



REVIEW

α -Synuclein Radiotracer Development and *In Vivo* Imaging: Recent Advancements and New Perspectives

Obada M. Alzghool, MSc,^{1,2*}  Guus van Dongen, PhD,¹ Elsmarieke van de Giessen, PhD,¹ 
Linda Schoonmade, MSc,³ and Wissam Beaino, PhD¹

¹Department of Radiology and Nuclear Medicine, Tracer Center Amsterdam, Amsterdam UMC, Vrije Universiteit, Amsterdam, The Netherlands

²Turku PET Centre, University of Turku, Turku, Finland

³Medical Library, Vrije Universiteit Amsterdam, Amsterdam, The Netherlands

ABSTRACT: α -Synucleinopathies including idiopathic Parkinson's disease, dementia with Lewy bodies and multiple systems atrophy share overlapping symptoms and pathological hallmarks. Selective neurodegeneration and Lewy pathology are the main hallmarks of α -synucleinopathies. Currently, there is no imaging biomarker suitable for a definitive early diagnosis of α -synucleinopathies. Although dopaminergic deficits detected with single-photon emission computed tomography (SPECT) and positron emission tomography (PET) radiotracers can support clinical diagnosis by confirming the presence of dopaminergic neurodegeneration, dopaminergic imaging cannot visualize the preceding disease process, nor distinguish α -synucleinopathies from tauopathies with dopaminergic neurodegeneration, especially at early symptomatic disease stage when clinical presentation is often overlapping. Aggregated α -synuclein (α Syn) could be a suitable imaging biomarker in α -synucleinopathies, because α Syn aggregation and therefore, Lewy pathology is evidently an early driver of α -synucleinopathies pathogenesis. Additionally, several antibodies and small molecule compounds targeting

aggregated α Syn are in development for therapy. However, there is no way to directly measure if or how much they lower the levels of aggregated α Syn in the brain. There is clearly a paramount diagnostic and therapeutic unmet medical need. To date, aggregated α Syn and Lewy pathology inclusion bodies cannot be assessed ante-mortem with SPECT or PET imaging because of the suboptimal binding characteristics and/or physicochemical properties of current radiotracers. The aim of this narrative review is to highlight the suitability of aggregated α Syn as an imaging biomarker in α -synucleinopathies, the current limitations with and lessons learned from α Syn radiotracer development, and finally to propose antibody-based ligands for imaging α Syn aggregates as a complementary tool rather than an alternative to small molecule ligands. © 2022 The Authors. *Movement Disorders* published by Wiley Periodicals LLC on behalf of International Parkinson Movement Disorder Society.

Key Words: α -synuclein; α -synucleinopathies; Parkinson's disease; positron emission tomography; radiotracers

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

*Correspondence to: Msc. Obada M. Alzghool, Turku PET Centre, University of Turku, Tykistökatu 6 A, 4th floor, 20520 Turku, Finland; E-mail: obalzg@utu.fi

Relevant conflicts of interest/financial disclosures: None of the authors has financial disclosures related to this review manuscript. All the authors report no conflicts of interest related to this review manuscript.

Funding agencies: This project has received funding from the European Union's Horizon 2020 research and innovation program under the Marie Skłodowska-Curie grant agreement no. 813528.

Received: 23 August 2021; **Revised:** 14 February 2022; **Accepted:** 21 February 2022

Published online 15 March 2022 in Wiley Online Library ([wileyonlinelibrary.com](https://www.wileyonlinelibrary.com)). DOI: 10.1002/mds.28984

α -Synucleinopathies form a subset of neurological disorders that include idiopathic Parkinson's disease (PD), dementia with Lewy bodies (DLB), multiple systems atrophy (MSA), and some rare disorders, such as pure autonomic failure.¹ These disorders share unclear etiology, overlapping symptoms and pathological hallmarks.²⁻⁸ Neurological disorders are the leading cause of disability in the world, the fastest growing of which is PD surpassing even Alzheimer's disease (AD).⁹ PD is also the most common type of α -synucleinopathies. The Global Burden of Disease Study in 2016 estimated that 6.1 million individuals had PD and that number is rising exponentially.¹⁰ Aging is the greatest risk factor for PD and other α -synucleinopathies. Genetic factors also

affect disease risk, onset, and progression. There are variants in more than 20 genes reported to cause PD.¹¹ Several missense mutations in *SNCA*, the α-synuclein (αSyn) encoding gene, are proven to cause parkinsonism.¹²⁻¹⁸ Gene multiplication is another abnormality associated with *SNCA* where the extra gene copies may cause an increased expression of αSyn and an increased tendency to self-aggregation and malfunction.¹⁹⁻²¹

αSyn is a ubiquitous protein in the central nervous system accounting for up to 1% of the total cytosol proteins.^{22,23} αSyn is predominantly expressed in the brain, and concentrated at the pre-synaptic nerve terminals.²⁴ The precise function of αSyn at physiological conditions remains unclear; however studies using cellular and animal models show that αSyn contributes to synaptic vesicles trafficking and neurotransmitter release.²⁵ In vitro aggregation studies revealed that wild type and mutated variants of recombinant human αSyn monomers aggregate to form oligomers, which further aggregate into fibrils, a process associated with structural transition from random coil to β-sheet.²⁶

The main hallmarks of α-synucleinopathies are selective dopaminergic neurodegeneration and Lewy pathology being the process of αSyn aggregation into inclusion bodies.²⁷⁻²⁹ Lewy pathology plays a central role in the pathogenesis of α-synucleinopathies as one of the main drivers of neurodegeneration. This happens via the ongoing process of αSyn aggregation, which disrupts cellular functions, and induces mitochondrial damage and synaptic dysfunction.³⁰ Lewy pathology develops in different anatomic patterns and cell types in different α-synucleinopathies. Neuropathological studies show that in PD and DLB αSyn accumulates inside the neurons as Lewy bodies (LBs) and Lewy neurites (LNs),^{31,32} whereas in MSA, αSyn accumulates inside oligodendrocytes as glial cytoplasmic inclusions (GCIs) and intraneuronal inclusions (Fig. 1A).^{33,34} In vitro and in vivo studies using primary neurons, rodents and patient brain tissue have demonstrated that GCI αSyn and LB αSyn strains are formed based on the different intracellular environments in oligodendrocytes and neurons, respectively, and that these strains possess distinct structural and biological properties. GCI αSyn aggregates are more structurally compact compared to LB αSyn. In addition, GCI αSyn demonstrate higher potency in inducing Lewy pathology and different pattern of transmission.^{35,36}

Lewy pathology spreading in PD brain tends to follow a consistent pattern. LBs and LNs first appear in the medulla oblongata and olfactory bulbs, then cover the pontine tegmentum, midbrain, limbic brain regions, and eventually extend into the neocortex.³⁷ In DLB, Lewy pathology is widely spread and depending on the distribution, pathology is classified as brainstem, limbic, or neocortical-predominant.³⁸ It should be noted that DLB and PD are increasingly considered a disease

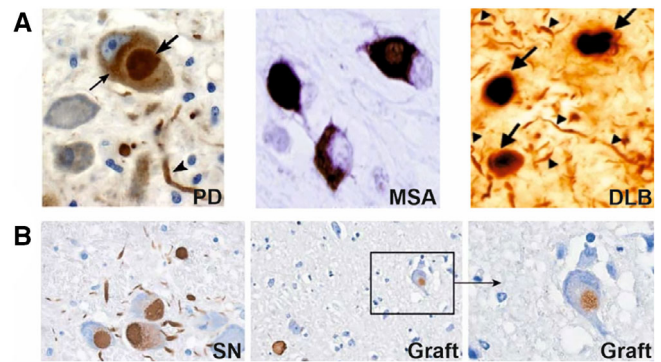


FIG. 1. Lewy pathology in different α-synucleinopathies shown by αSyn immunohistochemical staining in Parkinson's disease (PD) and multiple system atrophy (MSA) using paraffin-embedded brain tissue sections, and in dementia with Lewy bodies (DLB) using floating brain tissue sections. (A) Representative examples of Lewy bodies (bold arrow), Lewy neuritis (arrowhead) and αSyn aggregates (light arrow) in PD and DLB and glial cytoplasmic inclusions in MSA.^{32,34} (B) Lewy bodies and Lewy neurites in neurons of the substantia nigra (left) propagate to grafted neurons (middle and right) in PD.¹⁰⁰

continuum based on similar pathology, although in DLB and the intermediate form Parkinson's disease dementia (PDD), dementia is at the foreground of the clinical presentation.³⁹ Lewy pathology in MSA spreads from the basal ganglia, brainstem, and cortex into the cervical spinal cord and thalamus, then to the hippocampus and amygdala, and eventually into the occipital neocortex.⁴⁰ The unified staging system for Lewy body disorders⁴¹ and the staging/typing of Lewy body-related α-synuclein pathology⁴² are clinically used assessment protocols of Lewy pathology progression in different α-synucleinopathies.

Diagnosis and Therapy of α-Synucleinopathies

Diagnosis of α-synucleinopathies is performed according to clinical criteria. For example, clinical diagnosis of PD is based on the presence of bradykinesia in combination with at least resting tremor or rigidity and at least two supportive criteria (eg, beneficial response to dopaminergic therapy) without exclusion criteria (eg, cerebellar abnormalities or normal presynaptic dopaminergic imaging).⁴³ DLB is clinically diagnosed as progressive dementia accompanied with fluctuating cognition, visual hallucinations, parkinsonism, and REM sleep behavior disorder.⁴⁴ MSA is clinically diagnosed as autonomic dysfunction and either poorly levodopa-responsive parkinsonism or a cerebellar syndrome.⁴⁵

Single-photon emission computed tomography (SPECT) and positron emission tomography (PET) imaging could support the clinical diagnosis of α-synucleinopathies. Pre-synaptic dopaminergic imaging with

