

1 **Keep and promote biodiversity at polluted sites under**
2 **phytomanagement**

3 *Carlos Garbisu^{1*}, Itziar Alkorta², Petra Kidd³, Lur Epelde¹, Michel Mench⁴*

4

5 ¹Department of Conservation of Natural Resources, Soil Microbial Ecology Group, NEIKER-Basque
6 Institute for Agricultural Research and Development, Basque Research and Technology Alliance
7 (BRTA), Parque Científico y Tecnológico de Bizkaia, P812, 48160 Derio, Spain

8 ²Department of Biochemistry and Molecular Biology, University of the Basque Country, P.O. Box 644,
9 48080 Bilbao, Spain

10 ³Instituto de Investigaciones Agrobiológicas de Galicia (IIAG), Consejo Superior de Investigaciones
11 Científicas (CSIC), 15780 Santiago de Compostela, Spain

12 ⁴Univ. Bordeaux, INRAE, BIOGECO, F-33615 Pessac, France

13

14 ***Correspondence**

15 Dr. Carlos Garbisu

16 NEIKER, Department of Conservation of Natural Resources

17 Parque Científico y Tecnológico de Bizkaia P812

18 E-48160 Derio, SPAIN

19 Phone: +34 94 4034300; Fax: +34 94 4034310; E-mail: cgarbisu@neiker.eus

20

21 **Acknowledgements**

22 This work was supported by the Ministry of Economy, Industry and Competitiveness, Spanish
23 Government (AGL2016-76592-R) and European Union within the Interreg SUDOE (PhytoSUDOE-
24 SOE1/P5/E0189).

25

26 **Abstract**

27 The phytomanagement concept combines a sustainable reduction of pollutant linkages at risk-assessed
28 contaminated sites with the generation of both valuable biomass for the (bio)economy and ecosystem
29 services. One of the potential benefits of phytomanagement is the possibility to increase biodiversity in
30 polluted sites. However, the unique biodiversity present in some polluted sites can be severely impacted
31 by the implementation of phytomanagement practices, even resulting in the local extinction of endemic
32 ecotypes or species of great conservation value. Here we highlight the importance of promoting measures
33 to minimize the potential adverse impact of phytomanagement on biodiversity at polluted sites, as well as
34 recommend practices to increase biodiversity at phytomanaged sites without compromising its
35 effectiveness in terms of reduction of pollutant linkages and the generation of valuable biomass and
36 ecosystem services.

37

38 **Keywords:** contaminated soil; metal; metallophytes; phytoremediation; trace elements.

39

40 **1. Introduction**

41 The notion of phytomanagement is based on the combination of (i) a sustainable reduction of pollutant
42 linkages at degraded sites with (ii) the generation of valuable products and essential ecosystem services.
43 In other words, its main purpose is to grow profitable plants to minimize pollutant-induced environmental
44 risks while maximizing economic and/or ecological revenues. It is often claimed that one of the potential
45 benefits of phytomanagement is the possibility to enhance biodiversity in the degraded site under
46 recovery. Pertinently, it must be strongly emphasized that some polluted sites, most relevantly mining
47 sites, can harbour a unique biodiversity that must be carefully preserved. In any event, protecting
48 biodiversity is of the utmost importance as human well-being depends upon biodiversity in many
49 different ways (Naeem et al., 2016). In consequence, under the current scenario of global change and
50 biodiversity loss, it is crucial to use as many tools as possible to preserve the fabric of life and the natural
51 capital on which our survival and well-being depend. Biodiversity is known to be critical for the supply of
52 ecosystem services and, then, it is not surprising that much research effort has been directed at
53 understanding how biodiversity impacts ecosystem functioning and resilience, and concomitantly the
54 sustainable provision of goods and ecosystems services. This aspect has special relevance within the
55 phytomanagement framework since, as described above, the main purpose of phytomanagement is to
56 grow profitable plants in order to minimize pollutant-induced environmental risks while maximizing
57 economic and/or ecological revenues in terms of products and ecosystem services. However, when
58 implementing actions to promote such biodiversity in phytomanaged sites, in most cases, the only
59 initiative is to enhance the number of different plant species grown for phytomanagement purposes. We
60 must overcome such incomplete approach by widening our understanding of how the different taxonomic
61 groups can be positively or negatively affected by phytomanagement practices. In addition, the unique
62 biodiversity present in some polluted sites can be negatively affected by the implementation of
63 phytomanagement practices. In this review paper, the importance of promoting (i) measures to minimize
64 the potential adverse impact of phytomanagement on biodiversity; and (ii) practices to increase
65 biodiversity at phytomanaged sites, is highlighted.

66

67 **2. Phytomanagement: a sustainable gentle remediation option**

68 As a result of a wide variety of anthropogenic activities and accidental spills, many soils are currently
69 polluted with a myriad of potentially toxic compounds, such as trace elements (TEs), mineral oils,

70 polycyclic aromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs), pesticides, etc.
71 Unfortunately, the remediation of polluted soils is often a very expensive, environmentally-disruptive
72 activity, especially at large sites and/or in those soils simultaneously polluted with several contaminants
73 inducing adverse effects on biological receptors (Agnello et al. 2016).

74 Opportunely, in the last decades, various Gentle Remediation Options (GROs) have been
75 developed as more cost-effective, environmentally-friendly and aesthetically-pleasing technologies for
76 the remediation of large areas with polluted soils from mild up to medium levels of contamination
77 (Vangronsveld et al. 2009; Kidd et al. 2015; Mench et al. 2018). Among them, phytoremediation and
78 phytomanagement have shown their great potential, on the long term, for the sustainable remediation of
79 polluted sites due to their capacity to combine an effective mitigation of pollutant-induced risks with the
80 provision of valuable plant biomass and ecosystem services (Mench et al. 2018).

81 The term *phytoremediation* refers to a set of sustainable phytotechnologies focused on the use of
82 plant species to remediate polluted sites, mainly those affected by the presence of TEs via the
83 phytoextraction or phytostabilization options, which aim at (i) decreasing the available soil TEs, through
84 plant uptake and accumulation in the harvestable plant parts, or (ii) reducing the labile (“bioavailable”) TE
85 pool usually by combining the growth of TE-excluding plants with the application of soil amendments
86 (Garbisu and Alkorta 2001; Alkorta et al. 2004a,b). However, the commercial application of
87 phytoextraction has been seriously hampered by its intrinsic limitations, *e.g.* the long time required to
88 effectively extract TEs from medium and highly polluted soils, root depth, lack of plants that can
89 accumulate more than one or two TEs, decrease of metal(loid) market prices, etc. In turn, one constraint
90 for the application of phytostabilization is that many risk-assessment regulations for soil remediation are
91 still based on total soil TEs, not on their bioavailable concentrations or site-specific risk assessment.
92 Paradoxically, the harmful effects of TEs on soil biota and, hence, soil health, are related to the
93 sensitivity/tolerance of living organism populations and the bioavailable pool rather than total metal(loid)
94 concentrations (Kumpiene et al. 2009, 2017), the bioavailable fraction being subject to uptake by soil
95 organisms, leaching, and transfer to other environmental media (Madejón et al. 2006).

96 In any event, for effective clean-up of TE-polluted soils, the combination of different
97 approaches, *e.g.* phytoextraction with hyperaccumulators, chelate-assisted phytoextraction,
98 phytostabilization, microbe-assisted phytoremediation (bioaugmentation), traditional breeding and/or

99 genetic engineering of phytoremediation plants, etc., appears necessary to increase the applicability of
100 phytoremediation in the future (Yang et al. 2020a).

101 To overcome the phytoremediation limitations, the concept of *phytomanagement* evolved (Fig.
102 1), combining a sustainable reduction in pollutant linkages with the generation of plant biomass (mainly
103 non-food crops) and ecosystem services (Mench et al. 2009; Robinson et al. 2009; Fässler et al. 2010;
104 Robinson and McIvor 2013; Cundy et al. 2016; Burges et al. 2018). The phytomanagement objective is to
105 grow profitable plants to control the bioavailable pool of soil pollutants (*e.g.*, TEs), thereby minimizing
106 environmental risks, while maximizing economic and ecological revenues (Vangronsveld et al. 2009). In
107 this way, phytomanagement (commonly based on the interactions among plants, microorganisms and soil
108 amendments) is often considered a “holding strategy” for vacant sites until their remediation is
109 undertaken according to future land use (Mench et al. 2018). Most importantly, compared to other
110 remediation technologies, the requirements of phytomanagement for chemicals and energy are much
111 lower, as well as the total cost, making it a viable strategy for the remediation of large polluted areas
112 (Thewys et al. 2010; Kuppens et al. 2015; Giagnoni et al. 2020).

113 The production of valuable plant biomass (for timber, bioenergy, biofortified products,
114 ecomaterials, etc.) is considered essential for the commercial success of phytomanagement (Conesa et al.
115 2012). Energy crops, *e.g.* *Miscanthus* spp., *Ricinus communis* L. and *Brassica napus* L., can be grown for
116 biofuel production (Burges et al. 2018). Other plant species can be grown for the production of biochar,
117 raw materials (oil, paper, chemicals, essential oils, etc.), medicinal purposes, etc. (Pandey et al. 2016).
118 Likewise, the growth of fast growing trees opens the possibility to phytoextract some metals in excess
119 (*e.g.*, Cd, Zn, Ni, U, Cs, and Sr) while producing biomass for bioenergy and products (*e.g.*, timber, resin,
120 adhesives, etc.). Other phytomanagement options are aimed at removing the bioavailable metal(loid)
121 fraction, the so-called “bioavailable contaminant stripping (BCS)”, while providing ecosystem services
122 and feedstocks for biomass-processing (bio)technologies (Herzig et al. 2014).

123 Since the 2010s, an increasing attention is developing on the capacity of phytomanagement
124 options to provide a wide variety of co-benefits and ecosystem services, such as primary production,
125 control of soil erosion, water runoff/drainage management, carbon sequestration, amenity and recreation,
126 aesthetic value, habitat for animals and microorganisms, biodiversity, etc. (Evangelou et al. 2015; Kidd et
127 al. 2015; Cundy et al. 2016; Simek et al. 2017; Touceda-González et al. 2017a; Xue et al. 2018). Strictly
128 speaking, biodiversity *per se* is not an ecosystem service (Haines-Young and Potschin 2010); rather,

129 biodiversity supports the flow of vital ecosystem services that we depend upon (in other words,
130 biodiversity forms the biological infrastructure that supports the provision of ecosystem services). Indeed,
131 the ecosystem services (and, concomitantly, human well-being) depend essentially on the structures and
132 processes generated by living organisms and their interactions with, and processing of, abiotic materials
133 (Haines-Young and Potschin 2010; IPBES 2019). Although biodiversity has intrinsic value by itself (and,
134 then, it should be preserved in its own right), its utilitarian value has increasingly become the central
135 focus of the debates on the need to preserve our natural capital (Chang et al. 2007; Haines and Potschin
136 2010).

137 The phytomanagement capacity to promote biodiversity in polluted sites is of key importance, as
138 we are currently at a crucial juncture in human history, with biodiversity being lost at an accelerating pace
139 due to an increasingly affluent human population, climate change, uncontrolled development and habitat
140 destruction (Sandifer et al. 2015; IPBES 2019). Taking into consideration the links between biodiversity,
141 ecosystem functioning, ecosystem services and human well-being (Cardinale et al. 2006; Naeem et al.
142 2016), now more than ever, the importance of promoting biodiversity must be emphasized.

143

144 **3. Unique biodiversity at polluted sites**

145 One of the recurrently mentioned potential benefits of phytomanagement is the possibility to increase
146 biodiversity in the polluted land in question. However, the unique biodiversity often present in many
147 polluted sites (in particular, long-term abandoned mine sites) can be severely impacted by the
148 implementation of phytomanagement practices, even resulting in the local extinction of endemic ecotypes
149 or species of great conservation value. Here we highlight the importance of promoting measures to
150 minimize the potential adverse impact of phytomanagement on biodiversity at polluted sites. After all,
151 some polluted sites, most relevantly mining sites, harbour a unique biodiversity that must be
152 painstakingly preserved. Actually derelict soils can provide an interesting biodiversity for a variety of
153 uses (Vincent et al. 2018).

154 In particular, there is a need to conserve metallophytes (*i.e.*, unique plant species that have
155 evolved to survive on soils with high TE levels), which are nowadays increasingly under threat of
156 extinction from mining activity (Whiting et al. 2004; Batty 2005; Baker et al. 2010; Paul et al. 2018).
157 Indeed, regrettably, metalliferous ecosystems are presently threatened at a global scale by the growth of

158 mining activities with concomitant extinction risks for metallophyte diversity (Whiting et al. 2004; Séleck
159 et al. 2013).

