

**STUDY OF GENETIC DIVERSITY OF WHEAT  
(*Triticum aestivum* L.) GENOTYPES UNDER  
TERAI AGROCLIMATIC CONDITIONS**

**Thesis**

Submitted to the

Uttar Banga Krishi Viswavidyalaya

In partial fulfilment of the requirement for the degree

of

**MASTER OF SCIENCE AGRICULTURE**

in

**GENETICS AND PLANT BREEDING**

By

**SOURIK PODDAR**

**A-2019-036-M**



**DEPARTMENT OF GENETICS AND PLANT BREEDING**

**FACULTY OF AGRICULTURE**

**UTTAR BANGA KRISHI VISWAVIDYALAYA**

**PUNDIBARI-736165**

**2021**



**DEDICATED TO**

**MY PARENTS**

**Mr. Gopal Poddar**

**&**

**Mrs. Shapma Poddar**

**AND**

**MY CHAIRMAN**

**Dr. Saikat Das**



**UTTAR BANGA KRISHI VISWAVIDYALAYA**

**Faculty of Agriculture**

**Department of Genetics and Plant Breeding**

P.O.: Pundibari, Cooch Behar, West Bengal-736165, India.

**From:**

Dr. Saikat Das  
Associate professor and Chairman,  
Advisory Committee

Phone (M): +919434939334  
Email: saikat.ubkv@gmail.com

---

**CERTIFICATE**

This is to certify that the work recorded in the thesis entitled “**Study of genetic diversity of wheat (*Triticum aestivum* L.) genotypes under terai agroclimatic conditions**” submitted by **Mr. Sourik Poddar (A-2019-036-M)** Master of Science (Agriculture) in Genetics and Plant Breeding, Uttar Banga Krishi Viswavidyalaya is a genuine and original one carried out under my personal supervision and guidance. The results of the investigation reported in the thesis have not so far been submitted for any other Degree or Diploma. The assistance and help received from various sources during the courses of investigation is being duly acknowledged.

---

(SAIKAT DAS)

Chairman

Advisory Committee



# UTTAR BANGA KRISHI VISWAVIDYALAYA

Faculty of Agriculture

Department of Genetics and Plant Breeding

P.O.: Pundibari, Cooch Behar, West Bengal-736165, India.

**From:**

Dr. Saikat Das  
Associate professor and Chairman,  
Advisory Committee

Phone (M): +919434939334  
Email: saikat.ubkv@gmail.com

---

*CERTIFICATE*

Certificated that the thesis entitled “**Study of genetic diversity of wheat (*Triticum aestivum* L.) genotypes under terai agroclimatic conditions**” submitted by **Mr. Sourik Poddar** bearing Registration Number **A-2019-036-M** towards partial fulfilment of the requirement for the award of Master Degree from the Department of Genetics and Plant Breeding, Faculty of Agriculture under Uttar Banga Krishi Viswavidyalaya has been checked against plagiarism through Urkund software on 04-08-2021 and the similarity index has been achieved as 3% which is below the maximum tolerable range as per stipulation of this Viswavidyalaya. The thesis of **Mr. Sourik Poddar** may be accepted for the award of the **Master Degree in M.Sc. (Agriculture)** of Uttar Banga Krishi Viswavidyalaya.

.

Place:

Date:

---

(SAIKAT DAS)

Chairman

Advisory Committee

**APPROVAL OF THE EXAMINERS FOR THE AWARD OF THE  
DEGREE OF MASTER OF SCIENCE (AGRICULTURE) IN  
GENETICS AND PLANT BREEDING**

We, the undersigned, having been satisfied with the performance of Mr. Sourik Poddar, in the viva-voce examination on the Final evaluation of the thesis, conducted today,     /     / 2021, recommended that the thesis be accepted for the award of the degree of Master of Science (Agriculture) in Genetics and Plant Breeding.

<b>S.N</b>	<b>Name</b>	<b>Signature</b>
<b>1.</b>	<b>Dr. Saikat Das</b> Chairman Advisory committee Department of Genetics and Plant breeding	
<b>2.</b>	External Examiner	
<b>3.</b>	<b>Prof. Suwendu Kumar Roy</b> Member Advisory committee Head of the Department, GPB	
<b>4.</b>	<b>Dr. Rupsanatan Mandal</b> Member Advisory committee	
<b>5.</b>	<b>Dr. Satyajit Hembram</b> Member Advisory committee	
<b>6.</b>	<b>Dr. Puspendu Dutta</b> Member Advisory committee	
<b>7.</b>	<b>Prof Prabhat Kumar Pal</b> Dean (Actg.) Post Graduate Studies	

## **ACKNOWLEDGEMENT.**

This thesis is the end of my journey in obtaining my M.Sc. This thesis has been kept on track and seen through to completion with the support and encouragement of numerous people including my family members, respected teachers, well-wishers, my friends and colleagues. At the end of the thesis, I would like to thank all those people who gave immense encouragement and unforgettable experience for me.

At this moment of accomplishment, first of all I pay homage to my Chairman, **Dr. Saikat Das**. This work would not have been possible without his guidance, support, affectionate attitude, healthy criticism and encouragement. Under his guidance I have successfully overcome many difficulties and learned a lot. Thank you doesn't seem enough, but it is said with appreciation and respect to you for your support, encouragement, care, and understanding.

I gratefully acknowledge my advisory committee members, **Prof. S.K. Roy**, Head of the Department of Genetics and Plant Breeding, **Dr. Rupsanatan Mondal**, Assistant Professor of Department of Genetics and Plant Breeding, **Dr. Puspendu Dutta**, Assistant Professor of Department of Seed Science and Technology, and **Dr. S. Hembram**, Assistant professor of Department of Plant Pathology for their understanding, encouragement and personal attention which have provided good and smooth basis of my M.Sc. tenure.

I owe my special thanks to **Prof. Suvendu Kumar Roy**, Head of the Department of Genetics and Plant Breeding, Faculty of Agriculture, UBKV for understanding, support, and encouragement. I am grateful to University authority of UBKV, Cooch Behar, West Bengal, specifically **Dean, Postgraduate studies; Dean, Faculty of Agriculture** and **Director of Farm**, for providing all logistics and official support.

I would also like to express my sincere gratitude and unforgettable indebtedness to **International Maize and Wheat Improvement Center (CIMMYT), Mexico** for providing the genotypes under the 19HTWYT nursery to carry out my research.

I am thankful to all the teachers of Department of Genetics and Plant Breeding, **Prof. Arup Sarkar, Dr. Moumita Chakraborty, Dr. Lakshmi Hijam, Dr. Avijit Kundu, and Dr. Soumendra Chakraborty**, for their help, constant attention, useful suggestions and motivation during my post graduate studies.

I would like to express my appreciation and thankfulness to all the **non-teaching staff** of Dept. of GPB, **Mr. Manmatha Hansda** and field worker, **Bishu da** and all others for their kind help during my field works in entire of study.

I owe great deal of appreciation, respect and gratitude to **Mr. Sahanob Nath** for helping me to carry out major part of my research work about data collection and suggestions at his best, and also taking care of me as his little brother. This thesis would not have been obtained without a close collaboration with him.

I would also like to give a vote of thanks to my fellow classmates, **Subir, Soumayayan, Santanu, Vivekananda, Rakesh, Shivani, Kavita** and **Moumita** for their constant support and encouragement which made this journey easy and entertaining.

I would like to thank my beloved seniors, **Debajoti da, Anjan da, Bijaya di, Kajol di, Shiva da, Basid da** and many more for their constant support and encouragement.

My special thanks to **Subha di, Kuntal da, Pallabi di, Nandita mam** and **Monish da**, for tolerating my immature behaviour, mischiefs and still being so loving and caring to me throughout this whole journey.

A very special vote of thanks to **Prof. A.K. Choudhury**, Head of Department of Plant Pathology, UBKV for being a constant support, guide and source of encouragement since my B.Sc. days. Thank you for encouraging me to study in **Uttar Banga Krishi Vishwavidyalaya** and watching each step of mine and healthy criticisms that led me to learn a lot and push me to go out of my comfort zone.

This list is incomplete without mentioning my friends for life, **David, Mukul, Sonali, Ashish, Ananay, Jasvir, Nisha, Bhawna, Neeraj, Bharat, Vishu, Kriva, Ankur, Banashri, Smita, Nilahit, Anupam, Dipankar and Subham** for their constant support, understanding, moral support and inspirations.

A very special and loving thanks to **Mr. Bittu Mandal**, for always standing with me in ups and downs in this journey, understanding me, motivating me and taking care of me. This journey would have been different if his support was absent.

Last but not the least, I would like to thank the **Almighty** and my **Parents, Mr. Gopal Poddar and Mrs Shampa Poddar**, my elder brother, **Mr. Sagnik Poddar**, without whom this would be impossible for me to achieve whatever I have achieved till date. I am eternally grateful to them.

Besides this, several people have knowingly and unknowingly helped me in the successful completion of this M.Sc. tenure. I must emphasise that I am fully responsible for any shortcoming that remains.