[¹²³I]FP-CIT SPECT and [¹⁸F]FDOPA PET is used to distinguish neurodegenerative parkinsonisms (PD, MSA, progressive supranuclear palsy [PSP] and corticobasal degeneration [CBD]) from non-neurodegenerative parkinsonisms (eg, essential tremor, vascular or medication induced parkinsonism) with high sensitivity (98%) and specificity (98%).⁴⁶ Pre-synaptic dopaminergic imaging is also clinically useful to distinguish DLB from AD (Fig. 2).^{47,48} It can even be used to assess dopaminergic neurodegeneration in prodromal symptoms such as REM sleep behavior disorder⁴⁹ and hyposmia.⁵⁰ Although dopaminergic deficits detected with SPECT and PET radiotracers can support clinical diagnosis by confirming the presence of dopaminergic neurodegeneration, dopaminergic imaging cannot visualize the preceding disease process. Approximately 50% of nigral dopaminergic neurons are already degenerated at motor symptom manifestation and current clinical diagnosis.⁵¹ Diagnosing α -synucleinopathies is ideally performed at an earlier stage when neurodegeneration is still limited. This would provide a window of opportunity for therapeutic intervention to prevent, slow down, or halt the disease progression. Another clinical challenge is the differential diagnosis of atypical parkinsonisms, especially at early symptomatic disease stage when clinical presentation is often overlapping. Presynaptic dopaminergic imaging cannot reliably distinguish PD from the atypical parkinsonisms MSA, PSP, and CBD. [¹⁸F]FDG brain PET imaging can be used to aid the differential diagnosis of atypical parkinsonisms, but

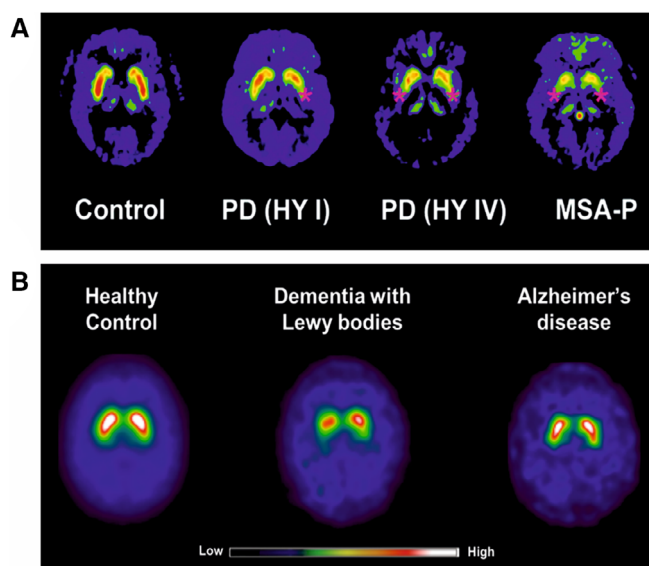


FIG. 2. Representative examples of dopamine transporter (DAT) imaging in α -synucleinopathies. (A) [¹⁸F]FDOPA uptake is reduced (red stars) in the putamen in Parkinson's disease early (PD HY I) and advanced (PD HY IV) stages, and multiple system atrophy with parkinsonian symptoms (MSA-P) compared to control.¹⁰⁴ (B) [¹²³I]FP-CIT show reduced DAT uptake in dementia with Lewy bodies, whereas DAT uptake is normal in healthy controls and Alzheimer's disease.⁴⁸

its reading requires training and semi-quantitative analysis software, and its value in the pre-symptomatic phase is probably limited.⁵² A good imaging biomarker to distinguish α -synucleinopathies (PD, MSA) from non- α -synucleinopathies (in particular the tauopathies PSP and CBD) could be clinically useful.⁵³

Currently, there is no reliable imaging biomarker for an early and definitive ante-mortem diagnosis of α -synucleinopathies. Lewy pathology is an early driver of α -synucleinopathies pathogenesis. There is evidence for an association between Lewy pathology and clinical symptoms severity,^{41,54,55} neuronal dysfunction,^{56,57} and decreased nigral neuronal density in early Braak stages.⁵⁸ Aggregated α Syn could be a suitable imaging biomarker in α -synucleinopathies. However, currently, aggregated α Syn and Lewy pathology inclusion bodies cannot be assessed ante-mortem with SPECT or PET radiotracers. An α Syn specific radiotracer capable for early diagnosis of α -synucleinopathies, potentially aiding differential diagnosis, is highly desired.

Additionally, as might be concluded from the information above, aggregated α Syn could be a suitable target for therapy in α -synucleinopathies considering its key role in Lewy pathology. There are currently no registered disease-modifying agents available for the treatment of α -synucleinopathies, current treatments are only acting on symptoms.⁵⁹ Several biological⁶⁰⁻⁶⁸ and small molecule compounds⁶⁹⁻⁷¹ targeting α Syn aggregates are in clinical and pre-clinical development. However, currently there is no way to directly measure if or how much they lower the levels of aggregated α Syn in the brain. Ideally, the same compound targeting aggregated α Syn can be used for therapy where it exerts a desired clinical effect and as a radiotracer for diagnosis and therapy monitoring where it confirms the specific target engagement and demonstrates the clinical benefit. The availability of α Syn radiotracer for monitoring α Syn load, target engagement, disease progression, and therapy response would greatly benefit the development process. In a comparable way, the availability of PET radiotracers for amyloid pathology in AD facilitated the clinical development of aducanumab in terms of clinical trials subject recruitment and efficacy evaluation,⁷²⁻⁷⁴ despite the controversial and yet to be established efficacy and clinical benefit of aducanumab. This example in the field of neurodegeneration demonstrates the potential of an α Syn radiotracer facilitating targeted therapy development in α -synucleinopathies.

Strategies for α Syn Radiotracer Development

Recommendations in the α Syn imaging field highlight the pressing need for SPECT and PET radiotracers to aid the diagnosis and treatment in α -synucleinopathies.

The Michael J. Fox Foundation established the α-Synuclein Imaging Consortium in 2011, launched a \$2 million prize for the development of a selective αSyn PET radiotracer in 2016, and in 2019 announced the \$10 million Ken Griffin Alpha-synuclein Imaging Competition. Despite these efforts, to date there is no radiotracer available for imaging αSyn. Several small-molecule ligands have been developed and tested for in vivo imaging and detection of aggregated αSyn and Lewy pathology inclusion bodies (LBs/LNs/GCIs) in α-synucleinopathies. This part of the review covers the used strategies for αSyn ligand development and findings from ligands that advanced to pre-clinical and clinical evaluation. More extensive and radiochemical-oriented description of all developed and tested probes targeting aggregated αSyn has been reviewed elsewhere.⁷⁵⁻⁷⁸

Repurposed Radiotracers from AD

Amyloid-β (Aβ), tau and αSyn form similar β-sheet structures on aggregation.⁷⁹⁻⁸¹ Presumably, imaging probes binding to Aβ or tau aggregates have the potential to bind to αSyn aggregates. Hence, the radiotracers [¹¹C]PIB, [¹¹C]BF227, [¹¹C]-PBB3, and its structural analog [³H]-C05-01 were evaluated for imaging αSyn aggregates. [¹¹C]PIB is a thioflavin-T derivative and the gold standard for staining β-sheet structured protein aggregates. Although PIB exhibited high binding affinity ($K_d = 4$ nM) for recombinant αSyn fibrils,⁸² it did not bind LBs-containing DLB brain homogenates.⁸³ In addition, PIB did not show interaction with PD brain sections containing LBs/LNs on autoradiography,⁸² and displayed poor binding selectivity for αSyn versus Aβ in DLB brain sections.⁸³ [¹⁸F]BF227 was developed to image Aβ plaques in AD.⁸⁴ The high binding affinity of [¹⁸F]BF227 ($K_d = 9.6$ nM) for recombinant αSyn fibrils conflicted with failed binding to LBs-containing DLB brain homogenates⁸⁵ and failed detection of GCIs in MSA brain with autoradiography.⁸⁶ [¹¹C]BF227 showed higher uptake in GCI-rich brain regions of MSA patients relative to control subjects (Fig. 3A), however, the group differences were small, and many individual values overlapped indicating low in vivo selectivity for aggregated αSyn.⁸⁷ In addition, as expected, this radiotracer showed high binding affinity to Aβ plaques in AD patients and clearly differentiated them from control subjects,⁸⁴ demonstrating high in vivo affinity for Aβ plaques. [¹¹C]-PBB3 emerged as one of the first generation tau radiotracers.⁸⁸ [¹¹C]-PBB3 showed in vitro autoradiographic binding to GCIs only in a subset of MSA cases,⁸⁹ and in vivo higher uptake in the brain of one MSA case compared to control subject (Fig. 3B).⁹⁰ These initial studies suggested that [¹¹C]-PBB3 displays some binding to

aggregated αSyn and the PBB3 analog [³H]-C05-01 was developed and displayed a reasonable binding affinity ($K_d = 24$ nM) to recombinant αSyn fibrils. In vitro autoradiography using tissue microarrays and fresh-frozen brain tissue showed that, whereas [³H] C05-01 selectively binds to αSyn aggregates in PD and MSA brain, the ligand also selectively binds to Aβ and tau aggregates in AD, and therefore, has limited specificity for αSyn.⁹¹ Apparently, these repurposed radiotracers lack selectivity toward αSyn aggregates, and therefore, are of no interest for further development because of limited applicability in α-synucleinopathies.

Re-Explored Chemical Entities

[¹¹C]anle253b is an analog of anle138b, an αSyn fibrilization inhibitor with a demonstrated therapeutic activity in rodent models.^{70,92} In vitro binding assays showed that [¹¹C]anle253b preferentially binds to αSyn fibrils over oligomeric and monomeric species. However, the high log *P* of 5.21 and atypical brain uptake kinetic curves in healthy rats indicated the need for physicochemical properties optimization.⁹³ More recently, MODAG-001 was developed by modifying the chemical structure of anle253b to overcome the suboptimal in vivo pharmacokinetic properties of [¹¹C] anle253b. In in vitro binding assays, [³H]MODAG-001 combined a very high binding affinity toward recombinant αSyn ($K_d = 0.6 \pm 0.1$ nM) with a good selectivity versus Aβ ($K_d = 20 \pm 10$ nM) and tau ($K_d = 19 \pm 6.4$ nM). [¹¹C]MODAG-001 showed suitable pharmacokinetic and biodistribution properties in mouse, and (d3)-[¹¹C]MODAG-001 binding to recombinant αSyn fibrils was confirmed in fibril-inoculated rat striata using in vivo PET imaging, but two radiometabolites from both radiotracers were detected in plasma and brain. Additionally, in vitro autoradiography showed no binding of (d3)-[¹¹C]MODAG-001 to αSyn aggregates in human brain sections of DLB cases. Nonetheless, MODAG-001 is still a promising lead structure for further development as it combines a high affinity and good selectivity with a suitable pharmacokinetics and biodistribution properties.⁹⁴ [¹¹C]14 ((3R)-7-(4'-[¹¹C]methoxynaphthyl-1-yl)methyl)-5-oxo-8-(3-(trifluoromethyl)-phenyl)-2,3-dihydro-5H-thiazolo [3,2-a]pyridine-3-carboxylic acid acetoxymethyl ester) is an analog of thiazolo-2-pyridone, FN075, an αSyn fibrillization accelerator, penetrated the brain in non-human primates, but showed poor pharmacokinetic properties for in vivo imaging with low initial brain uptake (≈ 0.8 standard uptake value), slow brain penetration (peak at 10 minutes post injection), and slow washout. Nearly half the radioactivity signal was still detectable as nonspecific binding by the end of the PET scan, which could be attributed to the high lipophilicity