160 Metallophytes are the consequence of powerful selective pressures over long evolutionary times
161 as a result of the presence of high total soil TEs (Ginocchio and Baker 2004). The intensity and duration
162 of the sustained evolutionary exposure to these high TE levels direct the degree of specialization of the
163 TE resistance trait. Thus, some populations of plant species can evolve TE resistance within a few years,
164 for example around metal smelters, if the selection pressure is high enough (Barrutia et al. 2011a).
165 Populations of pseudometallophytes present a greater capacity to withstand phytotoxicity induced by TE
166 excess, as compared with other populations of the same plant species from non-polluted sites (Whiting et
167 al. 2004; Barrutia et al. 2011a). But, as the duration of TE exposure increases, the mechanisms that allow
168 survival and growth in the presence of high TE levels become gradually more specialized, resulting in
169 true metallophytes or eumetallophytes that have developed evolutionary mechanisms to live and thrive on
170 metalliferous soils. As a matter of fact, true metallophytes have often diverged genetically and
171 morphologically to form new taxa endemic to their native metalliferous soils (Barrutia et al. 2011a).
172 Regrettably, their restricted geographic range is, partly, responsible for the current high rates of
173 population decline or, what is worse, irreversible extinction.

174 Plants growing in TE-polluted sites can be classified as (1) *excluders*: these plants limit TE
175 uptake and translocation and, then, maintain low TE concentrations in their aerial tissues; (2) *indicators*:
176 these plants accumulate TEs in their harvestable parts at concentrations similar to those present in the
177 polluted soil; and (3) *accumulators/hyperaccumulators*: these plants increase TE uptake, translocation
178 and accumulation in their aboveground biomass reaching levels that far exceed those present in the
179 polluted soil (van der Ent et al. 2013, 2015a; Malik et al. 2017; Massoura et al. 2014; Reeves et al.
180 2017a). Particularly, hyperaccumulators are exceptional plants that accumulate metal(loid)s in their
181 tissues to levels that can be hundreds or thousands of times greater than common ranges in other plant
182 species (van der Ent et al. 2013), and whose ecology is an active field of research, focusing on anti-
183 herbivore defences, allelopathy and biotic interactions (Reeves et al. 2017b). Logically, it is important to
184 protect not only TE hyperaccumulators as nature's oddities but also TE excluders, indicators and
185 accumulators.

186 Apart from their intrinsic value as remarkable rare species, metallophytes are suitable candidates
187 for the revegetation of mining sites, as well as for the implementation of phytoremediation

188 (phytoextraction/phytomining, phytostabilization) and phytomanagement initiatives (van der Ent et al.
189 2015b; Rosenkranz et al. 2019; Corzo Remigio et al. 2020). Thus, TE excluders and hyperaccumulators
190 have been extensively used for phytostabilization and phytoextraction purposes, respectively (Hernández-
191 Allica et al. 2006; Epelde et al. 2008, 2009, 2010; Barrutia et al. 2009; Pardo et al. 2014; Garaiyurrebaso
192 et al. 2017). There is nowadays an increasing interest in the use of native plant species and populations
193 for the revegetation of TE polluted sites, as opposed to non-native, introduced species (Parraga-Aguado et
194 al. 2014; Chen et al. 2019). In this respect, a sturdy commitment to conservation of metallophyte
195 biodiversity is self-evident (Whiting et al. 2002).

196 Then, before starting any phytomanagement initiative, it is imperative to study the native
197 vegetation of the polluted site in search of potential candidates (*e.g.*, metallophytes) for conservation
198 purposes. If such candidates are identified, then, an area of the site (preferably, the area where the most
199 interesting plant species have been identified) must be left unmanaged for conservation purposes (and, if
200 needed, protection barriers must be installed).

201 In addition to protecting the natural environment of valued and treasured plant species (*i.e.*, *in*
202 *situ* conservation in biotope “islands”), efforts must also be directed at conserving them *ex situ*, that is to
203 say, in germplasm banks, seed gardens, arboreta, botanic gardens, etc. (Whiting et al. 2004), ideally
204 maintaining the required degree of contaminant exposure (otherwise, contaminant-sensitive, non-adapted
205 individuals might again become dominant in the plant population after a few cultivations). Additionally,
206 when designing a strategy to preserve the valuable native vegetation at the polluted site, attention should
207 also be paid to plant assemblages (*e.g.*, metalliferous distinctive plant communities).

208 Apart from the presence of unique metallophytes in TE-polluted sites, these degraded
209 environments can also harbour a valuable microbial diversity that can likewise be used for
210 phytoremediation and/or phytomanagement initiatives. For instance, TE resistant plant growth-promoting
211 rhizobacteria and endophytes, as well as TE-resistant mycorrhiza, can be isolated from TE-polluted sites
212 and, subsequently, be used to improve plant survival, growth and performance under the harsh conditions
213 usually present in many TE-polluted sites, particularly mining sites (Weyens et al. 2011; Ma et al. 2015;
214 Burges et al. 2016, 2017; Harrison and Griffin 2020). For instance, rhizobacterial inoculants (*e.g.*,
215 *Arthrobacter nicotinovorans* SA40) have been shown to improve nickel phytoextraction by the
216 hyperaccumulator *Alyssum pintodasilvae* (Cabello-Conejo et al. 2014). Similarly, the inoculation of
217 ultramafic soils with *Microbacterium arabinogalactanolyticum* AY509224 increased soil Ni extractability

218 and uptake by *Alyssum murale* (Abou-Shanab et al. 2006). The inoculation with TE-resistant plant
219 growth-promoting bacteria has been reported to enhance the biomass of different plant species (e.g.,
220 *Brassica juncea*, *Ricinus communis*, *Helianthus annuus*, and *Sedum alfredii*) growing in TE-polluted soils
221 (Dell'Amico et al. 2008; Jiang et al. 2008; Mastretta et al. 2009; Zaidi et al. 2006). Likewise, Kolbas et al.
222 (2015) reported the positive effects of endophytic bacteria for Cu phytoextraction by sunflower plants.
223 Truyens et al. (2015) found that inoculation of *Agrostis capillaris* plants with endophytes can be
224 beneficial for their establishment during phytoextraction and phytostabilisation of Cd polluted soils. The
225 co-inoculation of *Paenibacillus mucilaginosus* and *Sinorhizobium meliloti* in Cu-contaminated soil
226 planted with *Medicago sativa* improved alfalfa growth and decreased Cu accumulation in shoots,
227 compared to the uninoculated control (Ju et al. 2020). The inoculation of *Pseudomonas vancouverensis*
228 promoted As accumulation efficiency in *Pteris vittata* and *Pteris multifida* (Yang et al. 2020b). However,
229 the effect of bacterial inoculants on plant growth and TE-accumulation has been shown to be plant
230 species-specific (Becerra-Castro et al. 2012).

231 Apart from the recognized conservation value of metallophytes present at TE-polluted sites,
232 plant species and, interestingly, microbial (bacteria, fungi) populations living in soils polluted with
233 organic compounds must also be considered for conservation purposes, owing to their intrinsic value as
234 well as their potential use for the rhizoremediation (i.e., degradation of pollutants by rhizosphere bacteria)
235 (Barrutia et al. 2011b; Lacalle et al. 2018; Brereton et al. 2020) and bioremediation of organically
236 polluted soils (Garbisu et al. 2017; Anza et al. 2018; Megloulou et al. 2019; Villaverde et al. 2019). Since
237 bacterial and fungal strains isolated from organically-polluted soils can then be used for bioremediation
238 via bioaugmentation purposes, after thoroughly testing their degrading capabilities and potentials, it is
239 recommended to keep them in a microbial bank for possible future biotechnological applications.

240

241 **4. Managing biodiversity during phytomanagement**

242 ***4.1. Phytomanagement under the current scenario of climate change***

243 The negative consequences of climate change can nowadays be undoubtedly identified in the more
244 frequent alteration of natural and agricultural ecosystems, owing to, for instance, higher temperatures,
245 extreme droughts and storms, and an increased likelihood of heat waves and heavy precipitation episodes
246 (Alkorta et al. 2017). Not surprisingly, plant survival and growth are being significantly altered under

247 changing climatic conditions. Furthermore, increasing CO₂ concentrations in the atmosphere are currently
248 changing the physiology of plants, affecting, among other aspects, their growth rate.

249 Specifically, regarding the choice of plant species for phytomanagement in semi-arid and arid
250 regions (*e.g.*, southern Europe) (Pulighe et al. 2019), and taking into account the critical importance of an
251 adequate water regime for the success of revegetation programs, special attention should be paid to the
252 selection of drought-resistant plant species and ecotypes, since the duration and frequency of extreme
253 droughts is nowadays increasing in many semi-arid and arid regions (Risueño et al. 2020).

254 The possibility of irrigating phytomanagement crops is decidedly controversial, since water is an
255 increasingly scarce resource in many parts of the world. A proper sustainable management of water
256 resources is currently one of the greatest challenges for our society worldwide. Above all, we must first
257 ensure availability of good quality water for human consumption and agricultural purposes. Regrettably,
258 in the coming decades, the problem of water scarcity will probably get worse than it is now. Predictably,
259 an increase in the world human population will imply more water for human consumption and
260 agricultural production (agriculture accounts for around 70% of the water currently used in the world).
261 Then, it follows that the consumption of good quality water for irrigation of phytomanaged sites is, in
262 general, not considered a valid option, especially in semi-arid and arid regions. An alternative is the use
263 of wastewater for irrigation. An appealing, and currently attention-grabbing, option is the possibility of
264 treating such wastewater by means of rhizofiltration (*i.e.*, the use of plant roots and associated microbes to
265 absorb, concentrate and precipitate pollutants, especially TEs, from polluted effluents and waters) and/or
266 biodegradation, notably using constructed wetlands (CWs) and floating islands (Dushenkov et al. 1995;
267 Schröder et al. 2007; Zhang et al. 2007; Olguin et al. 2017).

268 Urban wastewater is known to contain nitrogen, phosphorus and other nutrients, leading to an
269 extra beneficial effect for plant growth through fertilization. Irrigation with wastewater is only
270 recommended for non-food and non-fodder crops, and then it would be an ideal option for
271 phytomanagement. Evidently, it would be beneficial to have an efficient urban wastewater treatment plant
272 closed to the site to be phytomanaged, so that the wastewater, directly or preferably after rhizofiltration in
273 a CW does not need to be transported a long distance. Interestingly, CWs can also be used to treat acid
274 mine drainage and then there is the possibility of reusing the treated water to eventually irrigate mine
275 tailings (Pat-Espadas et al. 2018).

276 Therefore, especially in semi-arid and arid regions of the world, for phytomanagement purposes,
277 it is recommended to select plant species that are resistant to water stress, extreme droughts and heat
278 waves for increasing the long-term success of the phytomanagement strategy (Risueño et al. 2020). For
279 instance, as water supply and its distribution during the crop cycle is a key limiting factor for crop
280 production in SW France, the sunflower and tobacco ability to stand more frequent heat waves and long
281 droughts is certainly an advantage (Kidd et al. 2015; Mench et al. 2018). Although hundreds of plant
282 species are suitable candidates for phytoremediation and/or phytomanagement purposes, there is
283 nowadays an urgent need to identify those which can successfully be used under the current scenario of
284 climate change.

285 The potential indirect effects of climate change on the soil biota present in phytomanaged sites
286 must be also considered, through the abovementioned climate change-induced alterations in plant growth
287 and physiology. Although higher levels of atmospheric CO₂ are *a priori* not expected to directly affect
288 soil microbial communities (*i.e.*, CO₂ concentrations in the soil are much higher than in the atmosphere),
289 higher atmospheric CO₂ concentrations can indirectly impact on soil microbial communities through
290 higher plant growth, increases in litter deposition and rhizodeposition (often resulting in a stimulation of
291 soil microbial biomass and activity), faster nutrient uptake and water use efficiency (Phillips et al. 2011;
292 Bardgett et al. 2013; Burns et al. 2013; Alkorta et al. 2017). Such climate change-induced variations of
293 rhizodeposition patterns, and concomitant changes in the composition and activity of rhizosphere
294 microorganisms, can modify TE bioavailability in soils (Rajkumar et al. 2013), thus potentially affecting
295 plant performance during phytomanagement.

296 The consequences of climate change (via higher atmospheric CO₂ concentrations, heat waves,
297 extreme droughts, higher temperatures, etc.) on beneficial plant-microorganism interactions (*e.g.*, plant
298 growth-promoting rhizobacteria, endophytes, and mycorrhiza) are increasingly being studied (Compant et
299 al. 2005, 2010; Classen et al. 2015; Cavicchioli et al. 2019; Risueño et al. 2020). Plant growth-promoting
300 bacteria and fungi can positively affect water-stressed plants and, then, their inoculation should nowadays
301 be strongly considered for phytomanagement. On the other hand, climate-induced changes in soil
302 temperature and moisture can alter soil processes, such as organic matter decomposition and nutrient
303 cycling (Burns et al. 2013), supported, to a great extent, by the activity of soil microorganisms.