**Date:** \_\_\_\_\_

**Place:** Pundibari, Cooch Behar.

Signature

**(SOURIK PODDAR)**

## CONTENT

<b>Chapter No.</b>	<b>Particulars</b>	<b>Page No.</b>
<b>1.</b>	<b>Introduction</b>	1-3
<b>2.</b>	<b>Review of Literature</b>	4-12
	2.1 Genetic Diversity	4
	2.2 Physiological study	7
	2.3 Study on Spot Blotch Resistance	10
<b>3.</b>	<b>Materials and Methods</b>	13-26
<b>4.</b>	<b>Result and Discussion</b>	27-56
	4.1. Study on Morpho-phenetic traits	27
	4.2. Study on physiological characters.	34
	4.3. Study on disease (Spot blotch) resistance.	43
	4.4. Genetic divergence analysis ( $D^2$ analysis)	47
	4.5. Principal Component Analysis	50
	4.6. Hierarchical clustering of 50 genotypes based on Euclidean distance	52
	4.7. Correlation analysis	55
<b>5.</b>	<b>Summary and Conclusion</b>	57-59
<b>6.</b>	<b>Reference</b>	61-69
	<b>Annexure</b>	I-X
	<b>VITA</b>	XI



## LIST OF TABLES

<b>Table No.</b>	<b>Title</b>	<b>Page No.</b>
Table 3.1.	List of wheat genotypes evaluated during 2020-2021	14
Table 3.2.	Meteorological data pertaining to the period of experimentation	15
Table 3.3.	Experimental details	16
Table 4.1.	Analysis of variance of morpho phenetic traits	28
Table 4.2.	Mean values of morpho phenetic traits	29-30
Table 4.3.	Genetic parameters for different morpho-phenetic characters of 50 genotypes of wheat	33
Table 4.4a.	Two-way Analysis of variance of Chlorophyll Index values	34
Table 4.4b.	Statistical parameters of Chlorophyll Index values	34
Table 4.4c.	Two-way mean table for Chlorophyll Index under different growth stages	35
Table 4.5.	AUCIPC values pertaining to different genotypes under study	38
Table 4.6a.	Two-way Analysis of variance for Canopy Temperature Depression under different growth stages	39
Table 4.6b.	Statistical parameters for Canopy Temperature Depression	39
Table 4.6c.	Two-way mean table for Canopy Temperature Depression	40
Table 4.7.	Correlation Matrix between CT, CTD, GY and BY	42
Table 4.8a.	Two-way Analysis of variance for Disease severity (%) under different growth stages	43
Table 4.8b.	Statistical parameters for Disease severity (%)	43
Table 4.8c.	Two-way mean table for Disease severity (%) under different growth stages	44
Table 4.9.	AUDPC values pertaining to different genotypes under study	46
Table 4.10.	Distribution of 50 wheat genotypes into different clusters.	47
Table 4.11.	Average intra (diagonal) and inter (off diagonal) cluster $D^2$ values of 50 genotypes of wheat	48
Table 4.12.	Cluster mean for 15 characters of wheat genotypes	49
Table 4.13.	Summary of the contribution of the different principal components to variability	50
Table 4.14.	Five principal components along with their factor loadings	51
Table 4.15.	Hierarchical clustering of 50 genotypes of wheat	52
Table 4.16.	Correlation Matrix between 15 characteristics of wheat	56

### LIST OF FIGURES

<b>Figure No.</b>	<b>Title of the Figure</b>	<b>Page No.</b>
Fig 1.	Graphical representation of Chlorophyll Index	36
Fig 2.	Graphical representation of Canopy Temperature Depression	41
Fig 3.	Screen plot analysis of PCA	50
Fig 4.	PCA Biplot graph showing 50 genotypes by points (G-1 to G-50) and trait vectors as green arrow	52
Fig 5.	Hierarchical clustering of 50 genotypes of wheat based on Euclidean distance	54

**ANNEXURE**

<b>SL. NO.</b>	<b>Description</b>	<b>Page No.</b>
A	Table of Chlorophyll Index at four growth stages and Area Under Chlorophyll Index Progress Curve (AUCIPC)	I-III
B	Table of Canopy Temperature Depression (CTD) at two growth stages.	IV-VI
C	Table of Disease Severity % at four growth stages with Area Under Disease Progress Curve (AUDPC).	VII-X

## ABSTRACT

<b>Name</b>	Sourik Poddar
<b>Registration No</b>	A-2019-036-M
<b>Department</b>	Genetics and Plant Breeding
<b>Advisor</b>	Dr. Saikat Das, Associate Professor
<b>Thesis Title</b>	"Study of Genetic Diversity of Wheat ( <i>Triticum aestivum</i> L.) Genotypes Under Terai Agroclimatic Conditions"
<b>University</b>	Uttar Banga Krishi Viswavidyalaya

Rice-wheat cropping is the major cropping system on 13.5 mha in the Indo-Gangetic plains (IGP) (Pathak *et al.*, 2003; Panigrahy *et al.*, 2011) producing about 50% of the total food grain and feeding 40% of India population (Gupta *et al.*, 2016). Now, this rice-wheat cropping system has started showing declining trends in marginal yield due to 'heat stress' problem in last decade. Early onset of significantly higher temperature coincided with wheat grain filling stage especially in Indo-Gangetic plains leading to terminal heat stress and reduction in yield. The present investigation has been carried out with 50 advanced genotypes of wheat targeted for heat stressed areas to assess the genetic diversity of wheat genotypes for yield and yield components along with physiological attributes and also to screen the genotypes against prevailing diseases of this area.

Among the morpho-phenetic traits, only days to heading, plant height, awn length, spike length, spikelet per spike, 1000 grain weight and biological yield showed significant variation between genotypes. AUCIPC value calculated on the basis of Chlorophyll Index (CI), was highest in ENTRY 1 which indicated higher retention of chlorophyll at maturity. Canopy Temperature Depression (CTD) showed higher value for 13 genotypes at later stages which signified high physiological efficiency for these genotypes. Correlation study indicated cooler canopy temperature leads to higher grain yield and increased biomass. In terms of spot blotch resistance, no genotype was found either moderately resistant (MR) or resistant (R) category.

D<sup>2</sup> analysis showed significant diversity between genotypes and it divided 50 genotypes into 5 clusters. In terms inter cluster distance value, most divergent cluster was found as cluster I and V. Five principal components (PCs) showed Eigen value >1.00, which accounted for 69.03% of cumulative proportion of variance. Biplot curve showed, in PC 1 characters such as BY, PH, GPS, AUCIPC had high positive loadings while AUDPC and CTD had negative loadings. In PC 2, GY and HI had high positive loadings while DF had negative loading. Also, strong positive correlation was found between GY, BY, GPS while negative association between GY and DF, CTD and PH.

The correlation analysis among the 15 characters showed that only eight traits (GER, PH, AL, SL, GPS, AUCIPC, BY and HI) were positively associated with grain yield. AUCIPC was found positively correlated with PH, CTD, BY and GY. AUDPC was found to be positively correlated with HI whereas negatively correlated with DF, PH, GPS, AUCIPC and BY. This indicated that higher disease severity was negatively associated with high chlorophyll index value.

(DR. SAIKAT DAS)

Chairman-Advisory committee

(SOURIK PODDAR)

Signature of student

*Chapter 1*

# ***Introduction***

# Chapter 1

## INTRODUCTION

---

Bread wheat is an allohexaploid ( $2n=42$ ) and it can be grown in many different environments. Although it is best adapted in cool or temperate growing conditions, it is being cultivated in many areas of the world where heat stress is a major yield limiting factor, especially during maturity stage. These areas include lowland central and peninsular India, lowland of Terai region of Nepal, Bangladesh, Thailand, southern China, Nigeria, Sudan, Bolivian lowlands and parts of Brazil and Paraguay (Dubin and Rajaram, 1996).

The above-mentioned heat stressed areas of tropical lowlands represent 9 million hectares of wheat production and can be split into lowland humid areas (e.g., Bangladesh, eastern India, Terai of Nepal and lowlands of Bolivia and Paraguay) and lowland dry areas (e.g., central and peninsular India, Nigeria and Sudan). These areas are not considered as typical wheat-growing regions, but rather warmer, non-traditional wheat zones. In many of these areas, wheat is a new crop that came with the “Green Revolution” in the 1960s and 1970s. Wheat diseases in the lowland tropical environments can be severe and require significant efforts to control. Especially in lowland humid areas where humidity is high disease problem is severe.

After China, India is the world's second-largest wheat producer, accounting for roughly a quarter of global wheat output. Wheat output in India has grown by more than six times over the last four decades, from about 12 million tonnes in 1964-65 to around 107.18 million tonnes in 2019-20. Wheat acreage has risen from 13 million hectares to about 30 million hectares over this time, while productivity has improved from  $9.13 \text{ q}^{-\text{ha}}$  to  $35.0 \text{ q}^{-\text{ha}}$ . (Source: Project Directors Report, 2019-20, IIWBR, India). This was only possible due to the import of semi-dwarf genotypes from (CIMMYT) during the 1960s which was a crucial component in India's green revolution, which saw a quantum jump in wheat yield.

Rice-wheat cropping is the major cropping system on 13.5 mha in the Indo-Gangetic plains (IGP) (Pathak *et al.*, 2003; Panigrahy *et al.*, 2011) producing about 50% of the total food grain and feeding 40% of India population (Gupta *et al.*, 2016). In the last few decades climate change is changing our globe in which agricultural crops suffers badly (Rosenzweig *et al.*, 2014). Now, this rice-wheat cropping system has started showing declining trends in marginal

yield, groundwater depletion, soil deterioration and heat stress problem in last decade. Rise in temperature is one of the most important concerns as metabolic processes that eventually impact the agricultural production is governed by temperature (Teixeira *et al.*, 2013). Further increase in temperature will negatively affect agricultural production and more serious effect will be from short episodes of extremely high temperatures, or occurrence of 'heat stress'. This effect is being observed in India especially in wheat crop since last decade (Dubey *et al.*, 2020). Early onset of significantly higher temperature coincided with wheat grain filling stage especially in Indo-Gangetic plains leading to terminal heat stress and reduction in yield. The possible reason behind this is the proximity to the equator and late sowing of wheat in India which exposes the crop to high temperature during grain filling stage (Joshi *et al.*, 2007).