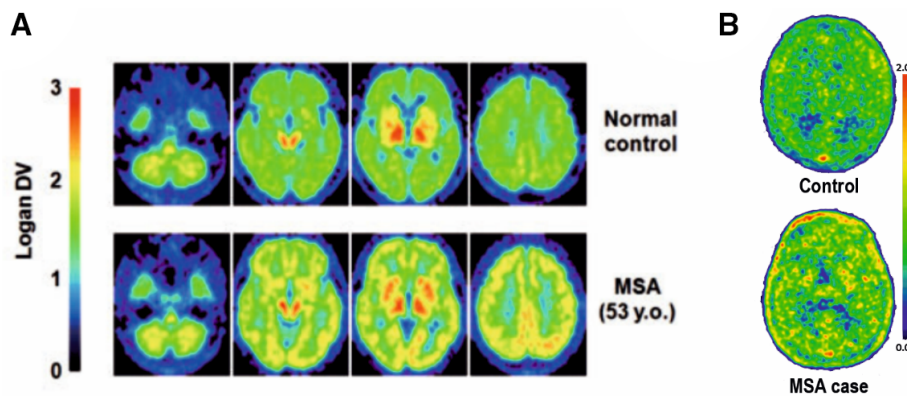


FIG. 3. Attempts made to image Lewy pathology in vivo in multiple system atrophy (MSA). **(A)** PET images of a normal subject and a case of MSA imaged with [^{11}C]BF227. The MSA case show increased cortical, basal ganglia, and white matter signal compared to the normal case, reflecting αSyn aggregates.⁸⁷ **(B)** PET images of a healthy control case and an MSA case imaged with [^{11}C]PBB3. The MSA brain showed mild uptake in basal ganglia, frontal and parietal, and cortex compared to the healthy control brain, which showed a lack of regionally specific binding.⁹⁰

($\text{cLogP} = 6.1$).⁹⁵ The lead compound [^{18}F]46a developed based on the 3-(benzylidene)-indolin-2-one scaffold showed good binding affinity toward αSyn ($K_d = 8.9$ nM) and selectivity over $\text{A}\beta$ ($K_d = 271$ nM) and tau ($K_d = 50$ nM) in in vitro binding assays. However, the high $\log P$ of 4.18 and structural limitations indicated potentially high nonspecific binding that terminated further development.⁹⁶

In Silico Modelling and High-Throughput Screening

This strategy uses chemical entities with reasonable binding properties to aggregated αSyn to generate potential ligands that undergo computational evaluation on αSyn fibrils structure to test ligands engagement with αSyn fibrils and feasibility for binding sites detection. The iodo-derivative [^{125}I]61 (2-(3,4-dimethylphenoxy)-N-(3-(4-[^{125}I]iodophenyl)isoxazol-5-yl) acetamide) bound to confirmed binding site location on αSyn fibrils structure and showed high in vitro binding affinity to αSyn fibrils ($K_d = 1.06$ nM) extracted from the brain of PD mouse model and 5-fold selectivity over $\text{A}\beta_{42}$ fibrils ($K_d = 5.56$ nM). Using in vitro autoradiography, [^{125}I]61 demonstrated binding to αSyn -rich regions on sections from the same PD mouse model, but also nonspecific binding. The non-ideal physicochemical properties of [^{125}I]61 made this radiotracer unsuitable for further evaluation.⁹⁷ [^{18}F]2FBox showed high in vitro binding affinity toward αSyn fibrils ($K_d = 3.3 \pm 2.8$ nM, $B_{\text{max}} = 0.128 \pm 0.025$ pmol/nmol of fibril) and selectivity over $\text{A}\beta_{1-42}$ ($K_d = 145.3 \pm 114.5$ nM, $B_{\text{max}} = 0.592 \pm 0.251$ pmol/nmol of fibril). However, [^{18}F]2FBox failed to label LBs in PD and MSA brain sections. Additionally, [^{18}F]2FBox non-selectively detected both αSyn and $\text{A}\beta_{1-42}$ fibrils in rats striatum injected with both proteins using

in vitro autoradiography and failed to detect injected αSyn fibrils in rats despite the reasonable pharmacokinetic properties using in vivo PET.⁹⁸

Lessons Learned from αSyn Radiotracer Development

Collectively, the above mentioned small-molecule ligands are not yet suitable for in vivo imaging of aggregated αSyn because of suboptimal binding affinity and/or specificity and physicochemical properties. However, many lessons have been learned that will help in the successful development of αSyn -selective radiotracer in the future. First, the radiotracer should possess high binding affinity and selectivity; blood brain barrier (BBB) penetration; adequate initial brain uptake and washout; absence of BBB-penetrating radioactive metabolites; absence of substrate activity on BBB efflux transporters, such as P-glycoprotein; optimal physicochemical properties, such as lipophilicity ($\log D$ 1–3 at pH 7.4) for effective BBB penetration via passive diffusion and limited nonspecific tissue binding.⁹⁹ Second, in vivo imaging of aggregated αSyn is challenging and the following aspects must be considered early in development. (1) Although αSyn aggregates (oligomers and fibrils) do spread in the brain with evidence for extracellular propagation,^{37,100-102} they are predominantly present in intracellular inclusion bodies.²⁷ Consequently, the radiotracer needs to cross the BBB and ideally penetrates the cell membrane either by an active transport mechanism or passively to bind intracellular αSyn aggregates. (2) Different types of cells are involved in different α -synucleinopathies. In PD and DLB, αSyn accumulates in the neurons as LBs and LNs,³¹ whereas in MSA αSyn accumulates in oligodendrocytes as GCIs.³³ Additionally, GCIs are structurally different from LBs and LNs^{35,36} (also see point 6 below). An

important implication of that to radiotracers design is that structural modifications could be necessary for imaging αSyn aggregates in oligodendrocytes GCI and neurons LBs/LNs. (3) Unlike Aβ plaques but like tau, αSyn aggregates present in the brain are low in abundance and small in size, which is demanding for adequate in vivo detection and visualization. The area covered by LBs in DLB brain sections was found to be 40× smaller than that of Aβ plaques.⁸³ Nonetheless, in vitro binding assays using recombinant αSyn fibrils and fibrils from PD brain showed that αSyn has sufficient binding site density to enable in vivo imaging, which is similar to tau, but less than Aβ plaques.¹⁰³ Moreover, the binding site density of αSyn (SIL23, αSyn fibrils $B_{\max} = 108\text{--}895\text{ nmol/L}$ ¹⁰³) is significantly higher than that of pre- and post-synaptic receptors in the dopaminergic and serotonergic systems (raclopride, D2 receptor $B_{\max} = 30\text{--}40\text{ nmol/L}$), for which PET radiotracers have been developed.¹⁰⁴ (4) The small-molecule ligands developed till now bind to the αSyn β-sheet structure, which is similar to tau NFTs and Aβ plaques.⁷⁹⁻⁸¹ Additionally, αSyn co-localizes with numerous other proteins inside the inclusion bodies including tau.²⁷ Therefore, developed ligands suffered from poor binding selectivity. These facts are problematic because Lewy pathology in PD and DLB often co-occurs with Aβ and tau pathologies as a result of co-existing AD and/or other tauopathies,^{105,106} which has been shown by amyloid PET imaging.¹⁰⁷ To overcome this challenge, more efforts are needed with the current and novel in silico approaches to develop small-molecule ligands that are specific for aggregated αSyn binding-sites.^{97,108} (5) αSyn aggregates form soluble oligomers and insoluble fibrillary structures in α-synucleinopathies.¹⁰⁹⁻¹¹¹ Ideally, these different structural conformations demand radiotracers with conformation-specific binding affinities for accurate and selective in vivo detection. (6) The majority of used αSyn materials in radiotracer development are recombinant fibrils. αSyn fibrils generated in vitro are not reliable for the evaluation of αSyn ligands because of poor reproducibility, and more importantly, not representing aggregated αSyn in the human brain. Thus far, screening ligands for αSyn aggregates has been based on the available NMR¹¹² and cryogenic electron microscopy (cryo-EM)¹¹³⁻¹¹⁵ structures of recombinant αSyn fibrils (Fig. 4B,C). More recently, the cryo-EM structure of authentic αSyn fibrils from the MSA brain has been solved for the first time, revealing that there are two types of αSyn fibrils that consist of four distinct protofibrils, which differ in conformation, folding, and predominance in different brain regions (Fig. 4A).¹¹⁶ These advancements are highly relevant to the development of αSyn-specific radiotracer considering that compared to the MSA-αSyn fibrils, in vitro-made αSyn fibrils are smaller, differ in conformation, and consist of fewer

protofibrils. Unlike αSyn fibrils from MSA brain, αSyn fibrils from DLB or PD brain are thinner and do not twist, which precluded solving their 3D cryo-EM structure. However, based on 2D analysis, αSyn from MSA and DLB are distinct.¹¹⁶ Therefore, using human brain tissue from α-synucleinopathies brains is the most reliable, although availability is problematic, especially without other co-pathologies.¹¹⁷ (7) Suitable rodent models of Lewy pathology need to be used when evaluating the in vivo binding properties of αSyn radiotracers. For a reliable evaluation, rodent models must develop sufficient Lewy pathology inclusion bodies using αSyn materials from α-synucleinopathies brain. Several reliable models exist, including the genetic models developed in mouse based on SNCA wild type or mutated variants overexpression, but the downside of the genetic models is that the developed Lewy pathology is generally mild and manifest after several months.¹¹⁸⁻¹²⁰ A more practical alternative is the seeding model developed in mouse and rat by injecting αSyn aggregates in the brain, which can develop acute, abundant, and progressive Lewy pathology.^{121,122} Those models develop αSyn aggregates with an onset of 1 month, which is not feasible with the genetic models.¹²³ Viral vector models are another alternative, which are developed in mouse and rat using a virus vector to deliver SNCA to the substantia nigra dopaminergic neurons where αSyn is overexpressed. Those models develop αSyn aggregates with an onset of days to weeks. An additional advantage of the viral vector models is that αSyn is overexpressed only in the substantia nigra dopaminergic neurons; therefore, αSyn aggregates are only localized to the nigrostriatal pathway, which is substantia nigra and striatum. This characteristic could be beneficial for evaluating radiotracers in vivo binding selectivity. Ideally, a radiotracer selective for αSyn aggregates should demonstrate binding in the substantia nigra and the striatum because of the presence of αSyn aggregates, and no nonspecific binding in other brain regions because of the absence of αSyn aggregates.¹²⁴⁻¹²⁶ The double-hit strategy combines the genetic and seeding models,¹²⁷⁻¹²⁹ or the seeding and viral vector models^{130,131} to generate αSyn aggregates that are even more abundant than a single model could generate.