304 Some phytomanagement practices (*e.g.*, the application of organic amendments, low- or no-
305 tillage practices, grassland implementation and afforestation) have great potential for carbon sequestration

306 and, hence, climate change mitigation. The incorporation of trees in phytomanagement initiatives (*e.g.*, as
307 part of intercropping systems) has also acknowledged positive effects in this respect (Schoeneberger et al.
308 2012; Alam et al. 2014; Zeng et al. 2019a,b; Brereton et al. 2020).

309 In theory, a possible option for adaptation to climate change in phytomanagement is to
310 incorporate to the planting scheme as many plant species as possible, and preferably from different
311 vegetation types: grasses, shrubs and trees. Nevertheless, in many situations this is not a realistic, feasible
312 option because the specific plant assemblages established for phytomanagement purposes are determined,
313 to a great extent, on the future land use and on the particular non-food crops intended to be delivered to
314 the local chains processing the harvestable biomass.

315 Moreover, the conservation of plant biodiversity (*e.g.*, the aforementioned metallophyte
316 diversity) is crucial for adaptation to climate change as part of an “insurance policy”: different species,
317 varieties and ecotypes may be needed in the future as environmental conditions are altered by climate
318 change.

319

320 **4.2. Promotion of biodiversity under phytomanagement**

321 Under the inherent constraints inevitably derived from the phytomanagement goals, it is certainly possible
322 to promote biodiversity in phytomanaged sites by means of, for instance, growing as many plant species
323 and varieties/ecotypes from different vegetation types (grasses, shrubs and trees) as possible (Table 1).
324 Interestingly, the establishment of different plant species in phytomanaged sites can result in the
325 generation of a wider variety of valuable products and ecosystem services (Evangelou et al. 2015; Pandey
326 and Bauddh 2018).

327 Many additional benefits can be obtained when combining different plant species for the
328 phytoremediation and phytomanagement of polluted sites. For instance, the combination of *Pteris vittata*
329 with *Morus alba* and *Broussonetia papyrifera* not only increased the phytoextraction of trace elements
330 but alleviated phytotoxicity as well (Zeng et al. 2019b). In a similar study, the co-planting of *P. vittata*
331 with *Arundo donax*, *M. alba* and *B. papyrifera* resulted in an improvement of soil health (Zeng et al.
332 2019a). Furthermore, intercropping with *Paspalum miliaceum* and *Axonopus affinis* was efficient in
333 promoting grapevine growth in Cu-polluted soil by reducing metal bioavailability (De Conti et al. 2019).
334 Interestingly, the combination of tree, shrub and grass species in a metal polluted soil resulted in a more
335 efficient employment of water resources and a higher biodiversity of soil microorganisms (Parraga-

336 Aguado et al. 2014). In contrast, the co-planting of *Odontarrhena chalcidica* or *Noccaea goesingensis*
337 with *Lotus corniculatus* for Ni removal led to reduced values of shoot biomass (Rosenkranz et al. 2019).

338 The beneficial effects of co-planting have also been reported for organically-polluted and mixed-
339 polluted soils. Wang et al. (2013) reported an enhanced degradation of PAHs, in the presence of trace
340 elements, when *S. alfredii* was combined with *Lolium perenne* or *Ricinus communis*. In agreement with
341 these results, in their studies on intercropping with *Medicago sativa* and *Festuca arundinacea*, Sun et al.
342 (2011) observed higher PAH degradation values under intercropping vs. monoculture. Likewise,
343 intercropping with *M. sativa*, *L. perenne* and *F. arundinacea* improved the degradation of phthalic acid
344 esters. Finally, *F. arundinacea* was also co-planted with *Salix miyabeana* and *M. sativa*, finding out that
345 when crops were cultivated in pairs they showed an enhanced rhizosphere community in terms of the
346 presence of plant growth-promoting bacteria (Brereton et al. 2020).

347 Aboveground and belowground organisms are closely linked: plants provide organic carbon for
348 soil decomposers and resources for root-associated organisms; in turn, soil decomposers break down dead
349 plant material and regulate plant growth by determining the nutrient supply (Wardle et al. 2004).
350 Different plant species differ in the quantity and quality of litter and root exudates, thus affecting the
351 biomass, activity and diversity (mainly, composition) of soil microbial communities. A more diversified
352 vegetation leads to a higher number of ecological niches and, hence, biodiversity (Risueño et al. 2020).
353 Indeed, higher plant richness results in a higher variety of root exudates and types of litter, thus
354 stimulating biodiversity belowground (Wardle et al. 2004; Haichar et al. 2008). Nonetheless, Li et al.
355 (2015) found no relationship between plant and soil bacterial diversity in an early successional forest, and
356 a negative correlation in a late successional forest. Similarly, Kowalchuk et al. (2000) reported a negative
357 correlation between grassland plant and soil ammonia-oxidizing bacterial diversity. These contradictory
358 results point out to the vast complexity of the multiple links and interactions between aboveground and
359 belowground diversity (Wardle et al. 2004; De Deyn and Van der Putten 2005; Kardol and Wardle 2010),
360 which, at the moment, are far from being well understood. Phytomanagement, apart from increasing soil
361 microbial biomass and activity, can induce shifts in the bacterial community structure at both the total
362 community and functional group levels (Touceda-González et al. 2017a). In a study on the effectiveness
363 of dolomite and compost as amendments for enhancing Cu phytostabilization with *Populus trichocarpa* x
364 *deltoides* cv. Beaupré and *Agrostis gigantea* L., Cu stabilization and phytomanagement induced positive

365 changes in the microbial community of soil leachates, enriching this community with plant beneficial
366 bacteria (Giagnoni et al. 2020).

367 The presence of phytopathogens and root herbivores in the rhizosphere can produce a negative
368 feedback on plant growth, whereas mycorrhizal fungi and plant-growth promoting rhizobacteria can have
369 a positive one on plant growth (Sessitsch et al. 2013; Sura-de Jong et al. 2015). In any case, the evidence
370 for positive or negative links between aboveground and belowground biodiversity is mixed, and not all of
371 the mechanisms by which aboveground organisms affect belowground diversity and vice versa
372 necessarily lead to correlations of species richness in both domains (Hooper et al. 2000). The common
373 perception that belowground biodiversity should follow similar patterns to those of plant diversity during
374 ecosystem development is challenged by Delgado-Baquerizo et al. (2020).

375 A higher richness of plant species can, for instance, be used to promote the biodegradation of
376 aged polycyclic aromatic compounds in soil: oxygenated PAHs (some of which are more toxic than their
377 related PAHs) can, however, accumulate in soils during such a plant-assisted remediation process
378 (Bandowe et al. 2019).

379 Through an increase in plant diversity and, hence, in the number of ecological niches and
380 possible habitats, it is also desirable to promote the aboveground and belowground diversity of animals
381 (*e.g.*, arthropods: insects, arachnids, myriapods, etc.; earthworms; nematodes; mammals; birds; and so
382 on), of course, always paying close attention to the potential risk of pollutant bioaccumulation and
383 biomagnification (*e.g.*, TE biomagnification along the trophic chain) (Peterson et al. 2003). Interestingly,
384 these animals can act as phytomanagement crop auxiliaries, helping to fight pests, pollinate the cultivated
385 plants, etc. (Verkerk et al. 1998; Ferron and Deguine 2005).

386 Finally, intercropping systems have been extensively investigated for phytoremediation purposes
387 (Sun et al. 2011; Ma et al. 2013; Wang et al. 2013; Alam et al. 2014; De Conti et al. 2019; Zeng et al.
388 2019a,b; Brereton et al. 2020) with additional benefits in terms of aboveground and belowground
389 diversity. Likewise, as individual plant species repeatedly possess a limited range of TE phytoremediation
390 capacities, functional complementarity principles could be of value for the phytoremediation of soils
391 polluted with multiple TEs by means of using assemblages of species (Desjardins et al. 2018).

392 Biodiversity provides a wide range of values, some of them indirectly such as aesthetic value,
393 cultural value, spiritual value, scientific value, educational value, etc. Arguably, from an anthropocentric
394 point of view, the most important value of biodiversity comes from the ecosystem services it provides.

395 Biodiversity preserves the structure and integrity on which healthy ecosystems depend to provide the vital
396 ecosystem services on which we rely on.

397 Among other values of biodiversity, the following two are often discussed when dealing with the
398 conservation of biodiversity and the human use of natural resources: (1) *intrinsic value*: as such, we have
399 the moral responsibility to preserve biodiversity (well-known nature writers such as Henry David
400 Thoreau, John Muir, Aldo Leopold, etc. have emphasized the intrinsic value of biodiversity); and (2)
401 *utilitarian value*: as such, focused on the commercial and subsistence benefits (*e.g.*, food, medicines, raw
402 materials, energy, etc.) of biodiversity to humankind. Within this utilitarian perspective, the idea is to
403 protect biodiversity so that we can utilize it later for our own benefit. Obviously, this utilitarian value of
404 biodiversity is inextricably linked to the phytomanagement concept. In any case, when designing a
405 phytomanagement initiative, it is unquestionably possible to promote biodiversity within the limits
406 imposed by the specific phytomanagement objectives (*e.g.*, by means of growing as many plant species as
407 possible) with the concomitant potential benefit of obtaining a wider variety of products and ecosystem
408 services.

409 Anyhow, the biodiversity concept is anything but simple. Among others, it includes the
410 following aspects: *richness* (or the number of species), *evenness* (relative abundances resulting in rare and
411 dominant species), *composition* (in terms of taxonomic groups), phylogenetic relatedness/distinctiveness,
412 and spatial and temporal distribution. Regarding species composition, biological species are certainly not
413 all equal: there are keystone species, foundation species, umbrella species, flagship species, charismatic
414 species, ecosystem engineers, invasive species, indicator species, chemical engineers, biological
415 regulators, etc., leading us to the difficult and arduous challenge to prioritize among them (Vane-Wright
416 et al. 1991).

417 Some authors proposed to assign more value to those species that lack close relatives, as by
418 maximizing the conservation of evolutionary diversity, we maximize genotypic, phenotypic and
419 functional diversity, and, hence, provide ecosystems with the most options to adapt to a changing world
420 (Vane-Wright et al. 1991; Cadotte et al. 2010). Besides, some species appear to perform phylogenetically
421 narrow processes (*e.g.*, nitrification, atmospheric nitrogen fixation) while others perform phylogenetically
422 broad processes (*e.g.*, denitrification). The former show a lower degree of functional redundancy,
423 compared to the latter.

424 To assess the influence of phytomanagement practices on biodiversity (such a broad concept) is
425 anything but easy. There are still many unanswered questions that research is yet to answer, *e.g.* What
426 number of species is a good number? What species composition is best? What degree of phylogenetic
427 distance is more adequate? How differently should we value the different types of species? Are
428 functionally redundant species less valuable than non-functionally redundant species? These questions
429 being answered, we must not take only richness into consideration when promoting biodiversity at
430 polluted sites under phytomanagement. To the best of our expertise and capacities, we must try to
431 consider other relevant aspects also included within the biodiversity concept.

432 To further complicate matters, biodiversity is difficult to quantify, at least partly, due to the
433 multitude of indices available to measure it (*e.g.*, species richness, Shannon-Wiener entropy, Simpson's
434 index, Berger-Parker index, etc.). This is not surprising because of the abovementioned complexity of all
435 the aspects of biodiversity, which inevitably leads to the fact that no single perfect indicator for
436 biodiversity can be devised (Duelli and Obrist 2003). As a matter of fact, the choice of index often
437 depends on the question(s) to be answered, as well as on the specific aspect(s) or entity of biodiversity to
438 be evaluated. Paradoxically, most diversity indices have traditionally relied on three untrue assumptions:
439 (i) all species are equal; (ii) all individuals are equal; and (iii) species abundances have been correctly
440 assessed with appropriate tools and in similar units (Magurran 2004). In any case, although the choice of
441 index(es) depends, to a great extent, on the specific questions and objectives of the study, three of the
442 most commonly used indices are the Margalef's index for richness, the Shannon-Weaver's index for
443 diversity and the Simpson's index for dominance.

444 Similarly, the use of indices for quantifying functional biodiversity (functional richness,
445 functional evenness, and functional divergence) is essential to better understand the links between
446 biodiversity, ecosystem functioning and environmental constraints (Mouchet et al. 2010). Indeed, many
447 studies on the impact of disturbances (*e.g.*, agronomic practices, contamination, climate change, nitrogen
448 deposition, etc.) on biological diversity are focused exclusively on structural biodiversity (usually, of only
449 one or a few taxonomic groups). But phytomanagement has a strong functional component related to the
450 provision of ecosystem services. Thus, it is highly beneficial to include both types of biodiversity, *i.e.*
451 structural and functional diversity, when promoting biodiversity under phytomanagement. Apart from a
452 selection, as wide as possible, of taxonomic groups, an analysis of functional groups, traits, guilds and so

453 on must be included in phytomanagement initiatives (Kumpiene et al. 2014; Durand et al. 2017; Touceda-
454 González et al. 2017a,b; Xue et al. 2018; Burges et al. 2020).