Terminal heat stress in wheat occurs when mean temperature during grain filling stage goes above 31°C. The IPCC (2014) projected that for Indian region, temperature will increase by 0.7–2.0 °C by 2030s and 3.3–4.8 °C by 2080s. The increase may be more in north India and during the rabi season (November to March). With a rise of temperature by 0.5–1.56 °C by 2080–2100, adverse impact on food production will occur which will cause a loss of 10–40% in food grain production in India (Parry *et al.*, 2004; IPCC, 2007). High temperature causes adverse impact on wheat physiology restricting growth and yield. At flowering stage, heat stress causes sterility of pollen and anther leads to underdeveloped embryo which in turn reduces grain number, while heat stress during grain filling stage leads to reduced grain filling rate and in turn reduces grain weight and overall yield (Mondal *et al.*, 2013). In a study it was found that even a rise of 1°C in the mean temperature in month of March-April leads to reduction in the duration of wheat crop by seven days and yield by about 400 kg per hectare (Singh *et al.*, 2011). In a study done by International Maize and Wheat Improvement Centre (CIMMYT) the areas with lower productivity potential will have more negative impact and in future with global warming, their productivity will further decrease under heat stress condition (Joshi *et al.*, 2007). All of these findings indicate that there is a need to understand the effects of rising temperatures on wheat growth and development as well as adoption of climate smart practices (Niles *et al.*, 2015).

CIMMYT nurseries have played an important role in wheat breeding throughout the world. CIMMYT has divided the world's various wheat producing zones into many mega-settings and distributed sophisticated breeding lines to a variety of conditions across the world, with an emphasis on evaluating genotypes for greater adaptability and selection for specific environments (Rajaram *et al.*, 1995; Braun *et al.*, 2010). Moreover, excellent CIMMYT-

derived bread wheat lines have been widely adopted in cross-breeding programmes throughout the developing world (Braun *et al.*, 1996), substantially increasing the genetic variety of wheat cultivars in many countries (Samale *et al.*, 2002). Advanced breeding lines targeted for heat stressed areas are annually distributed to internal co-operators through High Temperature Wheat Yield Trial (HTWYT) nursery.

West Bengal is not a large wheat-growing state and it occupies around 0.2 million hectares with an annual production of 0.6 million tons. Rice –wheat is the major cropping system and the crop faces much biotic and abiotic stresses. Terminal heat stress is a major concern as the crop sown late due late harvest of paddy. Temperature rises after February onwards which adversely affects the crop. Due to presence of high humidity, disease occurrence is also high. Spot blotch or foliar blight disease produced by *Bipolaris sorokiniana* (Sacc.) Shoem is one of the most serious diseases found in this region. This is a serious disease that creates tiny dark brown lesions on the leaf that quickly congeal and spread in sensitive genotypes. The severity is most prevalent in the eastern Gangetic plains of South Asia, which encompass India, Nepal, and Bangladesh (Sharma and Duveiller, 2006). In India, average yield losses owing to spot blotch have been found to be 15.5% (Dubin and Van Ginkel, 1991) and 17% (Saari, 1998), with grain yield losses ranging from 17.63% to 20% under favourable conditions (Goel *et al.*, 2006). Under severe infestation, however, yield loss might reach 80% (Joshi *et al.*, 2007). The Terai area of West Bengal, which has a high humidity level and a shorter winter season, is regarded a hotspot for spot blotch (Kumar *et al.*, 2016).

Considering the above facts, the present investigation entitled "Study of Genetic Diversity of Wheat (*Triticum aestivum* L.) Genotypes Under Terai Agroclimatic Conditions" was undertaken with the following objectives:

1. To study the genetic diversity of wheat genotypes for yield and yield components along with physiological attributes.
2. To screen the genotypes for the resistance towards major Wheat disease like Spot Blotch.



***Chapter 2***

***Review of Literature***

## Chapter 2

# REVIEW OF LITERATURE

---

### 2.1. Genetic diversity

Individuals' genetic variations with regard to different morphological characters are referred to as genetic variation. It can be used to evaluate the contribution of each character to total divergence and thus aid in the selection of superior parents for hybridization programmes.

Mahalanobis *et al.*, (1936), define statistical principles as a quantitative method for estimating genetic divergence between populations. Evolutionary forces trigger changes in the frequency of different populations, resulting in genetic divergence.

For the evaluation of genetic diversity, Rao *et al.*, (1952) proposed using  $D^2$  statistics in plant breeding. Mol *et al.*, (1962) discovered that geographical distribution has little effect on genetic diversity.

Mahalanobis generalised distance calculated by statistic is used to estimate the degree of divergence between biological populations and to compute the contribution of different components to the total divergence. In the field of plant breeding, Nair and Mukharjee *et al.*, (1960) were the first to use the  $D^2$  statistic as an indicator of genetic divergence for classification.

Multivariate cluster analysis was used by Voidani *et al.*, (1993) to categorise geographical sites as well as eco geographical sub-populations based on plant characters. Due to eco-geographical variation, no clear pattern of variation among the regions was discovered.

In 121 indigenous and exotic wheat varieties, Redhu *et al.*, (1995) looked at genetic variation for nine quantitative characters. They discovered that the varieties were divided into 27 groups. The presence of significant variability in plant height, number of grains per ear, 1000 grain weight, and grain yield per plant was revealed by cluster means for different characters. They also discovered that the clustering of varieties was unrelated to their geographical origin.

Walia and Garg *et al.*, (1996) used cluster analysis to look at grain yield and its related traits in 405 pure breeding lines. They discovered 13 distinct clusters, and the clustering pattern of genotypes from the same country showed that they were distributed in several clusters,

indicating that regional and genetic variation was not parallel. Clusters IV and IX had a lot of diversity, and Cluster VI had a lot of high mean values for grain yield, biological yield, number of tillers/unit area, and harvest index.

Sharma *et al.*, (1998) investigated genetic divergence in 51 spring wheat genotypes, clustering them using three Tocher's values: 1060, 500, and 300. They discovered that genotypes were divided into seven, nine, and ten distinct clusters, respectively. At Tocher's values of 500 and 300, the linkage dendrogram and minimum spanning tree showed conformity with the clustering pattern of the  $D^2$  statistic.

Dotlacil *et al.*, (2000) stated that 120 accessions of European winter wheat land races and obsolete cultivars were clustered into eight clusters, and that clustering reduced cultivar heterogeneity within clusters in most of the evaluated characters. However, it was difficult to find a clear correlation between cultivar geographic origin and presence in specific clusters.

Bergale *et al.*, (2001) looked at genetic divergence among fifty bread wheat cultivars and found that they were clustered into 11 clusters. The appearance of genotypes from various geographical origins in a single cluster suggests that the cultivars may have an ancestral relationship. They also said that plant height was the most important factor in genetic divergence.

According to Nimbalkar *et al.*, (2002), 24 genotypes were divided into 12 clusters, with cluster III and IV having the largest and lowest intra cluster distances, respectively. Cluster VII and Cluster XII had the greatest inter-cluster size. The number of grains per spike, grain weight per spike, and number of active tillers all had a significant impact on the genetic diversity.

Using Mahalanobis  $D^2$  study, Dwivedi *et al.*, (2002) investigated the genetic divergence within 72 lines of bread wheat. The genotypes were divided into eight clusters based on  $D^2$  values, with cluster I having the most genotypes and cluster VIII having the fewest. Clusters I, III, and IV genotypes were classified as varied and had higher mean values for the most significant yield component traits.

Leilah *et al.*, (2005) conducted two field trials, over two successive winter seasons using 'Yokorarogo' cultivar to show the relationship between wheat grain yield and its components under drought condition. Principal Component Analysis (PCA) has grouped three main components accounting for 74.4% of total variation of grain yield (PC1- 33.9%, PC2- 26.8% and PC3-13.7% respectively). Cluster analysis with wheat variables was used and was

found that similarity level increase as the number of cluster increases. The study proved that 100 grain weight, weight of grain/spike, harvest index and biological yield were variables most closely related to grain yield and can be considered under drought condition.

The thirty genotypes were divided into six clusters, according to Kumar *et al.*, (2009). Cluster VI has been defined for selecting parents for integrating grain yield per plant, tillers per plant, and plant height, cluster V for spike length, grains per spike, and early maturity, and cluster III for 1000 grain weight, according to cluster means.

Jaiswal *et al.*, (2010) divided 300 indigenous bread wheat germplasm into twenty-three clusters, each with a different mean value for the characters under study. They also suggested that genotypes with desired values from different clusters could be used in breeding programmes to improve yield and bread wheat characters.

The 49 bread wheat genotypes were classified into 22 distinct groups by Hailegiorgis *et al.*, (2011) using cluster analysis. This suggests that the genotypes examined have a lot of variation. Cluster 9 and 13 genotypes should be considered for direct use as parents in hybridization operations to create high yielding wheat varieties based on cluster mean values. Clusters 3 and 16 genotypes can be utilised to increase protein and gluten content, early maturity, and other desired traits other than grain yield. The principal components analysis indicated that nine principal components (PC1 to PC9) were responsible for almost 80% of the overall variance. It was also noticed that the tiny contribution of a few characteristics, rather than the cumulative effect of a lot of characters, was responsible for the separation of genotypes into various clusters. The results of this research may be utilised to design crosses and maximise the usage of genetic diversity and heterosis expression.

Singh *et al.*, (2014) used Tocher's and Euclidian methods of divergence to group thirteen wheat genotypes into four clusters. They also noticed that the genotypes and numbers of genotypes in each method's clusters were different.

Zaman *et al.*, (2014) studied divergence analysis in 30 drought tolerant genotypes of wheat. The result indicated presence of significant variation among the genotypes and was further classified into six clusters. The PCA analysis revealed PC1, PC2, PC3, PC4 and PC5 where the most contributors to the variance with 30.78%, 20.11%, 17.75%, 10.93% and 7.63% as their respective values. The divergence contributing traits (days to heading, spike/m<sup>2</sup>, and 1000 grain weight) as the most important for drought tolerance improvement through proper selection of parents. Depending upon cluster distance and cluster mean the genotypes of Cluster

II and VI could be considered as the parent materials for future drought tolerant high yielding hybridization programme.