Antibody-Based Ligands: A Complementary Strategy for Imaging αSyn Aggregates

Antibody-based radiotracers offer superior binding affinity and specificity over small-molecule radiotracers, and the capacity to target different structural conformations of a protein. Typically, antibodies have exceptionally high target binding affinity in the subnanomolar

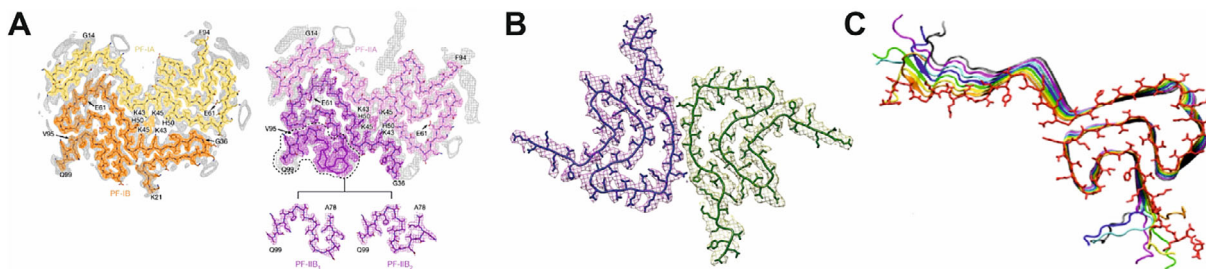


FIG. 4. Two structural models of α Syn fibril extracted from multiple system atrophy (MSA) brain resolved by cryo-EM¹¹⁶ (A) differ from structural models of recombinant human α Syn fibril resolved by cryo-EM¹¹⁵ (B) and solid-state NMR¹¹² (C).

range,¹³² such affinity range for small molecules is unattainable. In the case of α Syn aggregates, antibodies binding affinity is reported in the picomolar range (BIIB054 $K_d = \sim 120$ pM,⁶⁰ MEDI1341 $K_d = 74$ pM⁶³), whereas the highest reported binding affinity of a radiolabeled small-molecule is in the nanomolar range (MODAG-001 $K_d = 0.6 \pm 0.1$ nM.⁹⁴ Radiolabeled antibodies demonstrated a proof of concept in imaging A β aggregates using AD rodent models (Fig. 5A).¹³³⁻¹³⁶ Because antibodies are large molecules by nature, the primary limitation for radiolabeled antibodies is their restrained passage across the BBB. Approximately, only 0.1% of a peripherally administered antibody dose reaches the brain.^{137,138} Active transport into the brain by receptor-mediated transcytosis is one way to overcome this limitation where the transferrin receptor has been a successful shuttling system for enhanced targeting of A β aggregates by radiolabeled antibodies in AD mouse models.^{133,136,139,140} Another way is to use microbubble-mediated focused ultrasound (FUS), which is a non-invasive imaging technique for enhanced drug delivery to the brain by reversibly opening the BBB.^{141,142} Oncological applications using FUS in combination with radiolabeled-antibody PET radiotracers showed that this technique significantly enhanced BBB penetration of bevacizumab (Fig. 5B)¹⁴³ and ¹⁸F-FBPA-F¹⁴⁴ in glioma-bearing mouse and rat models, respectively.

A secondary limitation associated with radiolabeled antibodies is the typically slow pharmacokinetics of antibodies, that is slow target accumulation and clearance, which requires the use of long-lived radionuclides for PET imaging and therefore, results in a high radiation exposure.¹⁴⁵ This can partly be overcome by the introduction of total body PET/CT scanners, which have significantly higher sensitivity than the current standard PET/CT scanners.¹⁴⁶ Another approach to overcome this limitation is the *in vivo* pretargeting. In this approach, a slow kinetic tagged-antibody is administered and allowed to bind the desired target days before a fast kinetic radiolabeled agent with a short-lived radionuclide is administered and both components having to bind to each other in the body¹⁴⁷ and more

recently in the brain.¹⁴⁸ This way, the pretargeting strategy significantly reduces the radiation exposure as it allows the use of short-lived radionuclides that would

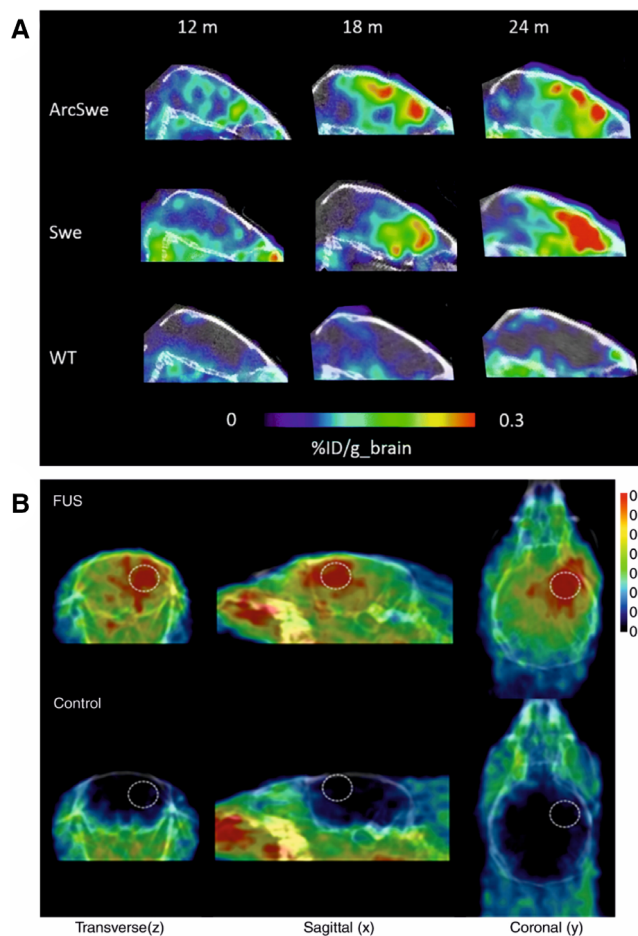


FIG. 5. Strategies to overcome the limited passage of radiolabeled antibodies across the blood brain barrier. (A) Sagittal PET images obtained at 3 days after administration of the bispecific radioligand [¹²⁴I]8D3-F (ab')₂-h158 in two mouse models of Alzheimer's disease (ArcSwe and Swe) and wild-type mice at 12, 18, and 24 months.¹⁴⁹ (B) Representative decay-corrected PET/CT fused images in focused ultrasound (FUS)-treated animals (top row) and control animals (bottom row) obtained 15 minutes after injection of ⁶⁸Ga-bevacizumab. Dashed circles, focused ultrasound targeting sites.¹⁴³

otherwise be incompatible with antibodies. Sehlin and colleagues¹⁴⁹ thoroughly reviewed the antibody-based ligands strategy and the associated limitations for imaging misfolded proteins in the brain, and Van Dongen and colleagues¹⁵⁰ reviewed the emerging application of antibodies in a theranostic setting where the same antibody is used for diagnosis as well as therapy.

In the case of imaging aggregated α Syn with radiolabeled antibodies, the predominant intracellular localization of aggregated α Syn²⁷ forms an intrinsic accessibility problem. Unlike passing the BBB, there does not seem to be an immediate solution for this limitation. Nonetheless, radiolabeled antibodies could still have a potential application in imaging the extracellular α Syn aggregates spreading in the brain, although it has to be seen whether extracellular α Syn concentrations are high enough to allow PET imaging. α Syn cell-to-cell spreading has been well demonstrated using in vitro cell culture studies^{151,152} and animal models.^{130,153} In humans, LBs and LN have been shown to spread from host to graft dopaminergic neurons in the substantia nigra of PD patients (Fig. 1B),¹⁰⁰ which has been replicated in a PD mouse model.¹⁰¹ Additionally, Lewy pathology propagation throughout the central nervous system (CNS)³⁷ strongly suggests α Syn spreading between interconnected brain regions. In silico modeling for the development of MEDI1341 antibody, which binds monomeric and aggregated α Syn, predicted that achieving target affinity of <100 pM is required to potently bind extracellular levels of α Syn in the brain. Accordingly, MEDI1341 ($K_d = 74$ pM) showed to successfully bind and lower the levels of extracellular α Syn in rats, monkeys, and mouse model of Lewy pathology.⁶³ The case of MEDI1341 antibody demonstrates that an antibody can successfully bind extracellular α Syn aggregates if a similar affinity level is accomplished, even though experimental measurements of the needed level of extracellular α Syn aggregates in the brain for an antibody to bind successfully are lacking. Another potential application of radiolabeled antibodies in imaging aggregated α Syn is that antibodies could have higher binding specificity for particular target species compared to small molecules. It has been demonstrated in AD mouse models that a radiolabeled antibody binding to A β oligomers and protofibrils is better able to monitor changes in A β levels after therapeutic intervention compared to a small molecule tracer binding only to A β fibrillar structures.¹⁵⁴ Considering that imaging intracellular α Syn aggregates using radiolabeled antibodies seems to be infeasible, it could be that a radiolabeled antibody binding predominantly to spreading α Syn oligomers, but not fibrils or vice versa, in the extracellular domain is better able to monitor changes in α Syn aggregates level and therapy response than a small-molecule radiotracer binding predominantly to intracellular α Syn aggregates. Collectively, the

evidence from the above-mentioned studies justifies evaluating the potential use of radiolabeled antibodies for imaging extracellular α Syn aggregates.