455 Although the identification of links between structural and functional biodiversity is undoubtedly
456 a challenging task, such identification is of much value from both an academic/scientific and management
457 point of view. Statistical multivariate analyses, applied to the group of variables used to measure
458 structural and functional diversity, are suitable tools for the establishment of hypotheses regarding the
459 abovementioned links.

460 The topic of the selection of the best indices to quantify both structural and functional
461 biodiversity is not within the scope of this document. Nonetheless, we encourage the use of various
462 indices for covering as much as possible the different aspects of the term biodiversity: richness,
463 abundance, phylogenetic relatedness, functional traits, etc. Ideally, one should make the best efforts
464 possible to evaluate the effect of phytomanagement practices on the various levels of biodiversity:
465 genetic, species, populations and communities/ecosystems. However, biodiversity is not simply the sum
466 of all ecosystems, species and genetic material, as it represents the variability within and among them.

467 In particular, for soil microorganisms, the assessment of genetic diversity is indispensable for
468 microbial ecologists since: (i) all the current definitions of “species” are inadequate for prokaryotes,
469 among other reasons due to the transfer of genes by horizontal gene transfer; and (ii) most
470 microorganisms cannot be cultivated and so we have no other choice than to study them by means of the
471 application of molecular biology techniques. In consequence, most soil microbial ecologists are nowadays
472 focused on the use of next generation sequencing techniques (*e.g.*, metabarcoding, metagenomics) for the
473 quantification of soil microbial diversity. But next generation sequencing has still many technical
474 limitations and then we must be cautious when drawing conclusions about the effect of disturbances or
475 practices (*e.g.*, phytomanagement) on soil microbial diversity.

476 Most studies on the effect of phytoremediation or phytomanagement practices on microbial
477 diversity are focused on soil microbial communities, especially rhizosphere microorganisms. In this
478 respect, more attention should be paid to plant microbiota and plant microbiomes (*e.g.*, in the
479 phyllosphere) under phytomanagement (Imperato et al. 2019).

480 Similarly, concerning genetic diversity, there are still many unanswered questions, such as, for
481 example: The more genes the better? Are all genes equally important? Can we talk about “good” genes

482 (*e.g.*, genes involved in contaminant biodegradation pathways) and “bad” genes (*e.g.*, antibiotic resistance
483 genes)? How can be combined data from metagenomic, metatranscriptomic and metaepigenomic studies?

484 Likewise, when dealing with ecosystem diversity (*i.e.*, the richness and complexity of biological
485 communities, including trophic levels and ecological processes, together with the chemical and physical
486 environment), additional questions emerge: How many trophic levels do we need? Are all of them equally
487 important? How many species per trophic level are needed?

488 Regarding the critical links between biodiversity and ecosystem functioning, one should take
489 into consideration the concept of emergent properties, *i.e.* those new qualities that appear on higher
490 integration levels and represent more than the sum of the low-level components (Reuter et al. 2005). For
491 understanding these emergent properties, the interaction between the different elements must be closely
492 studied (Reuter et al. 2005). In consequence, when possible, key biological interactions should be
493 identified and studied during phytomanagement initiatives, since they support the functioning of the
494 ecosystem and are the basis of emergent properties.

495 On the other hand, when promoting biodiversity under phytomanagement, it is important to
496 always include organisms from the different levels of the trophic chain. Instead, when evaluating their
497 effect on biodiversity, most phytomanagement initiatives only pay attention to aboveground botanical
498 diversity (richness, composition, vegetation structure) and, occasionally, include some belowground soil
499 biota, in many cases just microorganisms owing to their well-known key role in critical soil processes
500 and, hence, functions and ecosystem services (Xue et al. 2018; Burges et al. 2020). Nonetheless, for a
501 biodiversity assessment with more ecological relevance, it is desirable to include taxonomic groups from
502 the different levels of the trophic chain. As a matter of fact, we should study as many taxonomic groups
503 from the food web as possible (Garrouj et al. 2018; Ali et al. 2019; Prins et al. 2019).

504 Simplifying, the aboveground food web includes producers (plants), primary consumers
505 (herbivores) and secondary consumers (predators). Regarding consumers, there is an unresolved debate
506 regarding the benefits and disadvantages associated to the presence of animals in phytomanaged sites.
507 Actually, when dealing with the remediation of polluted sites, in many cases, animals are deliberately
508 excluded, in an attempt to avoid possible ecotoxic effects on exposed animals and, also, to minimize the
509 risk of bioaccumulation and biomagnification (Mann et al. 2011). But animals (*e.g.*, arthropods,
510 earthworms, mammals, birds, etc.) can act as phytomanagement crop auxiliaries, helping to fight pests,
511 pollinate the cultivated plants, etc. (Verkerk et al. 1998; Ferron and Deguine 2005).

512 Pertaining to the soil ecosystem, an amazing diversity of soil organisms make up its food web:
513 bacteria, fungi, algae, protozoa, nematodes, micro-arthropods, earthworms, insects, small vertebrates
514 (mice, moles), etc. Like all food webs, the soil food web is fueled by primary producers such as plants,
515 lichens, mosses, photosynthetic bacteria (*e.g.*, cyanobacteria) and unicellular algae. The remaining
516 members of the soil biota obtain energy and carbon by consuming the organic compounds produced by
517 primary producers.

518

519 *4.3. Adaptive monitoring during phytomanagement*

520 When dealing with long-term monitoring programs, such as the one for assessing the influence of
521 phytomanagement practices on biodiversity, as time passes, it is inevitable that (i) new analytical
522 techniques, methods and equipments might appear in the market; (ii) different approaches, concepts,
523 ideas, etc. might come up; (iii) changes in the ecosystem developmental stage will occur; (iv) unexpected
524 environmental threats might emerge; (v) budget fluctuations might threaten the initiative, and so forth
525 (Epelde et al. 2014). For that reason, we propose that the paradigm of adaptive monitoring (this paradigm
526 enables monitoring programs to evolve iteratively as new information emerges and research questions
527 change) should be incorporated to the long-term monitoring of the effect of phytomanagement practices
528 on biodiversity.

529 To this purpose, among other aspects, (i) well-formulated, clear and tractable questions must be
530 established at the beginning of the phytomanagement initiative; (ii) a rigorous statistical design must be
531 implemented from the onset of the study, notably accounting for the spatial variability of soil
532 contamination, contaminant exposure and pollutant linkage; and (iii) a conceptual model of the site under
533 phytomanagement must be created (Cundy et al. 2016).

534 As part of the adaptive monitoring program, periodically (the time period will depend on the
535 specific phytomanagement initiative), an expert judgment analysis must be organized to revise and, if
536 necessary, update the different aspects that make up the biodiversity monitoring program. Expert
537 judgment analyses often encourage the forging of partnership between researchers, policy-makers and
538 resource managers, an aspect of the utmost importance in phytomanagement (Cundy et al. 2013).

539 The accomplishment of economic, social and environmental benefits is a key aspect of
540 phytomanagement (Cundy et al. 2016). In particular, the provisioning of ecosystem services (carbon
541 sequestration, improvement of soil fertility, control of soil erosion, improvement of air quality, climate

542 and water regulation, production of atmospheric oxygen, provision of habitat, etc.) is a crucial component
543 of phytomanagement initiatives (Burges et al. 2018).

544 The provision of ecosystem services is underpinned by a variety of ecological processes and
545 functions which themselves are driven by biodiversity. Although changes in biodiversity can affect
546 ecosystem processes and, hence, the provision of ecosystem services, in some situations, biomass, species
547 composition, functional traits, etc. are more important than biodiversity itself for the provisioning of those
548 services. Nonetheless, trade-offs between biodiversity and ecosystem services might arise in some
549 situations (Bandowe et al. 2019). Also, trade-offs and conflicts between the different ecosystem services
550 themselves might also emerge, and, then, it is desirable to select from the onset what specific ecosystem
551 services to promote, and implement measures that minimize conflicts.

552

553 **Conclusion**

554 Paraphrasing the three well-known M's of successful trading (Mind, Money management, and Method),
555 we can visualize the links between biodiversity and phytomanagement according to three M's of
556 successful phytomanagement: (1) *Mind*: for effective phytomanagement, we must use our mind and
557 creativity to design the best strategy for each specific site and casuistry (here, following the medical
558 aphorism "there are no diseases but sick people", we can state that "there is no pollution but polluted
559 areas"; all of them are different and require a site-specific assessment). In this respect, biodiversity
560 provides ideas, models and strategies (tested through millions of years of evolution) that we can learn
561 from; (2) *Management*: for successful phytomanagement, we must apply scientifically-based adaptive
562 management, especially under the current scenario of climate change. Biodiversity provides a myriad of
563 species, metabolic capabilities, functional traits, etc. which we can use in response to changing
564 conditions; and (3) *Money*: a fruitful phytomanagement will provide economic value through products
565 (crops for biomass-processing technologies) and ecosystem services which can help fuel our bioeconomy.
566 Interestingly, the promotion of biodiversity in phytomanaged sites can result in the generation of a wider
567 variety of valuable products and ecosystem services, while minimizing pollutant-induced environmental
568 risks.

569

570

571

572 **References**

- 573 Abou-Shanab R, Angle J, Chaney R (2006) Bacterial inoculants affecting nickel uptake by *Alyssum*
574 *murale* from low, moderate and high Ni soils. *Soil Biol Biochem* 38:2882-2889. [https://doi](https://doi.org/10.1016/j.soilbio.2006.04.045)
575 [10.1016/j.soilbio.2006.04.045](https://doi.org/10.1016/j.soilbio.2006.04.045)
- 576 Agnello AC, Bagard M, van Hullebusch ED, Esposito G, Huguenot D (2016) Comparative
577 bioremediation of heavy metals and petroleum hydrocarbons co-contaminated soil by natural
578 attenuation, phytoremediation, bioaugmentation and bioaugmentation-assisted phytoremediation.
579 *Sci Total Environ* 563-564:693-703. [https:// doi:10.1016/j.scitotenv.2015.10.061](https://doi.org/10.1016/j.scitotenv.2015.10.061)
- 580 Alam M, Olivier A, Paquette A, Dupras J, Revéret J-P, Messier C (2014) A general framework for the
581 quantification and valuation of ecosystem services of tree-based intercropping systems.
582 *Agroforestry Systems* 88:679-691
- 583 Ali H, Khan E (2019) Trophic transfer, bioaccumulation, and biomagnification of non-essential hazardous
584 heavy metals and metalloids in food chains/webs-Concepts and implications for wildlife and
585 human health. *Human Ecol Risk Assess* 25:1353-1376. [https://doi](https://doi.org/10.1080/10807039.2018.1469398)
586 [10.1080/10807039.2018.1469398](https://doi.org/10.1080/10807039.2018.1469398)
- 587 Alkorta I, Epelde L, Garbisu C (2017) Environmental parameters altered by climate change affect the
588 activity of soil microorganisms involved in bioremediation. *FEMS Microbiol Lett* 364. [https://](https://doi.org/10.1093/femsle/fnx200)
589 [doi 10.1093/femsle/fnx200](https://doi.org/10.1093/femsle/fnx200)
- 590 Alkorta I, Hernández-Allica J, Becerril JM, Amezaga I, Albizu I, Garbisu C (2004a) Recent findings on
591 the phytoremediation of soils contaminated with environmentally toxic heavy metals and
592 metalloids such as zinc, cadmium, lead, and arsenic. *Rev Environ Sci Bio/Technol* 3:71-90
- 593 Alkorta I, Hernández-Allica J, Becerril JM, Amezaga I, Albizu I, Onaindia M, Garbisu C (2004b)
594 Chelate-enhanced phytoremediation of soils polluted with heavy metals. *Rev Environ Sci*
595 *Bio/Technol* 3:55-70
- 596 Anza M, Salazar O, Epelde L, Garbisu C (2018) Data on the selection of biostimulating agents for the
597 bioremediation of soil simultaneously contaminated with lindane and zinc. *Data in Brief*
598 *20:1371-1377*. [https://doi 10.1016/j.dib.2018.08.203](https://doi.org/10.1016/j.dib.2018.08.203)

599 Baker AJM, Ernst WHO, van der Ent A, Malaisse F, Ginocchi R (2010) Metallophytes: the unique
600 biological resource, its ecology and conservational status in Europe, central Africa and Latin
601 America. In: Batty LC, Hallberg KB (eds) Ecology of industrial pollution, Cambridge University
602 Press, British Ecological Society, pp. 7-40