Verma *et al.*, (2014) assessed the genetic variation of 108 bread wheat accessions from India and Australia for production and yield traits. They discovered that these genotypes were divided into eleven clusters with a distribution pattern suggesting that cluster IV (26) had the most genotypes, followed by cluster VI (22) and cluster II (12). In most cases, the inter-cluster gap was greater than the intra-cluster distance, suggesting greater genetic variability among accessions from different classes. Cluster VII and IX had the greatest inter-cluster difference (113.94), followed by VIII and X (97.72), indicating a great deal of variability within the clusters. Cluster X (13.96) had the greatest intra-cluster distance, while Cluster VII had the smallest (00.00). Cluster X genotypes had the largest mean grain yield, harvest index, and spike weight values. Perenjori, KRL 261 and KRL 283 from cluster X, and Gutha from cluster IX, can be used as possible donors for a hybridization programme to grow genotypes with high grain yields.

Salman *et al.*, (2014) separated 65 wheat accessions into six classes. Cluster 1 and cluster 4 had the most variety. This high level of diversity illustrates why prospective breeding programmes would have greater parental selection.

The 64 genotypes were grouped into nine clusters, according to Fikre *et al.*, (2015). Cluster I and IX had the greatest inter cluster gap ( $D^2=5112.1$ ), followed by clusters II and IX ( $D^2=4694.4$ ) and VIII and IX ( $D^2=3871.9$ ), indicating that they were genetically more divergent from one another than any other cluster. Crosses of genotypes from cluster I with cluster IX, cluster III with cluster IX, and cluster VIII with cluster IX are predicted to develop progenies with higher levels of genetic recombination and segregation.

## **2.2. Physiological study**

### **2.2.1. Canopy temperature depression (CTD)**

Under heat and drought stress, canopy temperature (CT) is a significant parameter that indicates relative resistance to terminal heat stress, and canopy temperature depression (CTD) is found to be closely associated with yield and yield attributes (Amani *et al.*, 1996 and Reynolds *et al.*, 1998).

In the season, Rosyara *et al.*, (2007) investigated canopy temperature depression as a correlative indicator of spot blotch resistance and heat stress tolerance. In both heat stressed

(late sowing) and non-stressed (timely sowing) field conditions, ten genetically diverse genotypes were grown. A negative association ( $r=-0.72^*$ ) was found between AUDPC per day and AUCTDPC, suggesting that AUCTDPC declines as foliar blight susceptibility increases. Based on AUCTDPC and AUDPC per day readings, genotypes may be classified as tolerant of either one or both stresses based on genetic variations for spot blotch resistance and heat stress response.

Under heat-stress conditions, Gowda *et al.*, (2011) investigated the relationship between canopy temperature depression, membrane stability, relative water quality, and grain yield in bread wheat (*Triticum aestivum*). They looked at 49 different bread wheat genotypes at three different sowing dates (15 November, 5 December, and 5 January) to see whether there was a connection between physiological parameters and grain yield in hot environment of New Delhi. The canopy temperature depression at anthesis and the canopy temperature depression at 10 days after anthesis demonstrated positive and important genotypic correlation coefficients in grain yield. Traits such as canopy temperature depression (at anthesis), canopy temperature depression (10 days after anthesis), and membrane damage both had high heritability estimates and could be used as selection criterion in stressful environments.

In South Asia, Mondal *et al.*, (2013) investigated wheat earliness as a key to adaptation under terminal and continuous high temperature stress. They looked at 30 wheat genotypes in 13 different areas in South Asia, as well as two different conditions in Mexico. For DH, PH, GY, and TKW, there were significant discrepancies between Mega Environments (ME). MEI sites had higher mean GY of 5.26 t<sup>ha</sup> and TKW of 41.8 g, compared to 3.63 t<sup>ha</sup> and 37.4 g for MES. Early heading entries (79 days mean DH) outperformed local checks in every region, with GY of 2-11 percent higher and 40-44 g TKW. CT was linked to GY in the Mexico study, implying that cooler canopies can lead to higher GY under both normal and high temperature stress conditions.

Ray and Ahmed (2015) investigated the impact of canopy temperature on yield and grain growth of various wheat genotypes sown at various times. On November 29, sowing, canopy temperature depression was almost identical in all genotypes (mean canopy temperature depression at various stages was 5.99°C for BARI gom 26 and 5.5°C for Pavon 76). The results showed that in the 30 December sowing, BARI gom 26 had a higher mean value of canopy temperature depression (232°C), while Pavon 76 had a lower value (0.88°C) at various points, suggesting that BARI gom 26 had cooler canopies even under post anthesis

heat stress. BARI gom 25 and BARI gom 26 continued to raise grain dry matter up to 32 days after sowing on December 30, while Pavon 76 stopped 8 days earlier, indicating that BARI gom 26 has a higher relative 1000-grain weight (96%) and grain yield (89%) than Pavon 76. Grain growth and yield were found to be greater in warmer environments while BARI gom 26 sustained a higher canopy temperature depression.

Under terminal heat stress, Jangid *et al.*, (2018) investigated 20 spring wheat genotypes for canopy temperature depression and remain green traits. The results of terminal heat were studied using delayed sowing at three separate dates: standard (S1; November 26, 2011), late (S2; December 25, 2011), and very late (S3; January 10, 2012). In comparison to S1, the chlorophyll content in flag leaf decreased during the anthesis stage in S2 and S3 plants. The temperature of the canopy rose as plant growth progressed and sowing was delayed. Based on susceptibility index and relative yield loss, the results showed that NW 1014 was the most resistant genotype and K 911 was the most vulnerable to terminal heat stress.

### **2.2.2. Chlorophyll index (CI)**

Rice-wheat cropping is the most common cropping method in India. Due to the late harvesting of rice in this cropping method, wheat sowing is delayed. Wheat crops suffer from terminal heat stress in such conditions, and crop yields are reduced due to disruptions in plant physiological processes (Jangid and Srivastava *et al.*, 2018). Stay green characters, lower photosynthetic rate reduction, lower canopy temperature, and higher leaf conductance were found to be consistent with terminal heat resistance in wheat (Fischer *et al.*, 1998).

Handheld chlorophyll metres such as the SPAD 502, Field scout CM 1000, and others will calculate leaf greenness, which is positively associated with leaf chlorophyll material.

The abundance of various leaf pigments such as chlorophyll and carotenoid can be predicted using spectral reflectance. Changes in photosynthesis are more closely related to changes in chlorophyll content; both of these changes occur during the grain filling period, which has an effect on grain weight (Guendouza and Maamari *et al.*, 2012). The cumulative chlorophyll content of the leaves is calculated using the chlorophyll index.

Murdock *et al.*, (2004) compared both reflectance and transmittance/absorbance chlorophyll metres on wheat and found the reflectance-type metre (Field Scout CM 10003) to be very effective in assessing leaf chlorophyll content. Ambient and reflected light is used to measure the chlorophyll index (CI). Leaf greenness improved with growing nitrogen treatments

applied at rates of 0, 30, 60, 90, 120, and 150 pounds of nitrogen per acre in a three-year trial on soft red winter wheat. When the R metre was used between 10 a.m. and 2 p.m., their similarity with both the T/A metre and the R metre was exceptional and nearly equal. This meant that in the case of wheat, CMI 1000 could be used to obtain an accurate estimation of leaf chlorophyll content.

Talebi *et al.*, (2011) investigated the use of chlorophyll content and canopy temperature as drought tolerance measures in durum wheat (*Triticum durum* Desf.). Two criteria were used to test twenty-four durum wheat genotypes (well-watered and moisture-stressed). The genotypes with high yield often had a high chlorophyll content and a low canopy temperature in well-watered conditions. Furthermore, genotypes with a low canopy temperature can sustain high transpiration, photosynthetic rate, and yield under moisture-stressed conditions. In both environments, there was a strong positive association between chlorophyll quality and yield. The important association between canopy temperature and chlorophyll content with yield under moisture-stressed conditions could explain the potential for screening wheat genotypes for drought conditions.

Aryal and his associates (2015). Drought resistant wheat I genotypes sown at normal and late conditions were measured for chlorophyll content as an indication of spot blotch resistance. A split plot pattern of three replications was used to test 20 genotypes. Aditya, CSISA DRYT 5204, and CSISA DYRT 5205 had longer periods of staying green, higher SPAD, and lower AUDPC values, indicating that these three genotypes are suitable for late sown conditions.

### **2.3. Study on Spot blotch resistance**

Many diseases afflict the warmer parts of the world, and among them is spot blotch or foliar blight caused by *Bipolaris sorokiniana* (Sacc. In Sorok). Due to its widespread prevalence and growing severity, Shoem is one of the most concerning diseases in India and other South Asian countries (Joshi *et al.*, 2002).

Van Ginkel *et al.*, (1998) and Rajaram *et al.*, (1998) described several spot blotch resistance sources and divided them into three groups: Latin America, China, and wheat wild relatives or alien species. Sanghai#4, Suzhoe#8, and Yangmai#6 were among the first Chinese sources of resistance used at CIMMYT. Latin American origins are mostly from Brazil, though



some may have Italian ancestors. According to Mehta *et al.*, (1985), older resistant Brazilian commercial varieties are BH-1 146 and CNT-11.

Chaurasia *et al.*, (1999) tested CIMMYT and Indian gene pool spring wheat lines and found 43 lines to be resistant; CIMMYT lines were more resistant than Indian lines. Different approaches to studying seedling/adult plant resistance to spot blotch have been used, and they can be divided into two groups. The studies in the first category used a Mendelian approach involving crosses between resistant and susceptible genotypes followed by an analysis of segregation pattern, there are some reports of the presence of monogenic (Amey *et al.*, 1951; Wilcoxson *et al.*, 1990) and polygenic (Griffie *et al.*, 1925; Mehta *et al.*, 1985; Steffenson *et al.*, 1996) types of resistance to spot blotch disease., and the second group of research used a quantitative genetics approach with molecular markers. Another study used F1 and F2 generations derived from six crosses involving three cultivars to infer additive effects between two or three recessive genes (two each in cultivars PBW 343 and HS361, and three in RAJ 3702) Bhushan *et al.*, (2002). In two other experiments, epistatic interactions among three dominant genes were inferred (Neupane *et al.*, 2007). In the four moderately resistant cultivars Gisuz, Cugap, Chirya 1, and Sabuf, polygenic control with two or three genes providing stable and durable resistance was also registered (Velazquez *et al.*, 1994).