Future Perspective

A-synucleinopathies and PD in particular will continue to be a heavy burden on healthcare systems and societies with the increasing prevalence of these disorders. Clearly, there is a paramount diagnostic and therapeutic unmet medical need to be addressed in α -synucleinopathies. The availability of the first α Syn PET or SPECT radiotracer will have a significant potential in resolving these unmet needs and improve our understanding of Lewy pathology and α -synucleinopathies. An α Syn radiotracer could possibly achieve early and differential diagnosis of the different α -synucleinopathies. Additionally, it could expedite the development of the several promising disease-modifying agents currently in clinical trials. Although developing a radiotracer specific for α Syn aggregates proved to be challenging, our understanding of this complex task improved vastly and considerable progress has been made. The learned lessons from previously tested radiotracers must be carefully considered in future developments, where both small-molecule ligands and antibodies should be the focus of development. For small-molecule ligands, the in silico approach is particularly promising to overcome the specificity hurdle. Small-molecule ligands offer superior BBB passage, cellular penetration, and target-reachability compared to radiolabeled antibodies, which are limited by poor BBB passage and cellular penetration, but offer superior binding specificity and the potential for theranostic applications. Additionally, based on the current evidence from radiolabeled antibodies evaluated in pre-clinical applications in AD, and the localization of α Syn aggregates in the brain, radiolabeled antibodies could have a complementary rather than an alternative role to small-molecule ligands. Neither radiolabeled antibodies nor small-molecule ligands are currently capable of imaging the complete spectrum of α Syn aggregates in the brain. Small-molecule ligands are better suited for imaging total (extra and intracellular) α Syn aggregates, whereas radiolabeled antibodies could be better suited for imaging extracellular α Syn aggregates and for monitoring the levels of α Syn aggregates and therapy response in α -synucleinopathies. ■

Data Availability Statement

The data that support the findings of this study are openly available in PubMed and Embase databases.

References

- Coon EA, Singer W, Low PA. Pure autonomic failure. *Mayo Clin Proc* 2019;94:2087–2098.
- Rajput AH, Rozdilsky B, Rajput A. Accuracy of clinical diagnosis in parkinsonism—a prospective study. *Can J Neurol Sci* 1991;18:275–278.
- Jankovic J, Rajput AH, McDermott MP, Perl DP. The evolution of diagnosis in early Parkinson disease. *Arch Neurol* 2000;57:369–372. <https://doi.org/10.1001/archneur.57.3.369>
- Palmqvist S, Hansson O, Minthon L, Londos E. Practical suggestions on how to differentiate dementia with Lewy bodies from Alzheimer's disease with common cognitive tests. *Int J Geriatr Psychiatry* 2009;24:1405–1412. <https://doi.org/10.1002/gps.2277>
- Weisman D, Cho M, Taylor C, Adame A, Thal LJ, Hansen LA. In dementia with Lewy bodies, Braak stage determines phenotype, not Lewy body distribution. *Neurology* 2007;69:356–359.
- Tiraboschi P, Attems J, Thomas A, Brown A, Jaros E, Lett DJ, et al. Clinicians' ability to diagnose dementia with Lewy bodies is not affected by β -amyloid load. *Neurology* 2015;84:496–499.
- Hughes AJ, Daniel SE, Ben-Shlomo Y, Lees AJ. The accuracy of diagnosis of parkinsonian syndromes in a specialist movement disorder service. *Brain* 2002;125:861–870. <https://doi.org/10.1093/brain/awf080>
- Maass F, Schulz I, Lingor P, Mollenhauer B, Bähr M. Cerebrospinal fluid biomarker for Parkinson's disease: an overview. *Mol Cell Neurosci* 2019;97:60–66.
- Feigin VL, Krishnamurthi RV, Theadom AM, Abajobir AA, Mishra SR, Ahmed MB, et al. Global, regional, and national burden of neurological disorders during 1990–2015: a systematic analysis for the global burden of disease study 2015. *Lancet Neurol* 2017;16:877–897.
- Ray Dorsey E, Elbaz A, Nichols E, Abd-Allah F, Abdelalim A, Adsuar JC, et al. Global, regional, and national burden of Parkinson's disease, 1990–2016: a systematic analysis for the global burden of disease study 2016. *Lancet Neurol* 2018;17:939–953.
- Blauwendraat C, Nalls MA, Singleton AB. The genetic architecture of Parkinson's disease. *Lancet Neurol* 2020;19:170–178.
- Polymeropoulos MH, Lavedan C, Leroy E, Ide SE, Dehejia A, Dutra A, et al. Mutation in the α -synuclein gene identified in families with Parkinson's disease. *Science* 1997;276:2045–2047.
- Krüger R, Kuhn W, Müller T, Woitalla D, Graeber M, Kösel S, et al. Ala30Pro mutation in the gene encoding α -synuclein in Parkinson's disease. *Nat Genet* 1998;18:106–108.
- Zarranz JJ, Alegre J, Gómez-Esteban JC, Lezcano E, Ros R, Ampuero I, et al. The new mutation, E46K, of α -Synuclein causes Parkinson and Lewy body dementia. *Ann Neurol* 2004;55:164–173. <https://doi.org/10.1002/ana.10795>
- Appel-Cresswell S, Vilarino-Guell C, Encarnacion M, Sherman H, Yu I, Shah B, et al. Alpha-synuclein p.H50Q, a novel pathogenic mutation for Parkinson's disease. *Mov Disord* 2013;28:811–813. <https://doi.org/10.1002/mds.25421>
- Kiely AP, Asi YT, Kara E, Limousin P, Ling H, Lewis P, et al. A-synucleinopathy associated with G51D SNCA mutation: a link between Parkinson's disease and multiple system atrophy? *Acta Neuropathol* 2013;125:753–769. <https://doi.org/10.1007/s00401-013-1096-7>
- Lesage S, Anheim M, Letournel F, Bousset L, Honoré A, Rozas N, et al. G51D α -synuclein mutation causes a novel parkinsonian-pyramidal syndrome. *Ann Neurol* 2013;73:459–471. <https://doi.org/10.1002/ana.23894>
- Pasanen P, Myllykangas L, Siitonen M, Raunio A, Kaakkola S, Lyytinen J, et al. A novel α -synuclein mutation A53E associated with atypical multiple system atrophy and Parkinson's disease-type pathology. *Neurobiol Aging* 2014;35:2180–2180.
- Devine MJ, Gwinn K, Singleton A, Hardy J. Parkinson's disease and α -synuclein expression. *Mov Disord* 2011;26:2160–2168.
- Miller DW, Hague SM, Clarimon J, Baptista M, Gwinn-Hardy K, Cookson MR, et al. α -Synuclein in blood and brain from familial Parkinson disease with SNCA locus triplication. *Neurology* 2004;62:1835–1838.
- Singleton AB, Farrer M, Johnson J, Singleton A, Hague S, Kachergus J, et al. α -Synuclein locus triplication causes Parkinson's disease. *Science* 2003;302:841. <https://doi.org/10.1126/science.1090278>
- Iwai A, Masliah E, Yoshimoto M, Ge N, Flanagan L, Rohan de Silva HA, et al. The precursor protein of non-A β component of Alzheimer's disease amyloid is a presynaptic protein of the central nervous system. *Neuron* 1995;14:467–475.
- Wang C, Zhao C, Li D, Tian Z, Lai Y, Diao J, et al. Versatile structures of α -synuclein. *Front Mol Neurosci* 2016;9:48. <https://doi.org/10.3389/fnmol.2016.00048/abstract>
- Jakes R, Spillantini MG, Goedert M. Identification of two distinct synucleins from human brain. *FEBS Lett* 1994;345:27–32. [https://doi.org/10.1016/0014-5793\(94\)90395-5](https://doi.org/10.1016/0014-5793(94)90395-5)
- Burré J, Sharma M, Tssetsenis T, Buchman V, Etherton MR, Südhof TC. α -Synuclein promotes SNARE-complex assembly in vivo and in vitro. *Science* 2010;329:1663–1667.
- Conway KA, Harper JD, Lansbury PT. Fibrils formed in vitro from α -synuclein and two mutant forms linked to Parkinson's disease are typical amyloid. *Biochemistry* 2000;39:2552–2563. <https://doi.org/10.1021/bi991447r>
- Wakabayashi K, Tanji K, Odagiri S, Miki Y, Mori F, Takahashi H. The Lewy body in Parkinson's disease and related neurodegenerative disorders. *Mol Neurobiol* 2013;47:495–508. <https://doi.org/10.1007/s12035-012-8280-y>
- Anderson JP, Walker DE, Goldstein JM, de Laat R, Banducci K, Caccavello RJ, et al. Phosphorylation of Ser-129 is the dominant pathological modification of α -synuclein in familial and sporadic lewy body disease. *J Biol Chem* 2006;281:29739–29752.
- Zhou J, Broe M, Huang Y, Anderson JP, Gai WP, Milward EA, et al. Changes in the solubility and phosphorylation of α -synuclein over the course of Parkinson's disease. *Acta Neuropathol* 2011;121:695–704. <https://doi.org/10.1007/s00401-011-0815-1>
- Mahul-Mellier AL, Bartscher J, Maharjan N, Weerens L, Croisier M, Kuttler F, et al. The process of Lewy body formation, rather than simply α -synuclein fibrillization, is one of the major drivers of neurodegeneration. *Proc Natl Acad Sci U S A* 2020;117:4971–4982. <https://doi.org/10.6084/m9.figshare.11842389.v2>
- Arima K, Ueda K, Sunohara N, Hirai S, Izumiyama Y, Tonozuka-Uehara H, et al. Immunoelectron-microscopic demonstration of NACP/ α -synuclein-epitopes on the filamentous component of Lewy bodies in Parkinson's disease and in dementia with Lewy bodies. *Brain Res* 1998;808:93–100.
- Braak H, Ghebremedhin E, Rüb U, Bratzke H, del Tredici K. Stages in the development of Parkinson's disease-related pathology. *Cell Tissue Res* 2004;318:121–134. <https://doi.org/10.1007/s00441-004-0956-9>
- Wakabayashi K, Yoshimoto M, Tsuji S, Takahashi H. α -Synuclein immunoreactivity in glial cytoplasmic inclusions in multiple system atrophy. *Neurosci Lett* 1998;249:180–182.
- Fujiwara H, Hasegawa M, Dohmae N, Kawashima A, Masliah E, Goldberg MS, et al. α -Synuclein is phosphorylated in synucleinopathy lesions. *Nat Cell Biol* 2002;4:160–164.
- Peng C, Gathagan RJ, Covell DJ, Medellin C, Stieber A, Robinson JL, et al. Cellular milieu imparts distinct pathological α -synuclein strains in α -synucleinopathies. *Nature* 2018;557:558–563.
- Lengyel-Zhand Z, Ferrie JJ, Janssen B, Hsieh CJ, Graham T, Xu KY, et al. Synthesis and characterization of high affinity fluorogenic α -synuclein probes. *Chem Commun* 2020;56:3567–3570.
- Braak H, del Tredici K, Rüb U, de Vos RAI, Jansen Steur EN, Braak E. Staging of brain pathology related to sporadic Parkinson's disease. *Neurobiol Aging* 2003;24:197–211.
- McKeith I, Mintzer J, Aarsland D, Burn D, Chiu H, Cohen-Mansfield J, et al. Dementia with Lewy bodies. *Lancet Neurol* 2004;3:19–28.
- Walker L, Stefanis L, Attems J. Clinical and neuropathological differences between Parkinson's disease, Parkinson's disease dementia and dementia with Lewy bodies – current issues and future