603 Bandowe BAM, Leimer S, Meusel H, Velescu A, Dassen S, Eisenhauer N, Hoffmann T, Oelmann Y,
604 Wilcke W (2019) Plant diversity enhances the natural attenuation of polycyclic aromatic
605 compounds (PAHs and oxygenated PAHs) in grassland soils. *Soil Biol Biochem* 129:60-70.
606 <https://doi.org/10.1016/j.soilbio.2018.10.017>

607 Bardgett R, Manning P, Morriën E, De Vries FT (2013) Hierarchical responses of plant-soil interactions
608 to climate change: consequences for the global carbon cycle. *J Ecol* 101:334-43. <https://doi.org/10.1111/1365-2745.12043>

610 Barrutia O, Epelde L, García-Plazaola JI, Garbisu C, Becerril JM (2009) Phytoextraction potential of two
611 *Rumex acetosa* L. accessions collected from metalliferous and non-metalliferous sites: effect of
612 fertilization. *Chemosphere* 74:259-264. <https://doi.org/10.1016/j.chemosphere.2008.09.036>

613 Barrutia O, Artetxe U, Hernández A, Olano JM, García-Plazaola JI, Garbisu C, Becerril JM (2011a)
614 Native plant communities in an abandoned Pb-Zn mining area of Northern Spain: Implications
615 for phytoremediation and germplasm preservation *Int J Phytorem* 13:256-270. <https://doi.org/10.1080/15226511003753946>

617 Barrutia O, Garbisu C, Epelde L, Sampedro MC, Goicolea MA, Becerril JM (2011b) Plant tolerance to
618 diesel minimizes its impact on soil microbial characteristics during rhizoremediation of diesel-
619 contaminated soils. *Sci Total Environ* 409:4087-4093. <https://doi.org/10.1016/j.scitotenv.2011.06.025>

621 Batty LC (2005) The potential importance of mine sites for biodiversity. *Mine Water Environ* 24:101-103

622 Becerra-Castro C, Monterroso C, Prieto-Fernández A, Rodríguez-Lamas L, Loureiro-Viñas M, Acea MJ,
623 Kidd PS (2012) Pseudometallophytes colonising Pb/Zn mine tailings: a description of the plant-
624 microorganism-rhizosphere soil system and isolation of metal-tolerant bacteria. *J Hazard Mater*
625 217-218:350-359. <https://doi.org/10.1016/j.jhazmat.2012.03.039>

626 Brereton NJB, Gonzalez E, Desjardins D, Labrecque M, Pitre FE (2020) Co-cropping with three
627 phytoremediation crops influences rhizosphere microbiome community in contaminated soil. *Sci*
628 *Total Environ* 711:135067. [https://doi 10.1016/j.scitotenv.2019.135067](https://doi.org/10.1016/j.scitotenv.2019.135067)

629 Burges A, Epelde L, Benito G, Artetxe U, Becerril JM, Garbisu C (2016) Enhancement of ecosystem
630 services during endophyte-assisted aided phytostabilization of metal contaminated mine soil. *Sci*
631 *Total Environ* 562:480-492. [https://doi 10.1016/j.scitotenv.2016.04.080](https://doi.org/10.1016/j.scitotenv.2016.04.080)

632 Burges A, Epelde L, Blanco F, Becerril JM, Garbisu C (2017) Ecosystem services and plant physiological
633 status during endophyte-assisted phytoremediation of metal contaminated soil. *Sci Total Environ*
634 584:329-338. [https://doi 10.1016/j.scitotenv.2016.12.146](https://doi.org/10.1016/j.scitotenv.2016.12.146)

635 Burges A, Alkorta I, Epelde L, et al. (2018) From phytoremediation of soil contaminants to
636 phytomanagement of ecosystem services in metal contaminated sites *Int J Phytorem* 20:384-397.
637 [https://doi:10.1080/15226514.2017.1365340](https://doi.org/10.1080/15226514.2017.1365340)

638 Burges A, Fievet V, Oustriere N, Epelde L, Garbisu C, Becerril JM, Mench M (2020) Long-term
639 phytomanagement with compost and a sunflower - tobacco rotation influences the structural
640 microbial diversity of a Cu-contaminated soil. *Sci Total Environ* 700:134529. [https://doi](https://doi.org/10.1016/j.scitotenv.2019.134529)
641 [10.1016/j.scitotenv.2019.134529](https://doi.org/10.1016/j.scitotenv.2019.134529)

642 Burns RG, DeForest JL, Marxsen J, Sinsabaugh RL, Stromberger ME, Wallenstein MD, Weintraub MN,
643 Zoppini A (2013) Soil enzymes in a changing environment: current knowledge and future
644 directions. *Soil Biol Biochem* 58:216-34. [https://doi 10.1016/j.soilbio.2012.11.009](https://doi.org/10.1016/j.soilbio.2012.11.009)

645 Cabello-Conejo MI, Becerra-Castro C, Prieto-Fernández A, Monterroso C, Saavedra-Ferro A, Mench M,
646 Kidd PS (2014) Rhizobacterial inoculants can improve nickel phytoextraction by the
647 hyperaccumulator *Alyssum pintodasilvae*. *Plant Soil* 379:35-50. [https://doi 10.1007/s11104-014-](https://doi.org/10.1007/s11104-014-2043-7)
648 [2043-7](https://doi.org/10.1007/s11104-014-2043-7)

649 Cadotte MW, Davies TJ, Regetz J, Kembel SW, Cleland E, Oakley TH (2010) Phylogenetic diversity
650 metrics for ecological communities: Integrating species richness, abundance and evolutionary
651 history. *Ecol Lett* 13: 96-105. [https://doi 10.1111/j.1461-0248.2009.01405.x](https://doi.org/10.1111/j.1461-0248.2009.01405.x)

652 Cardinale BJ, Srivastava DS, Emmett Duffy J, Wright JP, Downing AL, Sankaran M, Jouseau Z (2006)
653 Effects of biodiversity on the functioning of trophic groups and ecosystems. *Nature* 443:989-992

654 Cavicchioli R, Ripple WJ, Timmis KN et al. (2019) Scientists' warning to humanity: microorganisms and
655 climate change. *Nat Rev Microbiol* 17:569-586. [https://doi 10.1038/s41579-019-0222-5](https://doi.org/10.1038/s41579-019-0222-5)

656 Chan KM, Pringle RM, Ranganathan J, Boggs CL, Chan YL, Ehrlich PR, Haff PK, Heller NE, Al-
657 Khafaji K, Macmynowski DP (2007) When agendas collide: human welfare and biological
658 conservation. *Conservation Biology* 21:51-68. [https://doi 10.1111/j.1523-1739.2006.00570.x](https://doi.org/10.1111/j.1523-1739.2006.00570.x)

659 Chen F, Yang Y, Mi J, Liu R, Hou HP, Zhang SL (2019) Effects of vegetation pattern and spontaneous
660 succession on remediation of potential toxic metal-polluted soil in mine dumps. *Sustainability*
661 11:397. [https://doi 10.3390/su11020397](https://doi.org/10.3390/su11020397)

662 Classen AT, Sundqvist MK, Henning JA, Newman GS, Moore JAM, Cregger MA, Moorhead LC,
663 Courtney M. Patterson CM (2015). Direct and indirect effects of climate change on soil
664 microbial and soil microbial-plant interactions: What lies ahead? *Ecosphere* 6:1-21. [https://doi](https://doi.org/10.1890/ES15-00217.1)
665 [10.1890/ES15-00217.1](https://doi.org/10.1890/ES15-00217.1)

666 Compant S, Duffy B, Nowak J, Clément C, Barka EA (2005) Use of plant growth-promoting bacteria for
667 biocontrol of plant diseases: Principles, mechanisms of action, and future prospects. *Appl*
668 *Environ Microbiol* 71:4951-4959. [https://doi 10.1128/AEM.71.9.4951-4959.2005](https://doi.org/10.1128/AEM.71.9.4951-4959.2005)

669 Compant S, Van Der Heijden MGA, Sessitsch, A (2010) Climate change effects on beneficial plant-
670 microorganism interactions, *FEMS Microbiol Ecol* 73:197-214, [https://doi 10.1111/j.1574-](https://doi.org/10.1111/j.1574-6941.2010.00900)
671 [6941.2010.00900](https://doi.org/10.1111/j.1574-6941.2010.00900)

672 Conesa HM, Evangelou MWH, Robinson BH, Schulin R (2012) A critical view of current state of
673 phytotechnologies to remediate soils: Still a promising tool? *Scientific World J* 2012: Article ID
674 173829. [https://doi 10.1100/2012/173829](https://doi.org/10.1100/2012/173829)

675 Corzo Remigio A, Chaney RL, Baker AJM, Edraki M, Erskine PD, Echevarria G, van der Ent A (2020)
676 Phytoextraction of high value elements and contaminants from mining and mineral wastes:
677 opportunities and limitations. *Plant Soil*. [https://doi10.1007/s11104-020-04487-3](https://doi.org/10.1007/s11104-020-04487-3)

678 Cundy A, Witter N, Mench M, Friesl W, Müller I, Neu S (2013) Developing principles of sustainability
679 and stakeholder engagement for "gentle" remediation approaches: the European context. *J*
680 *Environ Manage* 129:283-291. [https://doi 10.1016/j.jenvman.2013.07.032](https://doi.org/10.1016/j.jenvman.2013.07.032)

681 Cundy AB, Bardos RP, Puschenreiter M, et al. (2016) Brownfields to green fields: Realising wider
682 benefits from practical contaminant phytomanagement strategies. *J Environ Manage* 184:67-77.
683 [https://doi 10.1016/j.jenvman.2016.03.028](https://doi.org/10.1016/j.jenvman.2016.03.028)

684 De Conti L, Ceretta CA, Melo GWB, Tiecher TL, Silva LOS, Garlet LP, Mimmo T, Cesco S, Brunetto G
685 (2019) Intercropping of young grapevines with native grasses for phytoremediation of Cu-
686 contaminated soils. *Chemosphere* 216:147-156. [https://doi 10.1016/j.chemosphere.2018.10.134](https://doi.org/10.1016/j.chemosphere.2018.10.134)

687 De Deyn GB, Van der Putten WH (2005) Linking aboveground and belowground diversity. *Trends Ecol*
688 *Evol* 20:625-633

689 Delgado-Baquerizo M, Bardgett RD, Vitousek PM, et al. (2019) Changes in belowground biodiversity
690 during ecosystem development. *Proc Natl Acad Sci* 116:6891-6896. [https://doi](https://doi.org/10.1073/pnas.1818400116)
691 [10.1073/pnas.1818400116](https://doi.org/10.1073/pnas.1818400116)

692 Dell'Amico E, Cavalca L, Andreoni V (2008) Improvement of *Brassica napus* growth under cadmium
693 stress by cadmium-resistant rhizobacteria. *Soil Biol Biochem* 40:74-84. [https://doi](https://doi.org/10.1016/j.soilbio.2007.06.024)
694 [10.1016/j.soilbio.2007.06.024](https://doi.org/10.1016/j.soilbio.2007.06.024)

695 Desjardins D, Brereton NJB, Marchand L, Brisson J, Pitre FE, Labrecque M (2018) Complementarity of
696 three distinctive phytoremediation crops for multiple-trace element contaminated soil. *Sci Total*
697 *Environ* 610-611:1428-1438. [https://doi 10.1016/j.scitotenv.2017.08.196](https://doi.org/10.1016/j.scitotenv.2017.08.196)

698 Duelli P, Obrist MK (2003) Biodiversity indicators: the choice of values and measures. *Agric Ecosyst*
699 *Environ* 98:87-98

700 Durand A, Maillard F, Foulon J, Gweon H S, Valot B, Chalot M (2017) Environmental metabarcoding
701 reveals contrasting belowground and aboveground fungal communities from poplar at a Hg
702 phytomanagement site. *Microbial Ecol* 74:795-809. [https://doi 10.1007/s00248-017-0984-0](https://doi.org/10.1007/s00248-017-0984-0)

703 Dushenkov V, Nanda Kumar PBA, Motto H, Raskin I (1995) Rhizofiltration: The use of plants to remove
704 heavy metals from aqueous streams. *Environ Sci Technol* 29: 1239-1245

705 Epelde L, Becerril JM, Hernández-Allica J, Barrutia O, Garbisu C (2008) Functional diversity as
706 indicator of the recovery of soil health derived from *Thlaspi caerulescens* growth and metal
707 phytoextraction. *Applied Soil Ecology* 39:299-310. [https://doi 10.1016/j.apsoil.2008.01.005](https://doi.org/10.1016/j.apsoil.2008.01.005)

708 Epelde L, Becerril JM, Mijangos I, Garbisu C (2009) Evaluation of the efficiency of a phytostabilization
709 process with biological indicators of soil health. *J Environ Quality* 38:2041-2049. <https://doi>
710 [10.2134/jeq2009.0006](https://doi.org/10.2134/jeq2009.0006)