Joshi *et al.*, (2004) looked at the segregating generations (F3, F4, F5, and F6) of three crosses with resistant (ace. no. 8226, Mon/Ald, Suzhoe#8) and susceptible (Sonalika) parents, finding that resistance was regulated by three additive genes. When both of these experiments are considered together, a polygenic or quantitative existence of resistance can be hypothesised, which is confirmed by studies involving QTL interval mapping and GWAS.

Spot blotch is a serious concern for wheat cultivation in warmer and humid regions of the world, according to Acharya *et al.*, (2011). Disease intensity was linked to humidity, temperature, and soil nutrient levels. When the flag leaf and the leaf underneath the flag leaf become infected before the head emerges, the yield loss is the greatest.

In 2004 and 2005, Sharma *et al.*, (2006) and Duveiller *et al.*, (2006) found that spot blotch reduced grain yields by 4% to 38% and 25% to 43%, respectively. In 2004, the weight of a thousand kernel and the number of kernels per spike were decreased by 15% and 10%, respectively, and by 18% and 11% in 2005. According to the findings, the new cultivar Gautam's level of resistance to spot blotch reflects a partial success in breeding for resistance.

Rai *et al.*, (2016) investigated the physiological trait for tolerance to spot blotch (*Bipolaris sorokiniana*) in wheat genotypes (*Triticum aestivum*). Three wheat varieties were tested for physiological traits as well as spot blotch resistance at various sowing dates. In the 5th of November, AUCTPC (207.15) and AUDPC (287.38) reported significantly lower values, while HD-2967 recorded significantly higher values (278.53) The AUDPC was lower, but the yield was highest on November 5th (0.59g/m<sup>2</sup>). There was a positive correlation between canopy temperature and disease, but a negative correlation with the ability to remain green. Yield and canopy temperature were found to be negatively associated with disease, with the severity of the disease accounting for up to 59% of the difference in yield. The plant's green index is the only variable that has a positive relationship with yield. The linear relationship shows that as the stay green property increases, the canopy temperature decreases, and it accounts for about 72% of the canopy temperature rise. The results showed that delaying planting raises canopy temperature and lowers crop greenness index, resulting in an increase in disease and a decrease in yield.

Rosyara *et al.*, (2010) investigated the effects of spot blotch and heat stress on hexaploid wheat genotypes' canopy temperature depression, chlorophyll fluorescence, and chlorophyll quality. Spot blotch and heat stress are two major stresses on bread wheat caused by *Cochliobolus sativus* (Ito and Kurib.) Drechsler ex Dastur (*Triticum aestivum* L.). In 2006 and 2007, eleven different bread wheat genotypes were tested in replicated field trials in Rampur, Nepal, under spot blotch epidemics and heat stress conditions. CTD (canopy temperature depression), CF (chlorophyll fluorescence), Chlorophyll content, percent disease leaf area, yield, and yield components were all measured. Individually, heat and spot blotch decreased CTD, CF, chlorophyll content, grain yield (GRY), and thousand kernel weights (TKW), with combined stress resulting in even greater reductions. Genotypes with lower GRY or TKW losses due to spot blotch also had lower yield losses due to heat stress or combined heat and disease stress, implying a connection between stress tolerance mechanisms. The highest values for chlorophyll content, CF, and CTD were found in genotypes with less disease.

*Chapter 3*

***Materials and  
Methods***

Under the different headings in the chapter, the specifics of the experimental material utilised and procedures employed during the current investigation are explained.

### 3.1. Experimental site

The research was carried out at the university instructional farm, Uttar Banga Krishi Vishwavidyalaya, Pundibari, Coochbehar, West Bengal, during the rabi season of 2020-2021. The farm is located at 26°19'86" North latitude, 89°23'53" East longitude, and is 43 metres above sea level.

### 3.2. Experimental material

The experimental material consists of 50 different wheat (*Triticum aestivum* L.) genotypes from the nursery sent by CIMMYT (19<sup>th</sup> High Temperature Wheat Yield Trial [19 HTWYT]). Whole list along with their pedigree is given in Table No. 3.1.

### 3.3. Meteorological features of the experimental site

**Climatic condition:** The experimental location is located in the Sub-Himalayan terai agro-climatic zone, with average rainfall of 3000 mm from June to September. The temperature started to increase towards the end of February and peaked in April and May. Throughout the year, the relative humidity remained high, ranging from 39.86 to 97.86 %. Early rains in March-April 2021 hindered the mature crop, but late monsoon rains in October 2020 provided an advantage.

**Temperature:** At the beginning, temperature was moderate but gradually increased during maturity.

**Rainfall:** During the growing season the crop had a couple of unseasonably wet days during February.

**Humidity:** During the experiment, humidity was high to moderate, ranging from a maximum of 97.86% to a minimum of 36.86% in 2020-2021.

**Table 3.1. List of wheat genotypes evaluated during 2020-2021**

S. N	GENOTYPE	PEDIGREE
1	ENTRY 1	DBW 187 (LOCAL CHECK)
2	ENTRY 2	NADI#1
3	ENTRY 3	WBLL1*2/BRAMBLING/4/BABAX/LR42//BABAX*2/3/SHAMA*2/5/...
4	ENTRY 4	QUAIU#1/SUP152
5	ENTRY 5	PBW343*2/KUKUNA/3/PASTOR//CHIL/PRL/4/GRACK/5/MUU/...
6	ENTRY 6	CHIBIA//PRLII/CM65531/3/FISCAL*2/4/TAM200/TURACO/5/...
7	ENTRY 7	WBLL1*2/BRAMBLING*2//BAVIS/3/CHYAK1/VILLA JUAREZ F2009/...
8	ENTRY 8	KACHU//WBLL1*2/BRAMBLING*2/6/BECARD#1/5/KIRITATI/4/...
9	ENTRY 9	SUP152/BAL#1*2/3/KINGBIRD#1//INQALAB 91*2/TUKURU
10	ENTRY 10	SUP152/BAL#1*2/3/KINGBIRD#1//INQALAB 91*2/TUKURU
11	ENTRY 11	SUP152/BAL#1*2/3/KINGBIRD#1//INQALAB 91*2/TUKURU
12	ENTRY 12	SUP152/BAL#1*2/3/KINGBIRD#1//INQALAB 91*2/TUKURU
13	ENTRY 13	SUP152/BAL#1*2/3/KINGBIRD#1//INQALAB 91*2/TUKURU
14	ENTRY 14	TUKUU//BAV92/RAYON/3/FRNCLN/4/2*FRNCLN*2/TECUE#1
15	ENTRY 15	ABLEU*2/BORL14
16	ENTRY 16	MILAN/KAUZ//BABAX/3/BAV92/4/WHEAR//2*PRL/2*PASTOR/5/...
17	ENTRY 17	KENYA SUNBIRD/2*KACHU//KFA/2*KACHU
18	ENTRY 18	BAVIS/NAVJ07//SUP152/BAJ#1
19	ENTRY 19	CHIPAK*2//SUP152/KENYA SUNBIRD
20	ENTRY 20	WBLL1*2/CHAPIO/6/CNDO/R143//ENTE/MEX175/3/AE.SQ/4/...
21	ENTRY 21	HEILO//MILAN/MUNIA/3/KIRITAI/2*TRCH/4/2*KACHU/KIRITATI
22	ENTRY 22	CHIBIA//PRLII/CM65531/3/FISCAL*2/2/TAM200/TURACO/5/...
23	ENTRY 23	WBLL1*2/BRAMBLING*2//BAVIS*2/4/SWSR22T.B./...
24	ENTRY 24	BECARD/FRNCLN//BORL14
25	ENTRY 25	TACUPETO F2001*2/KIRITATI//BLOUK#1/3/WBLL1*2/...
26	ENTRY 26	QUAIU#1/BECARD/3/WBLL1*2/BRAMBLING*2//BAVIS
27	ENTRY 27	BORL14*2/8/REH/HARE//2*BCN/3/CROC 1/AE.SQUARROSA (213)/...
28	ENTRY 28	BORL14*2/8/REH/HARE//2*BCN/3/CROC 1/AE.SQUARROSA (213)/...
29	ENTRY 29	SWSR22T.B./2*BLOUK#1//WBLL*2/KURUKU/3/BORL14/4/...
30	ENTRY 30	ELVIRA/5/CNDO/R143//ENTE/MEX175/3/AE.SQ/4/2*OCI/6/VEE/...
31	ENTRY 31	BOKOTA/3/ND643/2*WBLL1//2*BAJ#1
32	ENTRY 32	SUP152/BAJ#1/3/SWSR22T.B./2*BLOUK#1//WBLL1*2/KURUKU
33	ENTRY 33	MUTUS//ND643/2*WBLL1/3/BORL14
34	ENTRY 34	SHA7//PRL/VEE#6/3/FASAN/4/HAAS8446/2*FASAN/5/CBRD/KAUZ/...
35	ENTRY 35	KACHU/SAUAL//PRL/3/KACHU/KIRITATI
36	ENTRY 36	ROLF07//LALBMONO1*4/PVN/3/BORL14
37	ENTRY 37	NADI*2/3/EBW10 TALL#1/WESTONIA-Rht5//NAVJ07
38	ENTRY 38	SUP152/BAJ#1*2/4/WHEAR/VIVITSI//WHEAR/3/PANDORA
39	ENTRY 39	BECARD/FRNCLN//2*BORL14
40	ENTRY 40	BECARD/FRNCLN//KACHU/KIRITATI/3/BOKOTA
41	ENTRY 41	ATTILA/3*BCN//BAV92/3/PASTOR/4/TACUPETO F2001*2/...
42	ENTRY 42	MUNAL*2/WESTONIA/3/WBLL1*2/BRAMBLING*2//BAVIS/4/...
43	ENTRY 43	ROLF07*2/SHORTENED SR26 TRANSLOCATION//MUNAL#1/3/...
44	ENTRY 44	MUTUS*2//TAM200/TURACO*2/3/KFA/2*KACHU
45	ENTRY 45	MUTUS*2//TAM200/TURACO*2/3/KFA/2*KACHU
46	ENTRY 46	MUTUS*2/HARIL#1*2/3/SWSR22T.B./2*BLOUK#1//WBLL1*2/...
47	ENTRY 47	PASTOR//HXL7573/2*BAU/3/WBLL1/4/SOKOLL/3/PASTOR//...
48	ENTRY 48	ISENGRAIN/KBIRD//MUNAL#1*2/3/KFA/2*KACHU
49	ENTRY 49	CROC 1/AE.SQUARROSA (205) //BORL95/3/PRL/SARA//TSI/...
50	ENTRY 50	CROC 1/AE.SQUARROSA (205) //BORL95/3/PRL/SARA//TSI/...