- directions. *J Neurochem* 2019;150:467–474. <https://doi.org/10.1111/jnc.14698>
40. Brettschneider J, Suh ER, Robinson JL, Fang L, Lee EB, Irwin DJ, et al. Converging patterns of α -synuclein pathology in multiple system atrophy. *J Neuropathol Exp Neurol* 2018;77:1005–1016.
 41. Beach TG, Adler CH, Lue LF, Sue LI, Bachalakuri J, Henry-Watson J, et al. Unified staging system for Lewy body disorders: correlation with nigrostriatal degeneration, cognitive impairment and motor dysfunction. *Acta Neuropathol* 2009;117:613–634. <https://doi.org/10.1007/s00401-009-0538-8>
 42. Alafuzoff I, Ince PG, Arzberger T, Al-Sarraj S, Bell J, Bodi I, et al. Staging/typing of Lewy body related α -synuclein pathology: a study of the BrainNet Europe consortium. *Acta Neuropathol* 2009;117:635–652.
 43. Postuma RB, Berg D, Stern M, Poewe W, Olanow CW, Oertel W, et al. MDS clinical diagnostic criteria for Parkinson's disease. *Mov Disord* 2015;30:1591–1601. <https://doi.org/10.1002/mds.26424>
 44. McKeith IG, Boeve BF, Dickson DW, Halliday G, Taylor JP, Weintraub D, et al. Diagnosis and management of dementia with Lewy bodies. *Neurology* 2017;89:88–100.
 45. Gilman S, Wenning GK, Low PA, Brooks DJ, Mathias CJ, Trojanowski JQ, et al. Second consensus statement on the diagnosis of multiple system atrophy. *Neurology* 2008;71:670–676.
 46. Suwijn SR, van Boheemen CJM, de Haan RJ, Tissingh G, Booij J, de Bie RMA. The diagnostic accuracy of dopamine transporter SPECT imaging to detect nigrostriatal cell loss in patients with Parkinson's disease or clinically uncertain parkinsonism: a systematic review. *EJNMMI Res* 2015;5.
 47. McKeith I, O'Brien J, Walker Z, Tatsch K, Booij J, Darcourt J, et al. Sensitivity and specificity of dopamine transporter imaging with 123I-FP-CIT SPECT in dementia with Lewy bodies: a phase III, multicentre study. *Lancet Neurol* 2007;6:305–313.
 48. Cousins O, Yousaf T, Wilson H, Pagano G, Politis M. Molecular imaging of dementia with Lewy bodies. *Int Rev Neurobiol* 2019;144:59–93.
 49. Iranzo A, Valldeoriola F, Lomeña F, Molinuevo JL, Serradell M, Salameiro M, et al. Serial dopamine transporter imaging of nigrostriatal function in patients with idiopathic rapid-eye-movement sleep behaviour disorder: a prospective study. *Lancet Neurol* 2011;10:797–805.
 50. Ponsen MM, Stoffers D, Booij J, van Eck-Smit BLF, Wolters EC, Berendse HW. Idiopathic hyposmia as a preclinical sign of Parkinson's disease. *Ann Neurol* 2004;56:173–181. <https://doi.org/10.1002/ana.20160>
 51. Noyce AJ, Lees AJ, Schrag AE. The prediagnostic phase of Parkinson's disease. *J Neurol Neurosurg* 2016;87:871–878.
 52. Hellwig S, Amtage F, Kreft A, Buchert R, Winz OH, Vach W, et al. [18F]FDG-PET is superior to [123I]IBZM-SPECT for the differential diagnosis of parkinsonism. *Neurology* 2012;79:1314–1322.
 53. van Rumund A, Green AJE, Fairfoul G, Esselink RAJ, Bloem BR, Verbeek MM. α -Synuclein real-time quaking-induced conversion in the cerebrospinal fluid of uncertain cases of parkinsonism. *Ann Neurol* 2019;85:777–781.
 54. Mattila PM, Rinne JO, Helenius H, Dickson DW, Røyttä M. Alpha-synuclein-immunoreactive cortical Lewy bodies are associated with cognitive impairment in Parkinson's disease. *Acta Neuropathol* 2000;100:285–290. <https://doi.org/10.1007/s004019900168>
 55. Harding AJ, Stimson E, Henderson JM, Halliday GM. Clinical correlates of selective pathology in the amygdala of patients with Parkinson's disease. *Brain* 2002;125:2431–2445. <https://doi.org/10.1093/brain/awf251>
 56. Kramer ML, Schulz-Schaeffer WJ. Presynaptic α -synuclein aggregates, not Lewy bodies, cause neurodegeneration in dementia with lewy bodies. *J Neurosci* 2007;27:1405–1410.
 57. Katsuse O, Iseki E, Marui W, Kosaka K. Developmental stages of cortical Lewy bodies and their relation to axonal transport blockage in brains of patients with dementia with Lewy bodies. *J Neurol Sci* 2003;211:29–35.
 58. Dijkstra AA, Voorn P, Berendse HW, Groenewegen HJ, Rozemuller AJM, van de Berg WDJ. Stage-dependent nigral neuronal loss in incidental Lewy body and parkinson's disease. *Mov Disord* 2014;29:1244–1251. <https://doi.org/10.1002/mds.25952>
 59. Nutt JG, Wooten GF. Diagnosis and initial management of Parkinson's disease. *N Engl J Med* 2005;353:1021–1027. <https://doi.org/10.1056/NEJMc043908>
 60. Brys M, Fanning L, Hung S, Ellenbogen A, Penner N, Yang M, et al. Randomized phase I clinical trial of anti- α -synuclein antibody BIIB054. *Mov Disord* 2019;34:1154–1163. <https://doi.org/10.1002/mds.27738>
 61. Schenk DB, Koller M, Ness DK, Griffith SG, Grundman M, Zago W, et al. First-in-human assessment of PRX002, an anti- α -synuclein monoclonal antibody, in healthy volunteers. *Mov Disord* 2017;32:211–218. <https://doi.org/10.1002/mds.26878>
 62. Jankovic J, Goodman I, Safirstein B, Marmon TK, Schenk DB, Koller M, et al. Safety and tolerability of multiple ascending doses of PRX002/RG7935, an anti- α -Synuclein monoclonal antibody, in patients with Parkinson disease: a randomized clinical trial. *JAMA Neurol* 2018;75:1206–1214. <https://doi.org/10.1001/jamaneurol.2018.1487>
 63. Schofield DJ, Irving L, Calo L, Bogstedt A, Rees G, Nuccitelli A, et al. Preclinical development of a high affinity α -synuclein antibody, MED1341, that can enter the brain, sequester extracellular α -synuclein and attenuate α -synuclein spreading in vivo. *Neurobiol Dis* 2019;132:104582
 64. Zella SMA, Metzdorf J, Ciftci E, Ostendorf F, Muhlack S, Gold R, et al. Emerging immunotherapies for Parkinson disease. *Neurol Ther* 2019;8:29–44. <https://doi.org/10.1007/s40120-018-0122-z>
 65. Brás IC, Dominguez-Mejide A, Gerhardt E, Koss D, Lázaro DF, Santos PI, et al. Synucleinopathies: where we are and where we need to go. *J Neurochem* 2020;153:433–454. <https://doi.org/10.1111/jnc.14965>
 66. Meissner WG, Le Traon AP, Foubert-Samier A, Galabova G, Galitzky M, Kutzelnigg A, et al. A phase 1 randomized trial of specific active α -Synuclein immunotherapies PD01A and PD03A in multiple system atrophy. *Mov Disord* 2020;35:1957–1965. <https://doi.org/10.1002/mds.28218>
 67. Volc D, Poewe W, Kutzelnigg A, Lühns P, Thun-Hohenstein C, Schneeberger A, et al. Safety and immunogenicity of the α -synuclein active immunotherapeutic PD01A in patients with Parkinson's disease: a randomised, single-blinded, phase 1 trial. *Lancet Neurol* 2020;19:591–600.
 68. Poewe W, Volc D, Seppi K, Medori R, Lühns P, Kutzelnigg A, et al. Safety and tolerability of active immunotherapy targeting α -Synuclein with PD03A in patients with early Parkinson's disease: a randomized, placebo-controlled, phase 1 study. *J Parkinsons Dis* 2021;11:1079–1089.
 69. Price DL, Koike MA, Khan A, Wrasidlo W, Rockenstein E, Masliah E, et al. The small molecule alpha-synuclein misfolding inhibitor, NPT200-11, produces multiple benefits in an animal model of Parkinson's disease. *Sci Rep* 2018;8:16165
 70. Heras-Garvin A, Weckbecker D, Ryazanov S, Leonov A, Griesinger C, Giese A, et al. Anle138b modulates α -synuclein oligomerization and prevents motor decline and neurodegeneration in a mouse model of multiple system atrophy. *Mov Disord* 2019;34:255–263.
 71. Levin J, Schmidt F, Boehm C, Prix C, Bötzel K, Ryazanov S, et al. The oligomer modulator anle138b inhibits disease progression in a Parkinson mouse model even with treatment started after disease onset. *Acta Neuropathol* 2014;127:779–780.
 72. Sevigny J, Chiao P, Bussière T, Weinreb PH, Williams L, Maier M, et al. The antibody aducanumab reduces A β plaques in Alzheimer's disease. *Nature* 2016;537:50–56.
 73. Budd Haeblerlein S, O'Gorman J, Chiao P, Bussière T, von Rosenstiel P, Tian Y, et al. Clinical development of Aducanumab, an anti-A β human monoclonal antibody being investigated for the treatment of early Alzheimer's disease. *J Prev Alzheimer's Dis* 2017;4:255–263.
 74. Vaillancourt DE. Aducanumab reduces A β plaques in Alzheimer's disease. *Mov Disord* 2016;31:1631. <https://doi.org/10.1002/mds.26833>