711 Epelde L, Becerril JM, Kowalchuk GA, Deng Y, Zhou J, Garbisu C (2010) Impact of metal pollution and
712 *Thlaspi caerulescens* growth on soil microbial communities. *Appl Environ Microbiol* 76:7843-
713 7853. <https://doi> [10.1128/AEM.01045-10](https://doi.org/10.1128/AEM.01045-10)

714 Epelde L, Becerril JM, Alkorta I, Garbisu C (2014) Adaptive long-term monitoring of soil health in metal
715 phytostabilization: Ecological attributes and ecosystem services based on soil microbial
716 parameters. *Int J Phytorem* 16:971-981. <https://doi> [10.1080/15226514.2013.810578](https://doi.org/10.1080/15226514.2013.810578)

717 Evangelou MWH, Papazoglou EG, Robinson BH, Schulin R (2015) Phytomanagement: phytoremediation
718 and the production of biomass for economic revenue on contaminated land. In: Ansari AA, Gill
719 SS, Gill R, Lanza GR, Newman L (eds) *Phytoremediation: management of environmental*
720 *contaminants*, Volume 1, Springer International Publishing, Cham, pp 115-132

721 Fässler E, Robinson BH, Stauffer W, Gupta SK, Papritz A, Schulin R (2010) Phytomanagement of metal-
722 contaminated agricultural land using sunflower, maize and tobacco. *Agric Ecosyst Environ*
723 136:49-58. <https://doi> [10.1016/j.agee.2009.11.007](https://doi.org/10.1016/j.agee.2009.11.007)

724 Ferron P, Deguine J-P (2005) Crop protection, biological control, habitat management and integrated
725 farming. A review. *Agron Sustainable Development* 25:17-24

726 Garaiurrebaso O, Garbisu C, Blanco F, Lanzén A, Martín I, Epelde L, Becerril JM, Jechalke S, Smalla
727 K, Grohmann E, Alkorta I (2017) Long-term effects of aided phytostabilization on microbial
728 communities of metal-contaminated mine soil. *FEMS Microbiol Ecol* 93(3). <https://doi>
729 [10.1093/femsec/fiw252](https://doi.org/10.1093/femsec/fiw252)

730 Garbisu C, Alkorta I (2001) Phytoextraction: a cost-effective plant-based technology for the removal of
731 metals from the environment. *Bioresource Technol* 77:229-236.

732 Garbisu C, Garaiurrebaso O, Epelde L, Grohmann E, Alkorta I (2017) Plasmid-mediated
733 bioaugmentation for the bioremediation of contaminated soils. *Front Microbiol* 8(1966).
734 <https://doi> [10.3389/fmicb.2017.01966](https://doi.org/10.3389/fmicb.2017.01966)

735 Garrouj M, Marchand L, Frayssinet M, Mench M, Castagneyrol B (2018) Trace element transfer from
736 two contaminated soil series to *Medicago sativa* and one of its herbivores, *Spodoptera exigua*.
737 Int J Phytorem 20:650-657. [https://doi 10.1080/15226514.2017.1374342](https://doi.org/10.1080/15226514.2017.1374342)

738 Giagnoni L, dos Anjos Borges LG, Giongo A, de Oliveira Silveira A, Ardisson AN, Triplett EW, Mench
739 M, Renella G (2020) Dolomite and compost as amendments for enhancing phytostabilization
740 and increasing microbiota of the leachates from a Cu-contaminated soil. Agronomy 10:719.
741 [https://doi 10.3390/agronomy10050719](https://doi.org/10.3390/agronomy10050719)

742 Ginocchio R, Baker AJM. (2004) Metallophytes in Latin America: a remarkable biological and genetic
743 resource scarcely known and studied in the region. Revista Chilena de Historia Natural 77:185-
744 194. [http://doi 10.4067/S0716-078X2004000100014](http://doi.org/10.4067/S0716-078X2004000100014)

745 Haichar FZ, Marol C, Berge O, Rangel-Castro JI, Prosser J, Balesdent J, Heulin T, Achouak W (2008)
746 Plant host habitat and root exudates shape soil bacterial community structure. ISME J 2: 1221-
747 1230

748 Haines-Young R, Potschin M. (2010) The links between biodiversity, ecosystem services and human
749 well-being. In: Raffaelli DG, Frid CLJ (eds), Ecosystem ecology: a new synthesis, Cambridge
750 University Press. British Ecological Society, pp. 110-139

751 Harrison JG, Griffin EA (2020) The diversity and distribution of endophytes across biomes, plant
752 phylogeny and host tissues: how far have we come and where do we go from here? Environ
753 Microbiol. [https://doi 10.1111/1462-2920.14968](https://doi.org/10.1111/1462-2920.14968)

754 Hernández-Allica J, Becerril JM, Zárate O, Garbisu C (2006) Assessment of the efficiency of a metal
755 phytoextraction process with biological indicators of soil health. Plant and Soil 281:147-158

756 Herzig R, Nehnevajova E, Pfistner C, Schwitzguebel JP, Ricci A, Keller C (2014) Feasibility of labile Zn
757 phytoextraction using enhanced tobacco and sunflower: Results of five- and one-year field-scale
758 experiments in Switzerland. Int J Phytorem 16:735-754. [https://doi
759 10.1080/15226514.2013.856846](https://doi.org/10.1080/15226514.2013.856846)

760 Hooper DU, Bignell DE, Brown VK, et al. (2000) Interactions between aboveground and belowground
761 biodiversity in terrestrial ecosystems: Patterns, mechanisms, and feedbacks. Bioscience 50:1049-
762 1061

763 Imperato V, Portillo-Estrada M, McAmmond BM, Douwen Y, Van Hamme JD, Gawronski SW,
764 Vangronsveld J, Thijs S (2019) Genomic diversity of two hydrocarbon-degrading and plant
765 growth-promoting *Pseudomonas* species isolated from the oil field of Bóbrka (Poland). *Genes*
766 (Basel) 10:443. <https://doi.org/10.3390/genes10060443>

767 IPBES (2019) Summary for policymakers of the global assessment report on biodiversity and ecosystem
768 services of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem
769 Services. S. Díaz, J. Settele, E. S. Brondízio E.S., H. T. Ngo, M. Guèze, J. Agard, A. Arneth, P.
770 Balvanera, K. A. Brauman, S. H. M. Butchart, K. M. A. Chan, L. A. Garibaldi, K. Ichii, J. Liu,
771 S. M. Subramanian, G. F. Midgley, P. Miloslavich, Z. Molnár, D. Obura, A. Pfaff, S. Polasky, A.
772 Purvis, J. Razzaque, B. Reyers, R. Roy Chowdhury, Y. J. Shin, I. J. Visseren-Hamakers, K. J.
773 Willis, and C. N. Zayas (eds.). IPBES secretariat, Bonn, Germany. 56 pages.

774 Jiang C-Y, Sheng X-F, Qian M, Wang Q-Y (2008) Isolation and characterization of a heavy metal-
775 resistant *Burkholderia* sp. from heavy metal-contaminated paddy field soil and its potential in
776 promoting plant growth and heavy metal accumulation in metal-polluted soil. *Chemosphere*
777 72:157-164. <https://doi.org/10.1016/j.chemosphere.2008.02.006>

778 Kardol P, Wardle DA (2010) How understanding aboveground-belowground linkages can assist
779 restoration ecology. *Trends Ecol Evol* 25:670-679. <https://doi.org/10.1016/j.tree.2010.09.001>

780 Kidd P, Mench M, Álvarez-López V, et al. (2015) Agronomic practices for improving gentle remediation
781 of trace element-contaminated soils. *Int J Phytorem* 17:1005-1037. <https://doi.org/10.1080.15226514.2014.1003788>

783 Kolbas A, Kidd P, Guinberteau J, Jaunatre R, Herzig R, Mench M (2015) Endophytic bacteria take the
784 challenge to improve Cu phytoextraction by sunflower. *Environ Sci Pollut Res* 22:5370-5382.
785 <https://doi.org/10.1007/s11356-014-4006-1>

786 Kowalchuk GA, Stienstra AW, Stephen JR, et al. (2000) Changes in the community structure of ammonia
787 oxidizing bacteria during secondary succession of calcareous grasslands. *Environ Microbiol*
788 2:99-110

789 Kumpiene J, Guerri G, Landi L, et al. (2009) Microbial biomass, respiration and enzyme activities after *in*
790 *situ* aided phytostabilization of a Pb- and Cu-contaminated soil. *Ecotoxicol Environ Saf* 72:115-
791 119. <https://doi.org/10.1016/j.ecoenv.2008.07.002>

792 Kumpiene J, Bert V, Dimitriou I, Eriksson J, Friesl-Hanl W, Galazka F, Herzig R, Janssen JO, Kidd P,
793 Mench M, Müller I, Neu S, Oustriere N, Puschenreiter M, Renella G, Roumier PH, Siebielec G,
794 Vangronsveld J, Manier N (2014) Selecting chemical and ecotoxicological test batteries for risk
795 assessment of trace element-contaminated soils (phyto)managed by Gentle Remediation Options
796 (GRO). *Sci Total Environ* 496:510-522. <https://doi.org/10.1016/j.scitotenv.2014.06.130>

797 Kumpiene J, Renella G, Denys S, Marschner B, Mench M, Adriaensen K, Vangronsveld J, Friesl-Hanl
798 W, Puschenreiter M (2017) Review and evaluation of methods for determining the bioavailable
799 fraction of trace elements in soils. *Pedosphere* 27:389-406. [https://doi.org/10.1016/S1002-](https://doi.org/10.1016/S1002-0160(17)60337-0)
800 [0160\(17\)60337-0](https://doi.org/10.1016/S1002-0160(17)60337-0)

801 Kuppens T, Van Dael M, Vanreppelen K, Thewys T, Yperman J, Carleer R, Schreurs S, Van Passel S
802 (2015) Techno-economic assessment of fast pyrolysis for the valorization of short rotation
803 coppice cultivated for phytoextraction. *J Cleaner Production* 88:336-344. <https://doi.org/10.1016/j.jclepro.2014.07.023>

804

805 Ju W, Jin X, Liu L, Shen G, Zhao W, Duan C, Fang L (2020) Rhizobacteria inoculation benefits nutrient
806 availability for phytostabilization in copper contaminated soil: Drivers from bacterial community
807 structures in rhizosphere. *Appl Soil Ecol Applied* 150:103450. <https://doi.org/10.1016/j.apsoil.2019.103450>

808

809 Lacalle RG, Gómez-Sagasti MT, Artetxe U, Garbisu C, Becerril JM (2018) *Brassica napus* has a key role
810 in the recovery of the health of soils contaminated with metals and diesel by nano-
811 rhizoremediation. *Sci Total Environ* 618:347-356. <https://doi.org/10.1016/j.scitotenv.2017.10.334>

812 Li H, Wang XG, Liang C, et al. (2015). Aboveground-belowground biodiversity linkages differ in early
813 and late successional temperate forests. *Sci Rep* 5:12234. <https://doi.org/10.1038/srep12234>

814 Ma TT, Teng Y, Luo YM, Christie P. (2013) Legume-grass intercropping phytoremediation of phthalic
815 acid esters in soil near an electronic waste recycling site: a field study. *Int J Phytorem* 15:154-
816 167. <https://doi.org/10.1080/15226514.2012.687016>.

