. Chinese Frontier Health Quarantine. 2006;29:232-238.
- Declercq AM, Chiers K, Van DBW, Rekecki A, Teerlinck S, Adriaens D, Haesebrouck F, Decostere A. White necrotic tail tips in estuary seahorses, *Hippocampus kuda*, Bleeker. J. Fish Dis. 2014;37:501-504.
- Xu DH, Mei XT, Li BJ, Lin ZL, Xu SB. Pharmacological Effects of *Hippocampus* capsule on improving sexual function of rats. Journal of Chinese Medicinal Materials. 2003;26:807-808.
- 47. Mei XT, Xu DH, Lin ZL, Xu SB, Li BJ. The strengthening effect of hippocampal capsule on kidney deficiency rats. Chinese Traditional and Herbal Drugs. 2005;36: 409-140.
- Xu DH, Mei XT, Li BJ, Lin ZL, Xu SB. Pharmacological Effects of *Hippocampus* capsule on improving sexual function of rats. Journal of Chinese Medicinal Materials. 2003;26:807-808.
- 49. Meng XQ, Xu DH, Mei XT, Li BJ, Lv JY. Study on hippocampal capsule in the treatment of experimental benign prostatic hyperplasia. Chinese Pharmacological Journal. 2005;40:190-193.
- 50. Lu Y, Chen WX, Hua YQ, Meng DC. A study on hippocampus impotence yang capsule experimental pharmacological. Journal of Nangjing TCM university. 2001; 7:99-101.
- Xu DH, Wang LH, Mei XT, Li BJ, Lv JL, Xu SB. Protective effects of seahorse extracts in a rat castration and testosteroneinduced benign prostatic hyperplasia model and mouse oligospermatism model. Environ. Toxicol. Pharmacol. 2014;37:679-688.
- 52. Zhang H, Luo SD, Zhou BH, Yin WH. The Invigorating Yang and tonifying kidney

effect of *Hippocampus japonicas*. Chinese Journal of Marine Drugs. 1997;46:53-56.

- Wang H, Chen JJ, Wang LL, Zhang DL, Wang XR, Lee SC, Yang DF. Evaluation of DPPH free radical scavenging capability of *Hippocampus japonicus kuda* and their HPLC Fingerprint. Chin Pharm J. 2000;44: 1121-1125.
- 54. Lin FH, Zheng AE, Li YQ, Zhu H. Comparison of eight kinds of marine drugs *in vitro* antioxidant activity such as *Hippocampus*. Journal of Anhui Agr Sc. 2015;43:73-74.
- 55. Chen LP, Shen XR, Chen GH, Sun FF, Jin MN. Antioxidant activity of aqueous extract from Three-spot *Hippocampus*. Science and Technology of Food Industry. 2016;3: 114-116.
- 56. Yi MH, Li GQ, Xiao H, Lin ZS, Zeng DZ. Study on the antioxidative activities of extracts of *Hippocampus trimaculatus* leach and *Syngnathus* on fat and oil. China Tropical Medicine. 2006;6:1784-1785.
- 57. Qian ZJ, Ryu B, Kim MM, Kim SK. Free radical and reactive oxygen species scavenging activities of the extracts from seahorse, *Hippocampus kuda* Bleeler. Biotechnol. Bioprocess Eng. 2008;13:705-715.
- 58. Qian ZJ, Kang KH, Kim SK. Isolation and antioxidant activity evaluation of two new phthalate derivatives from seahorse, *Hippocampus Kuda* Bleeler. Biotechnol. Bioprocess Eng. 2012;17:1031-1040.
- Chen CB, Yuan XH, Chen Z, Yi MH. Study on extraction and antioxidant activities of liquefied protein of three-spot *Hippocampus*. China Tropical Medical. 2011;11:329-331.
- 60. Gu W, Xu YJ. Preparation of *Hippocampus* ACE inhibitory peptide and determination of antioxidant capacity. Science and Technology of Food Industry. 2016;5:201-211.
- 61. Liu Q, Dong ML, Zhang H, Kong L, Li W. Study on the activity of marine drughippocampal hydrolysate. Hebei Fisheries. 2015;9:10-15.
- 62. Hu JY, Li BF, Li ZJ, Lv P. A study on the anti-fatigue effect of eight marine drugs. Chinese Journal of Marine Drugs. 2000;2: 56-58.
- 63. Zhu AM. Pharmacologic Researches on ethanol extracts from *Hippocampus*. Chinese Pharmaceutical Affair. 2005;1:23-24.

- Peng WD, Chen QL. Anti-fatigue effect of enzymatic extract of *Hippocampus*. Chinese Journal of Food Hygiene. 2005;5: 404-407.
- 65. Chen LP, Shen XR, Chen GH, Cao XY, Yang J. A comparative study of the effects upon LPS induced macrophage RAW264.7 inflammation in vitro of the lipids of *Hippocampus trimaculatus* Leach. J Oleo Sci. 2015;64:1273-1281.
- Himaya SWA, Ryu B, Qian ZJ, Kim SK. Paeonol from *Hippocampus kuda* Bleeler suppressed the neuro-inflammatory responses *in vitro* via NF-κB and MAPK signaling pathways. Toxicol. *In Vitro*. 2012; 26:878-887.
- Ryu B, Qian ZJ, Kim SK. Purification of a peptide from seahorse, that inhibits TPAinduced MMP, iNOS and COX-2 expression through MAPK and NF-κB activation, and induces human osteoblastic

and chondrocytic differentiation. Chem. Biol. Interact. 2010;184:413-422.

- Ni JQ, Zhang ZH, Li WQ. Protective Effect of *Hippocampus* on ⁶⁰Co-γ induced Mice. China Journal of Chinese Materia Medica. 1997;12:750-751.
- Xu DH, Xu SB. Anti-thrombosis effect of extract of *Hippocampus kelloggi* on rats and its component analysis. Chinese Journal of Marine Drugs. 1997;1:11-13.
- Zhang ZH, Xu GJ, Xu L, Wang Q. Effects of five kinds of hippocampal extracts on Lglutamate-induced Antagonistic effect of calcium influx in neurons. Chinese Journal of Marine Drugs. 1994;4:6-9.
- 71. Feng X, Wu ZF, Yang L, Zhou Y, Wang Q, Liu YZ, Yang HZ. Effects of Haima extraction on experimental ischemia/ reperfusion brain damage. J of Hunan Normal Univ (Med Sci). 2005;2:1-4.

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