**Experimental soil:** The experimental field's soil is from the Teesta alluvial plain group, and it's a sandy loam with low water holding capacity and moderate fertility.

**Table 3.2. Meteorological data pertaining to the period of experimentation**

Weeks	Period	Temperature(°C)		Rainfall (mm)	Number of		Relative humidity	
		Min.	Max.		Rainy days	Sunshine hours	Min.	Max.
45	Nov. 04-Nov.10	14.70	30.17	0.00	0.00	6.69	48.00	67.29
46	Nov. 11 –Nov.17	13.63	31.46	0.00	0.00	7.56	42.86	60.14
47	Nov. 18 –Nov.24	13.03	27.26	0.00	0.00	5.53	52.29	82.71
48	Nov. 25 – Dec.01	11.26	29.40	0.00	0.00	5.33	50.14	68.57
49	Dec.02 – Dec.08	11.14	28.26	0.00	0.00	5.46	49.86	67.86
50	Dec. 09 – Dec.15	13.04	24.53	0.00	0.00	2.20	67.29	87.00
51	Dec.16 - Dec. 22	8.87	24.09	0.00	0.00	3.86	56.57	85.29
52	Dec. 23 – Dec.29	8.23	25.39	0.00	0.00	6.47	50.14	82.43
1	Dec. 30 – Jan.05	7.79	26.56	0.00	0.00	6.56	44.00	69.57
2	Jan.06 – Jan. 12	10.44	23.40	0.00	0.00	2.43	69.43	86.43
3	Jan.13 – Jan. 19	8.56	22.81	0.00	0.00	0.86	72.71	90.14
4	Jan. 20 – Jan. 26	7.69	22.00	0.00	0.00	2.39	69.57	89.00
5	Jan.27 – Feb.02	6.03	21.90	0.00	0.00	3.46	60.86	97.86
6	Feb. 03 – Feb.09	6.91	27.44	0.00	0.00	6.61	39.86	74.86
7	Feb.10 – Feb.16	9.19	27.59	0.00	0.00	5.46	41.86	84.29
8	Feb.17 – Feb.23	10.30	29.30	0.00	0.00	5.89	39.57	70.71
9	Feb.24 –Mar.02	13.59	28.29	0.00	0.00	3.09	56.71	82.29
10	Mar. 03 -Mar.09	11.99	28.97	0.00	0.00	4.23	47.29	67.14
11	Mar 10 – Mar16	16.17	30.24	14.30	4.00	3.01	52.14	72.14
12	Mar.17-Mar.23	15.57	32.70	0.00	0.00	5.57	41.43	67.71
13	Mar. 24-Mar.30	14.40	32.37	35.40	1.00	6.36	40.43	62.71
14	Mar.31 –Apr.06	14.77	31.49	2.80	1.00	5.07	50.57	61.00
15	Apr.07 – Apr.13	16.50	32.59	43.60	2.00	4.61	55.29	70.14
16	Apr. 14 –Apr.20	17.47	30.77	73.40	4.00	4.24	61.00	69.29
17	Apr. 21 –Apr.27	19.06	32.47	10.60	2.00	7.21	56.00	68.29

[Source: Gramin Krishi Mausam Sewa (GKMS) project, UBKV, Pundibari, Cooch Behar.]

### 3.4. Experimental details

The specifics of the experiment carried out in this study are listed in Table No. 3.3.

**Table 3.3. Experimental details**

Experimental Site	UBKV Instructional Farm Field
Year of experiment	Rabi 2020-21
Co-ordinates	26° 19' 86" N latitude, 89° 23' 53" E longitude
Sowing Date	28-11-2020 (Timely sown)
Plot size	2.5x1.5 (6rows per plot) row to row spacing 20 cm
No. of entries	50
Design of the experiment	Randomized Block Design (RBD)
Replications	2
Dates of harvesting	13-4-2021 (Timely harvested)

### 3.5. Agronomic practices.

Before planting, the soil was brought to a fine tilth. The fertiliser was administered at a rate of 150:60:50 kg/ha of N:P:K, with half of the nitrogen applied as a basal dosage and the remainder at the time of the first irrigation. At the CRI, Boot, and Milk stages, three irrigations were administered. Irrigation in the form of a flood was used.

### 3.6. Equipment used.

#### A) Meter scale.

The data on plant height and spike length was collected using a standard metre scale.

#### B) Chlorophyll meter.

The quantity of chlorophyll in the leaves was estimated using a Field scout CM 1000 chlorophyll metre (Spectrum technologies, Inc.). To determine the amount of chlorophyll in leaves, this metre detects light at wavelengths of 700 nm and 840 nm. At each wavelength, the ambient and reflected light is measured. As a result of the absorption of 700 nm light by chlorophyll a, the reflection of that wavelength from the leaf is decreased when compared to reflected 840 nm light. The wavelength of light with an 840 nm wavelength is unaffected by leaf chlorophyll concentration and may be used to estimate how much light is reflected owing to physical features of the leaf, such as the existence of a waxy or hairy surface. As soon as the trigger is pressed, the integrated lasers outline the target. The field of view is 0.434 inches (1.10 cm) in diameter at a distance of 11.2 inches (28.4 cm). The field vision expands to 7.4 inches (18.8 cm) in diameter at a distance of 72 inches (183 cm). From the observed ambient and reflected light data, a chlorophyll index value (0-999) is computed.

### **C) Infrared thermometer.**

The physiological data and canopy temperature were recorded using an AR20 (Intell smart) infrared thermometer. Using the concept of reflection, an infrared thermometer detects the temperature of the canopy. It includes laser sighting capabilities and monitors temperatures from -32°C to 380°C. By detecting the quantity of infrared radiation radiated by the object, this gadget can estimate temperature from a distance of 30cm (plant canopy).

### **3.7. Observation recorded.**

#### **Morpho-phenotypic trait study.**

This experiment looked at 12 morpho-phenotypic traits, including germination per metre, days to heading, plant height, awn length, spike length, grain per spike, spikelet per spike, tiller per metre, 1000 grain weight, grain yield, biological yield, and harvest index, to see if the genotypes were stable. In each of the two replications, data was gathered from five random plants from each plot, and an average value was calculated for statistical analysis. The methods for recording observations are explained further down.

- 1. Germination per meter:** The number of seedlings germinated in any three rows within a 1 metre length are counted individually and averaged .
- 2. Days to heading:** The number of days it took 50% of the plot's plants to flower was counted and recorded.
- 3. Plant height (cm):** The height of the fully grown plant was measured from the base to the tip of the panicle.
- 4. Awn length (cm):** The awns' length was measured from the base of the awns' attachment to the spikelet to the tip.
- 5. Spike length (cm):** The spike's length was measured from its neck node to its tip.
- 6. Grain per spike:** Each spike's total number of grains was recorded, and an average was computed.
- 7. Spikelet per spike:** The total number of spikelets from each spike was counted, and the average was computed.
- 8. Tiller per meter:** The number of matured tillers produced by a metre length of line sown plants was counted and represented as tillers per metre.



- 9. 1000 grain weight (g):** Thousands of grains were counted from harvested bulk grains, and their weight was recorded in gram using any electric balance, which was accurate to two decimal points.
- 10. Grain yield (t/ha):** After harvesting, the total weight of the grains was calculated, and the average of the two replications was stated in tonnes per hectare.
- 11. Biological yield (t/ha):** The plot's mature plants were uprooted and weighted, and the results represented in tonnes per hectare.
- 12. Harvest index:** It was calculated as per following formula:

$$\text{HI} = \text{Grain yield} / \text{Biological yield}.$$

### **3.8. Methods of recording physiological observations.**

- a) **Canopy temperature depression:** The canopy temperature was measured twice, at 68 DAS and 93 DAS, using an infrared thermometer. The canopy temperature as well as the temperature of the air were measured in five randomly selected plants of each genotype. Before recording the canopy temperature, the same infrared thermometer was used to obtain the air temperature by concentrating it on a blank sheet (white paper) positioned slightly above each plot. Data was collected in the same order in all two replications, and the average values were adjusted by two decimals. Using the formula, the raw data was transformed into a usable format.

$$\text{CTD} = \text{Air temperature} - \text{Canopy temperature}.$$

- b) **Chlorophyll index:** At four crop growth phases, 88 DAS, 95 DAS, 102 DAS, and 109 DAS, physiological parameters such as Chlorophyll index were measured. The chlorophyll index data were collected using a Field Scout CM 1000 chlorophyll metre. The metre was pointed at target row portions using the laser guide lights, and the value acquired was instantly displayed. Readings were obtained between the hours of 10 a.m. and 2 p.m., with the sun behind the reader and the ambient light receiver unobscured. The CM 1000 metre measurements were obtained 3 to 5 feet from the canopy at 45° or 90° angles to the wheat canopy surface. Only if the ambient light intensity is more than one on a scale of 0-nine is the chlorophyll index value evaluated. The measurements

are taken in a circular region of around 13-35 square inches (at 3-5 feet from the canopy), which includes a lot of plants and leaves.