817 Ma Y, Oliveira RS, Nai F, Rajkumar M, Luo Y, Rocha I, Freitas H (2015) The hyperaccumulator *Sedum*
818 *plumbizincicola* harbors metal-resistant endophytic bacteria that improve its phytoextraction
819 capacity in multi-metal contaminated soil. *J Environ Manage* 156:62-69. <https://doi.org/10.1016/j.jenvman.2015.03.024>

820

821 Madejón E, de Mora AP, Felipe E, et al. (2006) Soil amendments reduce trace element solubility in a
822 contaminated soil and allow regrowth of natural vegetation. *Environ Pollut* 139:40-52. <https://doi>
823 [10.1016/j.envpol.2005.04.034](https://doi.org/10.1016/j.envpol.2005.04.034)

824 Magurran AE (2004) *Measuring biological diversity*, 2nd ed. Blackwell Science Ltd, Oxford

825 Malik ZH, Ravindran KC, Sathiyara G (2017) Phytoremediation: a novel strategy and eco-friendly green
826 technology for removal of toxic metals. *Int J Agric Environ Res* 3:1-18

827 Mann RM, Vijver MG, Peijnenburg WJGM (2011) Metals and metalloids in terrestrial systems:
828 Bioaccumulation, biomagnification and subsequent adverse effects. In: Sánchez-Bayo F, van den
829 Brink PJ, Mann R (eds) *Ecological impacts of toxic chemicals*. Bentham Science Publishers, pp
830 43-62

831 Massoura ST, Echevarria G, Leclerc-Cessac E, Morel JL (2004) Response of excluder, indicator, and
832 hyperaccumulator plants to nickel availability in soils. *Australian J Soil Res* 42:933-938.
833 [https://doi 10.1071/SR03157](https://doi.org/10.1071/SR03157)

834 Mastretta C, Taghavi S, van der Lelie D, Mengoni A, Galardi F, Gonnelli C, Barac T, Boulet J, Weyens
835 N, Vangronsveld J (2009) Endophytic bacteria from seeds of *Nicotiana tabacum* can reduce
836 cadmium phytotoxicity. *Int J Phytoremediat* 11:251-267. <https://doi>
837 [10.1080/15226510802432678](https://doi.org/10.1080/15226510802432678)

838 Meglouli H, Fontaine J, Verdin A, Magnin-Robert M, Tisserant B, Hijri M, Lounès-Hadj Saharaoui A
839 (2019) Aided phytoremediation to clean up dioxins/furans-aged contaminated soil: Correlation
840 between microbial communities and pollutant dissipation. *Microorganisms* 7:523.
841 [doi:10.3390/microorganisms7110523](https://doi.org/10.3390/microorganisms7110523)

842 Mench M, Schwitzguébel JP, Schröder P, Bert V, Gawronski S, Gupta S (2009) Assessment of successful
843 experiments and limitations of phytotechnologies: contaminant uptake, detoxification and
844 sequestration, and consequences for food safety. *Environ Sci Pollut Res* 16:876-900. <https://doi>
845 [10.1007/s11356-009-0252-z](https://doi.org/10.1007/s11356-009-0252-z)

846 Mench MJ, Dellise M, Bes CM, et al. (2018) Phytomanagement and remediation of Cu-contaminated
847 soils by high yielding crops at a former wood preservation site: Sunflower biomass and ionome.
848 *Front Ecol Evol* 6:123. [https://doi 10.3389/fevo.2018.00123](https://doi.org/10.3389/fevo.2018.00123)

849 Mouchet MA, Villéger S, Mason NW, et al. (2010) Functional diversity measures: an overview of their
850 redundancy and their ability to discriminate community assembly rules. *Funct Ecol* 24:867-876.
851 [https://doi 10.1111/j.1365-2435.2010.01695.x](https://doi.org/10.1111/j.1365-2435.2010.01695.x)

852 Naeem S, Chazdon R, Duffy JE, Prager C, Worm B (2016) Biodiversity and human well-being: an
853 essential link for sustainable development. *Proc R Soc B* 283:20162091. [http://doi](http://doi.org/10.1098/rspb.2016.2091)
854 [10.1098/rspb.2016.2091](http://doi.org/10.1098/rspb.2016.2091)

855 Olguin EJ, Sanchez-Galvan G, Melo FJ, Hernandez VJ, Gonzalez-Portela RE (2017) Long-term
856 assessment at field scale of Floating Treatment Wetlands for improvement of water quality and
857 provision of ecosystem services in a eutrophic urban pond. *Sci Total Environ* 584:561-571.
858 [https://doi 10.1016/j.scitotenv.2017.01.072](https://doi.org/10.1016/j.scitotenv.2017.01.072)

859 Pandey VC, Bajpai O, Singh N (2016) Energy crops in sustainable phytoremediation. *Renewable*
860 *Sustainable Energy Rev* 54:58-73. [https://doi 10.1016/j.rser.2015.09.078](https://doi.org/10.1016/j.rser.2015.09.078)

861 Pandey VC, Baudh K (2018) Phytomanagement of Polluted Sites - Market Opportunities in Sustainable
862 Phytoremediation. Elsevier, pp. 626. [https://doi 10.1016/C2017-0-00586-4](https://doi.org/10.1016/C2017-0-00586-4)

863 Pardo T, Clemente R, Epelde L, Garbisu C, Bernal MP (2014) Evaluation of the phytostabilisation
864 efficiency in a trace elements contaminated soil using soil health indicators. *J Hazard Materials*
865 268:68-76. [https://doi 10.1016/j.jhazmat.2014.01.003](https://doi.org/10.1016/j.jhazmat.2014.01.003)

866 Parraga-Aguado I, Querejeta JI, Gonzalez-Alcaraz M-N, et al. (2014) Usefulness of pioneer vegetation
867 for the phytomanagement of metal(loid)s enriched tailings: Grasses vs. shrubs vs. trees. *J*
868 *Environ Manage* 133:51-58. [https://doi 10.1016/j.jenvman.2013.12.001](https://doi.org/10.1016/j.jenvman.2013.12.001)

869 Pat-Espadas AM, Loredó Portales R, Amabilis-Sosa LE, Gómez G, Vidal G (2018) Review of
870 constructed wetlands for acid mine drainage treatment. *Water* 10:1685. [https://doi](https://doi.org/10.3390/w10111685)
871 [10.3390/w10111685](https://doi.org/10.3390/w10111685)

872 Paul, ALD, Erskine PD, van der Ent A (2018) Metallophytes on Zn-Pb mineralised soils and mining
873 wastes in Broken Hill, NSW, Australia. *Australian J Bot* 66:124-133. [https://doi](https://doi.org/10.1071/BT17143)
874 [10.1071/BT17143](https://doi.org/10.1071/BT17143)

875 Peterson LR, Trivett V, Baker AJM et al. (2003) Spread of metals through an invertebrate food chain as
876 influenced by a plant that hyperaccumulates nickel. *Chemoecology* 13:103-108. [https://doi](https://doi.org/10.1007/s00049-003-0234-4)
877 [10.1007/s00049-003-0234-4](https://doi.org/10.1007/s00049-003-0234-4)

878 Phillips R, Finzi A, Bernhardt E (2011) Enhanced root exudation induces microbial feedbacks to N
879 cycling in a pine forest under long-term CO₂ fumigation. *Ecol Lett* 14:187-194. [https://doi](https://doi.org/10.1111/j.1461-0248.2010.01570.x)
880 [10.1111/j.1461-0248.2010.01570.x](https://doi.org/10.1111/j.1461-0248.2010.01570.x)

881 Prins CN, Hantziis LJ, Valdez-Barillas JR, Cappa JJ, Fakra SC, Milano de Tomasel C, Wall DH, Pilon-
882 Smits EAH (2019) Getting to the root of selenium hyperaccumulation – Localization and
883 speciation of root selenium and its effects on nematodes. *Soil Systems* 3:47. [https://doi](https://doi.org/10.3390/soilsystems3030047)
884 [10.3390/soilsystems3030047](https://doi.org/10.3390/soilsystems3030047)

885 Pulighe G, Bonati G, Colangeli M, Morese, MM, Traverso L, Lupia F, Khawaja C, Janssen R, Fava F
886 (2019) Ongoing and emerging issues for sustainable bioenergy production on marginal lands in
887 the Mediterranean regions. *Renewable Sustainable Energy Rev* 103:58-70. [https://doi:](https://doi.org/10.1016/j.rser.2018.12.043)
888 [10.1016/j.rser.2018.12.043](https://doi.org/10.1016/j.rser.2018.12.043)

889 Rajkumar M, Prasad MNV, Swaminathan S, et al. (2013) Climate change driven plant-metal-microbe
890 interactions. *Environ Int* 53:74-86. [https://doi 10.1016/j.envint.2012.12.009](https://doi.org/10.1016/j.envint.2012.12.009)

891 Reeves RD, Baker AJM, Jaffré T, Erskine PD, Echevarria G, van der Ent A (2017a) A global database for
892 plants that hyperaccumulate metal and metalloid trace elements. *New Phytologist* 218:407-411.
893 [https://doi 10.1111/nph.14907](https://doi.org/10.1111/nph.14907)

894 Reeves RD, Baker AJM, Jaffre T, Erskine PD, Echevarria G, van der Ent A (2017b) A global database for
895 plants that hyperaccumulate metal and metalloid trace element. *New Phytologist* 218:407-411.
896 [https://doi 10.1111/nph.14907](https://doi.org/10.1111/nph.14907)

897 Reuter H, Hölker F, Middelhoff U, et al. (2005) The concepts of emergent and collective properties in
898 individual-based models - Summary and outlook of the Bornhöved case studies. *Ecol Modell*
899 186:489-501. [https://doi 10.1016/j.ecolmodel.2005.02.014](https://doi.org/10.1016/j.ecolmodel.2005.02.014)

900 Risueño Y, Petri C, Conesa HM (2020) The importance of edaphic niches functionality for the
901 sustainability of phytomanagement in semiarid mining impacted ecosystems. *J Environ Manage*
902 266:110613. [https://doi 10.1016/j.jenvman.2020.110613](https://doi.org/10.1016/j.jenvman.2020.110613)

903 Robinson BH, Banuelos G, Conesa HM, Evangelou MWH, Schulin R (2009) The phytomanagement of
904 trace elements in soil. *Crit Rev Plant Sci* 28:240-266. [https://doi 10.1080/07352680903035424](https://doi.org/10.1080/07352680903035424)

905 Robinson BH, McIvor I (2013) Phytomanagement of contaminated sites using poplars and willows. In:
906 Leung DWM (ed) *Recent advances towards improved phytoremediation of heavy metal*
907 *pollution*, Bentham Science, Arab Emirates, pp. 119-133

908 Rosenkranz T, Hipfinger C, Ridard C, Puschenreiter M (2019) A nickel phytomining field trial using
909 *Odontarrhena chalcidica* and *Noccaea goesingensis* on an Austrian serpentine soil. *J Environ*
910 *Manage* 242:522-528. [https://doi 10.1016/j.jenvman.2019.04.073](https://doi.org/10.1016/j.jenvman.2019.04.073)

911 Sandifer PA, Sutton-Grier AE, Ward BP (2015) Exploring connections among nature, biodiversity,
912 ecosystem services, and human health and well-being: Opportunities to enhance health and
913 biodiversity conservation. *Ecosystem Services* 12:1-15. [https://doi 10.1016/j.ecoser.2014.12.007](https://doi.org/10.1016/j.ecoser.2014.12.007)

914 Schoeneberger M, Bentrup G, de Gooijer H, Soolanayakanahally R, Sauer T, Brandle J, Zhou X, Current
915 D (2012) Branching out: Agroforestry as a climate change mitigation and adaptation tool for
916 agriculture. *J Soil Water Conservation* 67:128A-136A. [https://doi 10.2489/jswc.67.5.128A](https://doi.org/10.2489/jswc.67.5.128A)

917 Schröder P, Navarro-Aviñó J, Azaizeh H, Goldhirsh AG, DiGregorio S, Komives T, Langergraber G,
918 Lenz A, Maestri E, Memon AR, Ranalli A, Sebastiani L, Smrcek S, Vanek T, Vuilleumier S,
919 Wissing F (2007) Using phytoremediation technologies to upgrade waste water treatment in
920 Europe. *Env Sci Pollut Res* 14:490-497

921 Séleck M, Bizoux JP, Colinet G, et al. (2013) Chemical soil factors influencing plant assemblages along
922 copper-cobalt gradients: implications for conservation and restoration. *Plant Soil* 373:455-469

923 Sessitsch A, Kuffner M, Kidd P, Vangronsveld J, Wenzel WW, Fallmann K, Puschenreiter M (2013) The
924 role of plant-associated bacteria in the mobilization and phytoextraction of trace elements in
925 contaminated soils. *Soil Biol Biochem* 60:182-194. [https://doi 10.1016/j.soilbio.2013.01.012](https://doi.org/10.1016/j.soilbio.2013.01.012)

926 Simek M, Elhottova D, Mench M, et al. (2017) Greenhouse gas emissions from a Cu-contaminated soil
927 remediated by *in situ* stabilization and phytomanaged by a mixed stand of poplar, willows, and
928 false indigo-bush. *Int J Phytorem* 19:976-984. [https://doi 10.1080/15226514.2016.1267706](https://doi.org/10.1080/15226514.2016.1267706)

929 Sun M, Fu D, Teng Y, et al. (2011) *In situ* phytoremediation of PAH-contaminated soil by intercropping
930 alfalfa (*Medicago sativa* L.) with tall fescue (*Festuca arundinacea* Schreb.) and associated soil
931 microbial activity. *J Soils Sediments* 11:980-989. [https://doi 10.1007/s11368-011-0382](https://doi.org/10.1007/s11368-011-0382)

932 Sura-de Jong M, Reynolds RJ, Richterova K, Musilova L, Hrochova I, Frantik T, Sakmaryova I, Strejcek
933 M, Cochran A, Staicu L, Cappa JJ, van der Lelie D, Pilon-Smits EAH (2015) Selenium
934 hyperaccumulators harbor a diverse endophytic bacterial community characterized by high
935 selenium resistance and plant growth promoting properties. *Frontiers in Plant Science* 6:113.
936 [https://doi 10.3389/fpls.2015.00113](https://doi.org/10.3389/fpls.2015.00113)