75. Kotzbauer PT, Tu Z, Mach RH. Current status of the development of PET radiotracers for imaging alpha synuclein aggregates in Lewy bodies and Lewy neurites. *Clin Transl* 2017;5:3–14. <https://doi.org/10.1007/s40336-016-0217-4>
76. Xu M, Ryan P, Rudrawar S, Quinn RJ, Zhang H, Mellick GD. Advances in the development of imaging probes and aggregation inhibitors for alpha-synuclein. *Acta Pharmacol Sin* 2020;41:483–498.
77. Korat Š, Bidesi NSR, Bonanno F, di Nanni A, Hoàng ANN, Herfert K, et al. Alpha-Synuclein PET tracer development—an overview about current efforts. *Pharmaceuticals* 2021;14:847.
78. Uzuegbunam BC, Librizzi D, Yousefi BH. PET radiopharmaceuticals for Alzheimer's disease and Parkinson's disease diagnosis, the current and future landscape. *Molecules* 2020;25:977.
79. Fowler DM, Koulou AV, Balch WE, Kelly JW. Functional amyloid - from bacteria to humans. *Trends Biochem* 2007;32:217–224.
80. Serpell LC, Berriman J, Jakes R, Goedert M, Crowther RA. Fiber diffraction of synthetic α -synuclein filaments shows amyloid-like cross- β conformation. *Proc Natl Acad Sci U S A* 2000;97:4897–4902.
81. Berriman J, Serpell LC, Oberg KA, Fink AL, Goedert M, Crowther RA. Tau filaments from human brain and from in vitro assembly of recombinant protein show cross- β structure. *Proc Natl Acad Sci U S A* 2003;100:9034–9038.
82. Ye L, Velasco A, Fraser G, Beach TG, Sue L, Osredkar T, et al. In vitro high affinity α -synuclein binding sites for the amyloid imaging agent PIB are not matched by binding to Lewy bodies in postmortem human brain. *J Neurochem* 2008;105:1428–1437. <https://doi.org/10.1111/j.1471-4159.2008.05245.x>
83. Fodero-Tavoletti MT, Smith DP, McLean CA, Adlard PA, Barnham KJ, Foster LE, et al. In vitro characterization of Pittsburgh compound-B binding to lewy bodies. *J Neurosci* 2007;27:10365–10371.
84. Kudo Y, Okamura N, Furumoto S, Tashiro M, Furukawa K, Maruyama M, et al. 2-(2-[2-Dimethylaminothiazol-5-yl]ethenyl)-6-(2-(fluoro)ethoxy)benzoxazole: a novel PET agent for in vivo detection of dense amyloid plaques in Alzheimer's disease patients. *J Nucl Med* 2007;48:553–561.
85. Fodero-Tavoletti MT, Mulligan RS, Okamura N, Furumoto S, Rowe CC, Kudo Y, et al. In vitro characterisation of BF227 binding to α -synuclein/Lewy bodies. *Eur J Pharmacol* 2009;617:54–58.
86. Verdurand M, Levigoureux E, Lancelot S, Zeinyeh W, Billard T, Quadrio I, et al. Amyloid-beta radiotracer [18 F]BF-227 does not bind to cytoplasmic glial inclusions of postmortem multiple system atrophy brain tissue. *Contrast Media Mol Imaging* 2018;2018:1–7.
87. Kikuchi A, Takeda A, Okamura N, Tashiro M, Hasegawa T, Furumoto S, et al. In vivo visualization of α -synuclein deposition by carbon-11-labelled 2-[2-(2-dimethylaminothiazol-5-yl)ethenyl]-6-[2-(fluoro)ethoxy]benzoxazole positron emission tomography in multiple system atrophy. *Brain* 2010;133:1772–1778. <https://doi.org/10.1093/brain/awq091>
88. Maruyama M, Shimada H, Suhara T, Shinotoh H, Ji B, Maeda J, et al. Imaging of tau pathology in a tauopathy mouse model and in alzheimer patients compared to normal controls. *Neuron* 2013;79:1094–1108.
89. Koga S, Ono M, Sahara N, Higuchi M, Dickson DW. Fluorescence and autoradiographic evaluation of tau PET ligand PBB3 to α -synuclein pathology. *Mov Disord* 2017;32:884–892. <https://doi.org/10.1002/mds.27013>
90. Perez-Soriano A, Arena JE, Dinelle K, Miao Q, McKenzie J, Neilson N, et al. PBB3 imaging in parkinsonian disorders: evidence for binding to tau and other proteins. *Mov Disord* 2017;32:1016–1024. <https://doi.org/10.1002/mds.27029>
91. Miranda-Azpiazu P, Svedberg M, Higuchi M, Ono M, Jia Z, Sunnemark D, et al. Identification and in vitro characterization of C05-01, a PPB3 derivative with improved affinity for alpha-synuclein. *Brain Res* 2020;1749:147131
92. Węgrzynowicz M, Bar-On D, Calo' L, Anichtchik O, Iovino M, Xia J, et al. Depopulation of dense α -synuclein aggregates is associated with rescue of dopamine neuron dysfunction and death in a new Parkinson's disease model. *Acta Neuropathol* 2019;138:575–595. <https://doi.org/10.1007/s00401-019-02023-x>
93. Maurer A, Leonov A, Ryazanov S, Herfert K, Kuebler L, Buss S, et al. 11C radiolabeling of anle253b: a putative PET tracer for Parkinson's disease that binds to α -Synuclein fibrils in vitro and crosses the blood-brain barrier. *ChemMedChem* 2020;15:411–415. <https://doi.org/10.1002/cmdc.201900689>
94. Kuebler L, Buss S, Leonov A, Ryazanov S, Schmidt F, Maurer A, et al. [11C]MODAG-001—towards a PET tracer targeting α -synuclein aggregates. *Eur J Nucl Med Mol Imaging* 2021;48:1759–1772. <https://doi.org/10.1007/s00259-020-05133-x>
95. Cairns AG, Vazquez-Romero A, Moein MM, Ådén J, Elmore CS, Takano A, et al. Increased brain exposure of an alpha-Synuclein fibrillization modulator by utilization of an activated Ester prodrug strategy. *ACS Chem Neurosci* 2018;9:2542–2547. <https://doi.org/10.1021/acscchemneuro.8b00236>
96. Chu W, Zhou D, Gaba V, Liu J, Li S, Peng X, et al. Design, synthesis, and characterization of 3-(Benzylidene)indolin-2-one derivatives as ligands for α -Synuclein fibrils. *J Med Chem* 2015;58:6002–6017. <https://doi.org/10.1021/acs.jmedchem.5b00571>
97. Ferrie JJ, Lengyel-Zhand Z, Janssen B, Lougee MG, Giannakoulis S, Hsieh CJ, et al. Identification of a nanomolar affinity α -synuclein fibril imaging probe by ultra-high throughput: in silico screening. *Chem Sci* 2020;11:12746–12754.
98. Verdurand M, Levigoureux E, Zeinyeh W, Berthier L, Mendjel-Herda M, Cadarossanesaib F, et al. In silico, in vitro, and in vivo evaluation of new candidates for α -Synuclein PET imaging. *Mol Pharm* 2018;15:3153–3166. <https://doi.org/10.1021/acs.molpharmaceut.8b00229>
99. Mathis CA, Lopresti BJ, Ikonovic MD, Klunk WE. Small-molecule PET tracers for imaging proteinopathies. *Semin Nucl Med* 2017;47:553–575.
100. Li JY, Englund E, Holton JL, Soulet D, Hagell P, Lees AJ, et al. Lewy bodies in grafted neurons in subjects with Parkinson's disease suggest host-to-graft disease propagation. *Nat Med* 2008;14:501–503.
101. Desplats P, Lee HJ, Bae EJ, Patrick C, Rockenstein E, Crews L, et al. Inclusion formation and neuronal cell death through neuron-to-neuron transmission of α -synuclein. *Proc Natl Acad Sci U S A* 2009;106:13010–13015.
102. Grozdanov V, Danzer KM. Release and uptake of pathologic alpha-synuclein. *Cell Tissue Res* 2018;373:175–182. <https://doi.org/10.1007/s00441-017-2775-9>
103. Bagchi DP, Yu L, Perlmutter JS, Xu J, Mach RH, Tu Z, et al. Binding of the radioligand SIL23 to α -Synuclein fibrils in Parkinson disease brain tissue establishes feasibility and screening approaches for developing a Parkinson disease imaging agent. *PLoS One* 2013;8:e55031. <https://doi.org/10.1371/journal.pone.0055031>
104. Heiss WD, Herholz K. Brain receptor imaging. *J Nucl Med* 2006;47:302–312.
105. Kövari E, Horvath J, Bouras C. Neuropathology of Lewy body disorders. *Brain Res Bull* 2009;80:203–210.
106. Kotzbauer PT, Cairns NJ, Campbell MC, Willis AW, Racette BA, Tabbal SD, et al. Pathologic accumulation of α -synuclein and A β in Parkinson disease patients with dementia. *Arch Neurol* 2012;69:1326–1331. <https://doi.org/10.1001/archneurol.2012.1608>
107. Donaghy P, Thomas AJ, O'Brien JT. Amyloid PET imaging in lewy body disorders. *Am J Geriatr Psychiatry* 2015;23:23–37.
108. Hsieh CJ, Ferrie JJ, Xu K, Lee I, Graham TJA, Tu Z, et al. Alpha Synuclein fibrils contain multiple binding sites for small molecules. *ACS Chem Neurosci* 2018;9:2521–2527. <https://doi.org/10.1021/acscchemneuro.8b00177>
109. Paleologou KE, Kragh CL, Mann DMA, Salem SA, Al-Shami R, Allsop D, et al. Detection of elevated levels of soluble α -synuclein oligomers in post-mortem brain extracts from patients with dementia with Lewy bodies. *Brain* 2009;132:1093–1101. <https://doi.org/10.1093/brain/awn349>
110. Sharon R, Bar-Joseph I, Frosch MP, Walsh DM, Hamilton JA, Selkoe DJ. The formation of highly soluble oligomers of α -synuclein is regulated by fatty acids and enhanced in Parkinson's disease. *Neuron* 2003;37:583–595.