Area under chlorophyll index progress curve (AUCIPC) was calculated as per following formula adapted from Rosyara *et al.*, 2007:

$$\text{AUCIPC} = \Sigma 1/2 (S_{i+1} + S_i) d$$

Where,

$S_i$  = Chlorophyll index value at the end of time 'i'

$S_{i+1}$  = Chlorophyll index value at the end of time 'i+1'

$d$  = Day's interval between two observations.

### 3.9. Methods of recording disease observation (Spot Blotch).

The disease spot blotch (*Bipolaris sorokiniana*) (Sacc) Shoem was seen at four crop growth stages: 88 DAS, 95 DAS, 102 DAS, and 109 DAS in this study. A double-digit scale (00-99) was created as a variation of Saari and Prescott's severity scale to score diseases (Saari and Prescott, 1975). The first digit (D1) denotes disease progression from ground level to canopy height; the second digit (D2) denotes disease severity as assessed by diseased leaf area. D1 and D2 are both graded on a scale of 1 to 9.

For each score, the percentage of disease severity is estimated based on the following formula:

$$\text{Severity}(\%) = (D1/9) \times (D2/9) \times 100$$

### The Area Under Disease Progress Curve (AUDPC).

To analyse the severity of the disease, Area Under Disease Progress Curve (AUDPC) was calculated by using the following formula given by Wilcoxson *et al.*, (1975). The AUDPC has no unit.

$$\text{AUDPC} = \Sigma 1/2 (X_{i+1} + X_i) d$$

Where,

$X_{i+1}$  = Disease severity on 'i+1'<sup>th</sup> day

$X_i$  = Disease severity on 'i'<sup>th</sup> day

$d$  = Day's interval between two observations.

Genotypes were classified into resistant (R), moderately resistant (MR), moderately resistant to moderately susceptible (MR-MS), moderately susceptible (MS), moderately susceptible to susceptible (MS-S), susceptible (S), susceptible to highly susceptible (S-HS) and highly susceptible (HS) as per AUDPC values suggested by Liatukas and Ruzgas, 2012. The AUDPC scale is as follows:

AUDPC value	Type of resistance
< 100.0	Resistant (R)
100.1-150.00	Moderately Resistant (MR)
150.1-200.00	MR-MS
200.1-250.00	Moderately susceptible (MS)
250.1-300.00	MS-S
300.1-350.00	S
350.1-400.00	S-HS
> 400.00	Highly susceptible (HS)

### 3.10. Statistical analysis.

Statistical analysis was done by software such as R, Genstat and OPSTAT.

#### 3.10.1. Analysis of variance

For statistical analysis, the mean genotype values in each replication were utilised. To assess the importance of variance among the genotypes (treatments) for various characteristics, the data was analysed using a randomised full block design. Panse and Sukhatme (1989) detailed the procedures required in analysing the randomised full block design.

The following mathematical model was used in the analysis

$$Y_{ij} = \mu + t_i + b_j + e_{ij}$$

Where,  $i = 1, 2, 3, 4, \dots, t$ , number of treatments (t)

$j = 1, 2, 3, 4, \dots, r$ , number of replications (r)

$Y_{ij}$  = Performance of  $i^{\text{th}}$  genotype in  $j^{\text{th}}$  replication

$\mu$  = general mean of the population

$t_i$  = effect of  $i^{\text{th}}$  treatment

$b_j$  = effect of  $j^{\text{th}}$  replication

$e_{ij}$  = random error associated with  $i^{\text{th}}$  treatment and  $j^{\text{th}}$  block

The partitioning of total variance, due to block, treatments and error and their expectation are given in the following table.

**Analysis of Variance (ANOVA) for RCBD**

Source of variance	df	Sum of squares	Mean sum of square	F value
Replication	r-1	SSr	MSr	MSr/MSe
Genotypes	t-1	SSt	MSt	MSt/MSe
Error	(r-1)(t-1)	SSe	MSe	
<b>Total</b>	(rt-1)			

Where,

- r** = number of replications
- t** = number of genotypes
- df** = degrees of freedom
- SSr** =sum of square of replication
- SSt** = sum of square of treatment
- SSe** = sum of square of error
- MSr** = Mean square for replication
- MSt** = Mean square for treatment
- MSe** = Mean square for error

**Genotypic variance ( $\sigma^2g$ )=**  $(MSt - MSe)/r$

**Phenotypic variance ( $\sigma^2p$ )=**  $\sigma^2g + \sigma^2e$

**Error variance ( $\sigma^2e$ ) =**  $MSe$

For each character, the significance of variance among the treatment means was evaluated using the ‘F’ test at a 5% or 1% level of significance. It was indicated as significant if the computed ‘F’ value was higher than the table value, or vice versa. The critical difference (CD) was calculated to evaluate the significance of the difference between treatment means whenever the ‘F’ value was determined to be significant.

**CD = SEd × t (5%) at error d.f.**

Where, **t** = table value of ‘t’ at error d.f.

**SEd** = standard error of difference between two treatment means

$$\mathbf{SEd} = \sqrt{2\mathbf{MSe}/r}$$

Where,  $\mathbf{MSe}$  = Mean sum square of error  
 $\mathbf{r}$  = Number of replications.

If the difference between the two-treatment means is higher than the CD value (at 5% or 1%), it is said to be significantly different or visa-versa

### 3.10.2. Heritability and Genetic advance

#### Heritability

Heritability in broad sense  $h^2$  (b) was computed as a ratio of genotypic variance to phenotypic variance (Allard, 1960)

$$h^2(\mathbf{b}) = \frac{\sigma^2\mathbf{g}}{\sigma^2\mathbf{p}} \times 100$$

Where,  $h^2(\mathbf{b})$  = Heritability in broad sense  
 $\sigma^2\mathbf{g}$  = Genotypic variance  
 $\sigma^2\mathbf{p}$  = Phenotypic variance

#### Genetic advance

The expected genetic advance under selection for the different characters was estimated as suggested by Allard (1960).

$$\mathbf{G.A.} = h^2(\mathbf{b}) \times \sigma_p \times \mathbf{k}$$

Where,  
 $\mathbf{G.A.}$  = Expected genetic advance  
 $h^2\mathbf{b}$  = Heritability in broad sense  
 $\sigma_p$  = Phenotypic standard deviation ( $\sqrt{\sigma_p^2}$ )

$\mathbf{K}$  = Intensity of selection, the value of which is 2.06 when 5 percent of the individual are selected from the population as given by Lush (1949).

Genetic advance as percent of mean for each character was calculated as suggested by Johnson *et al.* (1955).

$$\text{Genetic advance as percent of mean} = \frac{\text{GA}}{\text{Mean}} \times 100$$

### 3.10.3. Coefficient of variance

Following Burton and Devane (1953), the genotypic coefficient of variation (GCV), phenotypic coefficient of variation (PCV), and environmental coefficient of variation (ECV) were calculated.

$$\text{GCV} = \frac{\text{Genotypic standard deviation } \sqrt{\sigma_g^2}}{\text{Mean}} \times 100$$

$$\text{PCV} = \frac{\text{Phenotypic standard deviation } \sqrt{\sigma_p^2}}{\text{Mean}} \times 100$$

The estimates of genotypic and phenotypic standard deviations were obtained from the respective analysis of variance table for different characters.

For convenience following classifications were used for describing various parameters of variability in the text:

PCV and GCV	>25% High 10-25% Moderate <10% Low
Heritability ( $h^2$ )	>60% high 30-60% Medium <30% Low
Genetic advance	>20% High 10-20% Moderate <10% Low

The range of heritability and genetic advance as percentage of mean was classified as suggested by Johnson *et al.* (1955).

### 3.10.4. Estimation of Correlation coefficient

Correlation was calculated as the relationship between different character pairings (between two variables). As proposed by Searle (1961), the genotypic association was evaluated using analysis of variance and covariance.

#### Genotypic correlation between character x and y

$$r_{xy}(g) = \frac{\text{Cov}_{xy}(g)}{\sqrt{\text{Var}_x(g) \times \text{Var}_y(g)}}$$

Where,  $\text{Cov}_{xy}(g)$  = Genotypic covariance between two-character x and y

$\text{Var}_x(g)$  = Genotypic variance for characters x

$\text{Var}_y(g)$  = Genotypic variance for characters y

The significance of correlation coefficient (r) was tested by comparing the observed value of correlation coefficient with the tabulated value for (n-2) degrees of freedom. If the observed value is more than the table value, the correlation coefficient is said to be significant.

$$tc = \frac{r\sqrt{n-2}}{\sqrt{1-r^2}}$$

Where,  $r$  = correlation coefficient

$n$  = number of genotypes

$tc$  = t calculated

### 3.10.5. Genetic Diversity.

The generalised distance between two population is defined by Mahalanobis (1936) as

$$D^2 = \sum \sum \lambda_{i,j} \cdot d_i \cdot d_j$$

Where,  $\lambda_{i,j}$  = Reciprocal matrix to the common dispersion matrix

$d_i$  = difference between the mean values of two population for  $i^{\text{th}}$  character.

$d_j$  = difference between the mean values of two populations for  $j^{\text{th}}$  character.

In the current investigation, estimating  $D^2$  values from the above formula is extremely difficult since it requires inverting a thirteenth order determinant and then evaluating  $B(BH) / 2$

terms whose total equals  $D^2$ . Working using a collection of uncorrelated characters derived from the original measurements was found to be more convenient.  $D^2$  is simplified to the evaluation of a simple sum of uncorrelated character derived from original measurements when such converted variables are used.  $D^2$  is simplified to a simple sum of squares evaluation when such modified variables are used. The pivotal condensation approach was used to transform the data (Singh and Chaudhary, 1977). The coefficients for the transformation were obtained by dividing the first row of the reduced matrix by the square root of the corresponding pivotal condensation elements.

### **3.10.6. Determination of group constellations**

Because a cluster is a poorly defined concept, no guidelines can be established for locating them. The only requirement seems to be that any two groups belonging to the same cluster should have a lower  $D^2$  score on average than two groups belonging to separate clusters. The genotypes were classified using Tocher's technique (Rao, 1952).