937 Touceda-González M, Prieto-Fernández A, Renella G, Giagnoni L, Sessitsch A, Brader G, Kumpiene J,
938 Dimitriou I, Eriksson J, Friesl-Hanl W, Galazka R, Janssen J, Mench M, Müller I, Neu S,
939 Puschenreiter M, Siebielec G, Vangronsveld J, Kidd PS (2017a) Microbial community structure
940 and activity in trace element contaminated soils phytomanaged by Gentle Remediation Options
941 (GRO). *Environ Pollut* 231:237-251. [http://doi 10.1016/j.envpol.2017.07.097](http://doi.org/10.1016/j.envpol.2017.07.097)

942 Touceda-González M, Álvarez-López V, Prieto-Fernández A, Rodríguez-Garrido B, Trasar-Cepeda C,
943 Mench M, Puschenreiter M, Quintela-Sabaris C, Macías-García F, Kidd PS (2017b) Aided
944 phytostabilisation reduces metal toxicity, improves soil fertility and enhances microbial activity
945 in Cu-rich mine tailings. *J Environ Manage* 186:301-313. [https://doi
10.1016/j.jenvman.2016.09.019](https://doi.org/10.1016/j.jenvman.2016.09.019)

947 Truyens S, Jambon I, Croes S, Janssen J, Weyens N, Mench M, Carleer R, Cuypers A, Vangronsveld J
948 (2014) The effect of long-term Cd and Ni exposure on seed endophytes of *Agrostis capillaris*
949 and their potential application in phytoremediation of metal-contaminated soils. *Int J Phytorem*
950 16:643-659. [https://doi 10.1080/15226514.2013.837027](https://doi.org/10.1080/15226514.2013.837027)

951 Thewys T, Witters N, Meers E, Vangronsveld J (2010) Economic viability of phytoremediation of a
952 cadmium contaminated agricultural area using energy maize. Part II: Economics of anaerobic
953 digestion of metal contaminated maize in Belgium. *Int J Phytorem* 12:663-679. [https://doi
10.1080/15226514.2010.493188](https://doi.org/10.1080/15226514.2010.493188)

955 van der Ent A, Baker AJM, Reeves RD, Pollard AJ, Henk Schat H (2013) Hyperaccumulators of metal
956 and metalloid trace elements: Facts and fiction. *Plant Soil* 362:319-334. [https://doi
10.1007/s11104-012-1287](https://doi.org/10.1007/s11104-012-1287)

958 van der Ent A, Baker AJM, Reeves RD, Pollard AJ, Schat H (2015a) A Commentary on “Toward a more
959 physiologically and evolutionarily relevant definition of metal hyperaccumulation in plants”.
960 Front Plant Sci 6:1-3. [https://doi 10.3389/fpls.2015.00554](https://doi.org/10.3389/fpls.2015.00554)

961 van der Ent A, Baker AJM., Reeves RD, Chaney RL, Anderson CWN, Meech JA, Erskine PD, Simonnot
962 M-O, Vaughan J, Morel JL, Echevarria G, Fogliani B, Rongliang Q, Mulligan DR (2015b)
963 Agromining: farming for metals in the future? Environ Sci Technol 49:4773-4780. [https://doi
964 10.1021/es506031u](https://doi.org/10.1021/es506031u)

965 Vane-Wright RI, Humphries CJ, Williams, PH (1991) What to protect - systematics and the agony of
966 choice. Biol Conservation 55:235-254

967 Vangronsveld J, Herzig R, Weyens N, et al. (2009) Phytoremediation of contaminated soils and
968 groundwater: lessons from the field. Environ Sci Pollut Res Int 16:765-794. [https://doi
969 10.1007/s11356-009-0213-6](https://doi.org/10.1007/s11356-009-0213-6)

970 Verkerk RHJ, Leather SR, Wright DJ (1998) The potential for manipulating crop-pest-natural enemy
971 interactions for improved insect pest management. Bull Entomol Res 88:493-501

972 Villaverde J, Láiz L, Lara-Moreno A, González-Pimentel JL, Morillo E (2019) Bioaugmentation of PAH-
973 contaminated soils with novel specific degrader strains isolated from a contaminated industrial
974 site. Effect of hydroxypropyl- β -cyclodextrin as PAH bioavailability enhancer. Front Microbiol 14
975 November 2019. [https://doi 10.3389/fmicb.2019.02588](https://doi.org/10.3389/fmicb.2019.02588)

976 Vincent Q, Auclerc A, Beguiristain T, Leyval C (2018) Assessment of derelict soil quality: Abiotic, biotic
977 and functional approaches. Sci Total Environ 614-614:990-1002. [https://doi
978 10.1016/j.scitotenv.2017.09.118](https://doi.org/10.1016/j.scitotenv.2017.09.118)

979 Wang K, Huang H, Zhu Z, et al. (2013) Phytoextraction of metals and rhizoremediation of PAHs in co-
980 contaminated soil by co-planting of *Sedum alfredii* with ryegrass or castor. Int J Phytorem
981 15:283-298. [https://doi 10.1080/15226514.2012.694501](https://doi.org/10.1080/15226514.2012.694501)

982 Wardle DA, Bardgett RD, Klironomos JN, et al. (2004) Ecological linkages between aboveground and
983 belowground biota. Science 304:1629-1633

984 Weyens N, Truyens S, Saenen E, Boulet J, Dupae J, Taghavi S, Lelie D, Carleer R, Vangronsveld J
985 (2011) Endophytes and their potential to deal with co-contamination of organic contaminants

986 (toluene) and toxic metals (nickel) during phytoremediation. *Int J Phytoremediation* 13:244-255.
987 [https://doi 10.1080/15226511003753920](https://doi.org/10.1080/15226511003753920)

988 Whiting SN, Reeves RD, Baker AJM (2002) Mining, metallophytes and land reclamation. *Min Environ*
989 *Manag* 10:11-16

990 Whiting SN, Reeves RD, Richards DG, et al. (2004) Research priorities for conservation of metallophyte
991 biodiversity and their potential for restoration and site remediation. *Rest Ecol* 12:106-116.
992 [https://doi 10.1111/j.1061-2971.2004.00367.x](https://doi.org/10.1111/j.1061-2971.2004.00367.x)

993 Xue K, Zhou J., Van Nostrand J D, Mench M, Bes C, Giagnoni L, Arenella M, Renella G. (2018)
994 Functional activity and functional gene diversity of a Cu-contaminated soil remediated by aided
995 phytostabilization using compost, dolomitic limestone and a mixed tree stand. *Environ Pollut*
996 242:229-238. [https://doi 10.1016/j.envpol.2018.06.057](https://doi.org/10.1016/j.envpol.2018.06.057)

997 Yan A, Wang Y, Tan SN, Yusof MLM, Ghosh S, Chen Z (2020) Phytoremediation: a promising
998 approach for revegetation of heavy metal-polluted land. *Front Plant Sci* 11:359. [https://doi](https://doi.org/10.3389/fpls.2020.00359)
999 [10.3389/fpls.2020.00359](https://doi.org/10.3389/fpls.2020.00359)

1000 Yang W, Zhao F, Wang Y, et al. (2020a) Differences in uptake and accumulation of copper and zinc by
1001 *Salix* clones under flooded *versus* non-flooded conditions. *Chemosphere* 241:125059. [https://doi](https://doi.org/10.1016/j.chemosphere.2019.125059)
1002 [10.1016/j.chemosphere.2019.125059](https://doi.org/10.1016/j.chemosphere.2019.125059)

1003 Yang C, Ho Y-H, Makita R, Inoue C, Chien M-F (2020b) A multifunctional rhizobacterial strain with
1004 wide application in different ferns facilitates arsenic phytoremediation. *Sci Total Environ*
1005 712:134504. [https://doi 10.1016/j.scitotenv.2019.134504](https://doi.org/10.1016/j.scitotenv.2019.134504)

1006 Zaidi S, Usmani S, Singh BR, Musarrat J (2006) Significance of *Bacillus subtilis* strain SJ-101 as a
1007 bioinoculant for concurrent plant growth promotion and nickel accumulation in *Brassica juncea*.
1008 *Chemosphere* 64:991-997. [https://doi 10.1016/j.chemosphere.2005.12.057](https://doi.org/10.1016/j.chemosphere.2005.12.057)

1009 Zeng P, Guo Z, Xiao X, Peng C, Huang B, Feng WL (2019a) Complementarity of co-planting a
1010 hyperaccumulator with three metal(loid)-tolerant species for metal(loid)-contaminated soil
1011 remediation. *Ecotoxicol Environ Safety* 169:306-315. [https://doi 10.1016/j.ecoenv.2018.11.017](https://doi.org/10.1016/j.ecoenv.2018.11.017)

- 1012 Zeng P, Guo ZH, Xiao XY, Peng C, Feng WL, Xin LQ, Xu Z (2019b) Phytoextraction potential of *Pteris*
1013 *vittata* L. co-planted with woody species for As, Cd, Pb and Zn in contaminated soil. *Sci Total*
1014 *Environ* 650:594-603. [https://doi 10.1016/j.scitotenv.2018.09.055](https://doi.org/10.1016/j.scitotenv.2018.09.055)
- 1015 Zhang XB, Liu P, Yang YS, Chen WR (2007) Phytoremediation of urban wastewater by model wetlands
1016 with ornamental hydrophytes. *J Environ Sci (China)* 19:902-909. [https://doi 10.1016/S1001-](https://doi.org/10.1016/S1001-0742(07)60150-8)
1017 [0742\(07\)60150-8](https://doi.org/10.1016/S1001-0742(07)60150-8)
- 1018

1019 **Table 1:** Ten examples of effects of biodiversity under phytomanagement.

Plant species	Contaminants	Main finding	Reference
<i>Pteris vittata</i> co-planted with <i>Morus alba</i> and <i>Broussonetia papyrifera</i>	As, Cd, Pb and Zn	Co-planting alleviated toxicity and improved phytoextraction	Zeng et al. 2019b
<i>Pteris vittata</i> co-planted with <i>Arundo donax</i> , <i>Morus alba</i> and <i>Broussonetia papyrifera</i>	As, Cd, Pb, and Zn	Co-planting enhanced <i>P. vittata</i> growth and metal(oid) accumulation, and improve soil quality	Zeng et al. 2019a
Co-planting <i>Sedum alfredii</i> with <i>Lolium perenne</i> or <i>Ricinus communis</i>	Metals and PAHs	Co-planting <i>S. alfredii</i> with ryegrass or castor enhanced pyrene and anthracene dissipation	Wang et al. 2013
Intercropping: <i>Medicago sativa</i> with <i>Festuca arundinacea</i>	PAHs	Removal PAHs under intercropping was higher than under monoculture	Sun et al. 2011
<i>Odontarrhena chalcidica</i> or <i>Noccaea goesingensis</i> co-planted with <i>Lotus corniculatus</i>	Ni	Intercropping with <i>L. corniculatus</i> tended to decrease the shoot biomass of both species	Rosenkranz et al. 2019
The grass <i>Piptatherum miliaceum</i> , the shrub <i>Helichrysum decumbens</i> , and the trees <i>Pinus halepensis</i> and <i>Tetraclinis articulata</i>	Metal(loid)s	A diverse set of plant species with contrasting life forms may result in a more efficient employment of water resources and a higher biodiversity not only in relation to flora but also soil microbes	Parraga-Aguado et al. 2014
<i>Medicago sativa</i> , <i>Lolium perenne</i> and <i>Festuca arundinacea</i>	Phthalic acid esters (PAEs)	Intercropping with the three species was the most effective treatment for PAEs removal	Ma et al. 2013
Monocultures and polycultures of <i>Festuca arundinacea</i> , <i>Medicago sativa</i> and <i>Salix miyabeana</i>	Ag, As, Cd, Cr, Cu, Pb, Se and Zn	Co-cropping with the three species was the most robust scenario for remediation of multiple trace element contaminated soil	Desjardins et al. 2018
Grapevine was grown in monocropping, intercropping with <i>Paspalum plicatulum</i> and intercropping with <i>Axonopus affinis</i>	Cu	Intercropping with <i>P. plicatulum</i> and <i>A. affinis</i> was efficient in promoting the growth of grapevines at moderate and low levels of Cu contamination by reducing its bioavailability	De Conti et al. 2019
Co-cropping of <i>Festuca arundinacea</i> , <i>Salix miyabeana</i> and <i>Medicago sativa</i>	Trace elements and persistent organic pollutants (POPs)	The crops cultivated in pairs retained rhizosphere microbiome bacteria involved in plant growth promotion, POP tolerance and degradation, and improved nutrient acquisition	Brereton et al. 2020

1020

1021

1022 **Figure legends**

1023 **Figure 1:** Evolution from phytoremediation to phytomanagement.