111. Campbell BCV, McLean CA, Culvenor JG, Gai WP, Blumbergs PC, Jäkälä P, et al. The solubility of α -synuclein in multiple system atrophy differs from that of dementia with Lewy bodies and Parkinson's disease. *J Neurochem* 2001;76:87–96. <https://doi.org/10.1046/j.1471-4159.2001.00021.x>
112. Tuttle MD, Comellas G, Nieuwkoop AJ, Covell DJ, Berthold DA, Kloepper KD, et al. Solid-state NMR structure of a pathogenic fibril of full-length human α -synuclein. *Nat Struct Mol Biol* 2016; 23:409–415.
113. Guerrero-Ferreira R, Taylor NMI, Mona D, Ringler P, Lauer ME, Riek R, et al. Cryo-EM structure of alpha-synuclein fibrils. *Elife*. eLife 2018;7:e36402.
114. Li B, Ge P, Murray KA, Sheth P, Zhang M, Nair G, et al. Cryo-EM of full-length α -synuclein reveals fibril polymorphs with a common structural kernel. *Nat Commun* 2018;9:1–10.
115. Li Y, Zhao C, Luo F, Liu Z, Gui X, Luo Z, et al. Amyloid fibril structure of α -synuclein determined by cryo-electron microscopy. *Cell Res* 2018;28:897–903.
116. Schweighauser M, Shi Y, Tarutani A, Kametani F, Murzin AG, Ghetti B, et al. Structures of α -synuclein filaments from multiple system atrophy. *Nature* 2020;585:464–469.
117. Yu L, Cui J, Padakanti PK, Engel L, Bagchi DP, Kotzbauer PT, et al. Synthesis and in vitro evaluation of α -synuclein ligands. *Bioorg Med Chem* 2012;20:4625–4634.
118. Chesselet MF, Richter F, Zhu C, Magen I, Watson MB, Subramaniam SR. A progressive mouse model of Parkinson's disease: the Thy1-aSyn ("line 61") mice. *Neurotherapeutics* 2012;9: 297–314. <https://doi.org/10.1007/s13311-012-0104-2>
119. Giasson BI, Duda JE, Quinn SM, Zhang B, Trojanowski JQ, Lee VMY. Neuronal α -synucleinopathy with severe movement disorder in mice expressing A53T human α -synuclein. *Neuron* 2002; 34:521–533. <https://doi.org/10.1016/S0896-6273>
120. Freichel C, Neumann M, Ballard T, Müller V, Woolley M, Ozmen L, et al. Age-dependent cognitive decline and amygdala pathology in α -synuclein transgenic mice. *Neurobiol Aging* 2007; 28:1421–1435.
121. Luk KC, Kehm V, Carroll J, Zhang B, O'Brien P, Trojanowski JQ, et al. Pathological α -synuclein transmission initiates Parkinson-like neurodegeneration in nontransgenic mice. *Science* 2012;338: 949–953.
122. Peelaerts W, Bousset L, Van der Perren A, Moskalyuk A, Pulizzi R, Giugliano M, et al. α -Synuclein strains cause distinct synucleinopathies after local and systemic administration. *Nature* 2015;522:340–344.
123. Paumier KL, Luk KC, Manfredsson FP, Kanaan NM, Lipton JW, Collier TJ, et al. Intrastriatal injection of pre-formed mouse α -synuclein fibrils into rats triggers α -synuclein pathology and bilateral nigrostriatal degeneration. *Neurobiol Dis* 2015;82: 185–199.
124. Bourdenx M, Dovero S, Engeln M, Bido S, Bastide MF, Duthiel N, et al. Lack of additive role of ageing in nigrostriatal neurodegeneration triggered by α -synuclein overexpression. *Acta Neuropathol Commun* 2015;3:46. <https://doi.org/10.1186/s40478-015-0222-2>
125. van der Perren A, Toelen J, Casteels C, Macchi F, van Rompuy AS, Sarre S, et al. Longitudinal follow-up and characterization of a robust rat model for Parkinson's disease based on overexpression of alpha-synuclein with adeno-associated viral vectors. *Neurobiol Aging* 2015;36:1543–1558.
126. Gombash SE, Manfredsson FP, Kemp CJ, Kuhn NC, Fleming SM, Egan AE, et al. Morphological and behavioral impact of AAV2/5-mediated overexpression of human wildtype alpha-synuclein in the rat nigrostriatal system. *PLoS One* 2013;8:e81426. <https://doi.org/10.1371/journal.pone.0081426>
127. Luk KC, Kehm VM, Zhang B, O'Brien P, Trojanowski JQ, Lee VMY. Intracerebral inoculation of pathological α -synuclein initiates a rapidly progressive neurodegenerative α -synucleinopathy in mice. *J Exp Med* 2012;209:975–986.
128. Watts JC, Giles K, Oehler A, Middleton L, Dexter DT, Gentleman SM, et al. Transmission of multiple system atrophy prions to transgenic mice. *Proc Natl Acad Sci U S A* 2013;110: 19555–19560.
129. Lau A, So RWL, Lau HHC, Sang JC, Ruiz-Riquelme A, Fleck SC, et al. α -Synuclein strains target distinct brain regions and cell types. *Nat Neurosci* 2020;23:21–31.
130. Thakur P, Breger LS, Lundblad M, Wan OW, Mattsson B, Luk KC, et al. Modeling Parkinson's disease pathology by combination of fibril seeds and α -synuclein overexpression in the rat brain. *Proc Natl Acad Sci U S A* 2017;114:E8284–E8293. <https://doi.org/10.1073/pnas.1710442114>
131. Espa E, Clemensson EKH, Luk KC, Heuer A, Björklund T, Cenci MA. Seeding of protein aggregation causes cognitive impairment in rat model of cortical synucleinopathy. *Mov Disord* 2019; 34:1699–1710. <https://doi.org/10.1002/mds.27810>
132. Kaplon H, Reichert JM. Antibodies to watch in 2021. *MAbs* 2021; 13:1860476.
133. Sehlin D, Fang XT, Cato L, Antoni G, Lannfelt L, Syvänen S. Antibody-based PET imaging of amyloid beta in mouse models of Alzheimer's disease. *Nat Commun* 2016;7:1–11.
134. Syvänen S, Fang XT, Hultqvist G, Meier SR, Lannfelt L, Sehlin D. A bispecific Tribody PET radioligand for visualization of amyloid-beta protofibrils—a new concept for neuroimaging. *Neuroimage* 2017;148:55–63.
135. Sehlin D, Fang XT, Meier SR, Jansson M, Syvänen S. Pharmacokinetics, biodistribution and brain retention of a bispecific antibody-based PET radioligand for imaging of amyloid- β . *Sci Rep* 2017;7: 17254.
136. Syvänen S, Hultqvist G, Gustavsson T, Gumucio A, Laudon H, Söderberg L, et al. Efficient clearance of A β protofibrils in A β PP-transgenic mice treated with a brain-penetrating bifunctional antibody. *Alzheimer's Res Ther* 2018;10:49. <https://doi.org/10.1186/s13195-018-0377-8>
137. Poduslo JF, Curran GL, Berg CT. Macromolecular permeability across the blood-nerve and blood-brain barriers. *Proc Natl Acad Sci U S A* 1994;91:5705–5709.
138. Bard F, Cannon C, Barbour R, Burke RL, Games D, Grajeda H, et al. Peripherally administered antibodies against amyloid β -peptide enter the central nervous system and reduce pathology in a mouse model of Alzheimer disease. *Nat Med* 2000;6: 916–919.
139. Hultqvist G, Syvänen S, Fang XT, Lannfelt L, Sehlin D. Bivalent brain shuttle increases antibody uptake by monovalent binding to the transferrin receptor. *Theranostics* 2017;7:308–318.
140. Niewoehner J, Bohrmann B, Collin L, Urich E, Sade H, Maier P, et al. Increased brain penetration and potency of a therapeutic antibody using a monovalent molecular shuttle. *Neuron* 2014;81: 49–60.
141. Dasgupta A, Liu M, Ojha T, Storm G, Kiessling F, Lammers T. Ultrasound-mediated drug delivery to the brain: principles, progress and prospects. *Drug Discov Today Technol* 2016;20: 41–48.
142. Du M, Chen Z, Chen Y, Li Y. Ultrasound-targeted delivery technology: a novel strategy for tumor-targeted therapy. *Curr Drug Targets* 2018;20:220–231.
143. Liu HL, Hsu PH, Lin CY, Huang CW, Chai WY, Chu PC, et al. Focused ultrasound enhances central nervous system delivery of bevacizumab for malignant glioma treatment. *Radiology* 2016; 281:99–108. <https://doi.org/10.1148/radiol.2016152444>
144. Yang FY, Chang WY, Li JJ, Wang HE, Chen JC, Chang CW. Pharmacokinetic analysis and uptake of 18f-fbpa-fr after ultrasound-induced blood-brain barrier disruption for potential enhancement of boron delivery for neutron capture therapy. *J Nucl Med* 2014;55:616–621.
145. Börjesson PKE, Jauw YWS, De Bree R, Roos JC, Castelijns JA, Leemans CR, et al. Radiation dosimetry of 89Zr-labeled chimeric monoclonal antibody U36 as used for immuno-PET in head and neck cancer patients. *J Nucl Med* 2009;50:1828–1836.
146. Cherry SR, Jones T, Karp JS, Qi J, Moses WW, Badawi RD. Total-body PET: maximizing sensitivity to create new opportunities for clinical research and patient care. *J Nucl Med* 2018;59: 3–12.

147. Altai M, Membreno R, Cook B, Tolmachev V, Zeglis BM. Pretargeted imaging and therapy. *J Nucl Med* 2017;58:1553–1559.
148. Lopes van den Broek S, Shalgunov V, Sehlin D, Syvanen S, Herth M. Development of trans—Cyclooctene modified antibodies for Pretargeted imaging within the central nervous system. *J Nucl Med* 2020;61:196–196.
149. Sehlin D, Syvänen S, Ballanger B, Barthel H, Bischof GN, Boche D, et al. Engineered antibodies: new possibilities for brain PET? *Eur J Nucl Med Mol Imaging* 2019;46:2848–2858. <https://doi.org/10.1007/s00259-019-04426-0>
150. van Dongen GAMS, Beaino W, Windhorst AD, Zwezerijnen GJC, Oprea-Lager DE, Hendrikse NH, et al. The role of 89Zr-Immuno-PET in navigating and Derisking the development of bio-pharmaceuticals. *J Nucl Med* 2021;62:438–445.
151. Danzer KM, Haasen D, Karow AR, Moussaud S, Habeck M, Giese A, et al. Different species of α -synuclein oligomers induce calcium influx and seeding. *J Neurosci* 2007;27:9220–9232.
152. Danzer KM, Krebs SK, Wolff M, Birk G, Hengerer B. Seeding induced by α -synuclein oligomers provides evidence for spreading of α -synuclein pathology. *J Neurochem* 2009;111:192–203. <https://doi.org/10.1111/j.1471-4159.2009.06324.x>
153. Mougnot AL, Nicot S, Bencsik A, Morignat E, Verchère J, Lakhdar L, et al. Prion-like acceleration of a synucleinopathy in a transgenic mouse model. *Neurobiol Aging* 2012;33:2225–2228.
154. Meier SR, Sehlin D, Roshanbin S, Lim Falk V, Saito T, Saido TC, et al. 11 C-PIB and 124 I-antibody PET provide differing estimates of brain amyloid-beta after therapeutic intervention. *J Nucl Med* 2021;63:302–309.