#### **3.10.6.1. Average intra cluster distance**

The intra cluster distance was calculated at  $\sum D_i^2/n$ ; where,  $\sum D_i^2$  is the sum of distances between all possible combinations (n) of the population included in a cluster.

#### **3.10.6.2. Average inter-cluster distance**

The average inter cluster distance was calculated as  $\sum D_i/(n_i n_j)$  where  $n_i$  is the number of populations in cluster-I and  $n_j$  is the number of populations in cluster j.

#### **3.10.6.3. Cluster mean**

Cluster means were calculated for individual characters on the basis of mean performance of the genotypes included in that cluster.

#### **3.10.6.4. Cluster diagram**

With the help of  $D^2$  values between the clusters, a diagram showing the relationship between different populations was drawn.

#### **3.10.6.5. Relative contribution of characters towards genetic diversity**

With the help of  $D^2$  statistics, the relative contribution of each component traits to the total divergence was worked out.



### **.10.7. Principal component analysis:**

The variables under investigation are frequently highly linked, and as a result, they effectively communicate the same thing. The original collection of variables can be transformed into a new set of uncorrelated variables known as principal components. These new variables are linear combinations of original variables that are derived in decreasing order of significance, with the first principal component accounting for as much variance in the original data as feasible. PCA is also a linear dimensional reduction approach that finds orthogonal directions of maximum variance in the original data and projects it into a lower-dimensional space made up of a subset of the highest variance component.

Let  $X_1, X_2, X_3, \dots, X_p$  are the variables under study, then first principal component may be define as

$$Z_1 = a_{11}X_1 + a_{12}X_2 + \dots + a_{1p}X_p$$

Such that variance of  $Z_1$  is as large as possible subject to the condition that

$$a_{11}^2 + a_{12}^2 + \dots + a_{1p}^2 = 1$$

This constraint is introduced because if this is not done, then  $\text{var}(Z_1)$  can be increased simply by multiplying any  $a_{1j}$ s by a constant factor. The second principal component define as

$$Z_2 = a_{21}X_1 + a_{22}X_2 + \dots + a_{2p}X_p$$

Such that  $\text{var}(Z_2)$  is as large as possible to  $\text{var}(Z_1)$  subject to the constraint that

$$a_{21}^2 + a_{22}^2 + \dots + a_{2p}^2 = 1 \text{ and } \text{cov}(Z_1, Z_2) = 0 \text{ and so on.}$$

*Chapter 4*

***Result and  
Discussion***

## Chapter 4

# RESULT AND DISCUSSION.

---

### 4.1. Study on Morpho-phenetic traits

#### 4.1.a. Analysis of variance (ANOVA).

The ANOVA for the 12 characters indicated significant differences between the genotypes for 50% Heading, plant height, awn length, spike length, spikelet per spike, 1000 grain weight and biological yield. This revealed that there was substantial variability present among the wheat genotypes for those traits (Table 4.1). Replications were also found to be differed significantly for traits such as plant height, spikelet per spike, tiller per meter, 1000 grain weight, biological yield and harvest index. This indicates the correct choice of design for statistical analysis.

#### 4.1.b. Mean performance along with different genetic components

The mean values for the characters were produced in the Table 4.2 whereas the other genotypic parameters were given in Table 4.3.

#### Germination per meter

The best performance for germination per meter was exhibited by ENTRY 3 (65) and ENTRY 2 (62.5). The lowest performance was exhibited by ENTRY 32 (33.5), ENTRY 35 (33.5), ENTRY 33 (34.65), ENTRY 26 (35), ENTRY 18 (35.65), ENTRY 31 (36), ENTRY 41 (36), ENTRY 29 (36.84), ENTRY 34 (37.17), ENTRY 21 (38.67) and ENTRY 6 (39.84). CV is moderate for this trait (19.75%) while heritability (0.089) and genetic advance (3.78%) is poor for this trait.

#### Days to 50% heading

The highest days to 50% heading was exhibited by ENTRY 22 (78), ENTRY 21 (76.5), ENTRY 36 (76.5), ENTRY 39 (74), ENTRY 29 (73), ENTRY 25 (72), ENTRY 26 (72), ENTRY 34 (72), ENTRY 1 (71), ENTRY 7 (71), ENTRY 14 (71), ENTRY 15 (71), ENTRY 28 (71) and ENTRY 45 (70). The shortest time required was by ENTRY 19 (58), ENTRY 31 (58) and ENTRY 11 (58.5). CV is found to be low (4.71%) while heritability (0.645) is quite high and genetic advance (10.5%) is moderate for this trait.

**Table 4.1. : Analysis of variance of morpho phenetic traits**

Source	df	Mean sum of Square											
		GER	DF	PH	AL	SL	GPS	SPS	TM	TGW	GY	BY	HI
Replication	1	13.915	5.760	996.288**	0.014	1.000	37.450	9.120*	643.470**	151.610**	0.000	61.202**	0.121**
Genotype	49	90.434	45.644**	38.959**	1.957***	3.752*	60.970	3.386**	79.230	24.124**	0.437	3.628**	0.007
Error	49	75.696	9.862	18.589	0.227	2.334	42.660	1.430	65.650	8.248	0.400	1.917	0.006

\*significant at 5% probability level, \*\*significant at 1% probability level.

**GER**= Germination per meter, **DF**= 50% Heading, **PH**= Plant Height (cm), **AL**= Awn Length (cm), **SL**= Spike Length (cm), **GPS**= Grain Per Spike, **SPS**= Spikelet Per Spike, **TM**= Tiller per Meter, **TGW**= 1000 Grain Weight (g), **GY**= Grain Yield (t<sup>ha</sup>), **BY**= Biological Yield (t<sup>ha</sup>), **HI**= Harvest Index

**Table 4.2. : Mean values of morpho phenetic traits**

<b>Genotypes</b>	<b>GER</b>	<b>DF</b>	<b>PH</b>	<b>AL</b>	<b>SL</b>	<b>GPS</b>	<b>SPS</b>	<b>TM</b>	<b>TGW</b>	<b>GY</b>	<b>BY</b>	<b>HI</b>
<b>1</b>	42.33	71	98.3	7.6	13.3	61.5	20.7	55.17	43.01	3.448	12.770	0.267
<b>2</b>	62.5	66	90	7.1	10.1	56.1	21.6	69.335	40.65	4.036	12.030	0.337
<b>3</b>	65	66	91.7	6.8	9.4	60.6	19.8	66.335	36.7	3.654	10.450	0.353
<b>4</b>	45.5	66	93.8	8.3	10	49.1	19.5	58.665	39.58	3.644	12.070	0.309
<b>5</b>	41.835	66	85.4	7.6	10	44.2	18.9	52.165	41.13	3.380	7.360	0.492
<b>6</b>	39.835	64	84.4	7.1	9.4	52.6	17.3	57.17	37.07	3.018	9.420	0.336
<b>7</b>	45.5	71	92.7	7.4	11.2	48.5	19.1	59.5	37.51	3.654	9.380	0.396
<b>8</b>	44.835	63	89.1	7.8	10.2	54.2	18.6	57.165	44.49	2.996	8.090	0.371
<b>9</b>	41.335	67	89.2	7.6	10.3	54.7	20.1	60.835	44.38	3.820	9.400	0.409
<b>10</b>	52.5	61.5	85.6	6.4	9.5	49.9	21	64.33	39.29	3.278	8.370	0.394
<b>11</b>	56.17	58.5	87.5	6.3	8.9	45.9	18.3	57.165	42.17	3.594	8.920	0.403
<b>12</b>	42.17	61.5	82.1	6.6	10.3	43.4	17.8	67.5	46.81	3.080	8.300	0.372
<b>13</b>	42.17	61	83.2	7.4	9.3	49.4	18.5	65	35.75	3.738	8.010	0.476
<b>14</b>	44.5	71	89.6	8.5	8.9	43.3	15.6	60.67	37.36	2.982	8.450	0.352
<b>15</b>	36.665	71	84.6	7.3	9.3	50.2	18.3	57.835	36.47	3.286	8.690	0.380
<b>16</b>	41.665	65.5	85.1	8	10.1	46.5	17.4	61.165	45.84	3.246	7.720	0.425
<b>17</b>	45.83	62	79.2	6.6	9.3	43	17.3	72.335	43.89	2.938	8.280	0.354
<b>18</b>	35.665	62	83.6	7.8	9.7	40	15.4	62.5	38.75	3.768	8.790	0.437
<b>19</b>	44.665	58	85.3	7.3	17.7	54.9	18.7	58	39.72	4.302	8.760	0.503
<b>20</b>	42.83	66.5	85.6	6.9	10.5	46.1	19.6	72.165	38.33	3.848	9.030	0.429
<b>21</b>	38.67	76.5	86.4	6.7	9.8	44.7	17.7	61	39.86	2.546	8.460	0.299
<b>22</b>	46.5	78	85.2	7	9.4	44.8	17.4	62	39.69	3.036	7.800	0.398
<b>23</b>	47	66	80.8	7.8	11.5	50.9	19.2	57.665	35.94	3.400	9.430	0.361
<b>24</b>	51.165	68	91.4	7.3	10.1	54.4	19.1	66	34.73	3.808	9.920	0.384
<b>25</b>	42.665	72	82.3	7.3	9.9	51.8	17.2	72.165	37.3	3.434	9.100	0.367
<b>26</b>	35	72	91.1	7.3	10.4	45.3	18.8	62.335	36.32	3.224	9.480	0.357
<b>27</b>	44	67	95.4	6.9	9.8	48.8	19.4	54.665	37.48	3.434	9.640	0.366

<b>Genotypes</b>	<b>GER</b>	<b>DF</b>	<b>PH</b>	<b>AL</b>	<b>SL</b>	<b>GPS</b>	<b>SPS</b>	<b>TM</b>	<b>TGW</b>	<b>GY</b>	<b>BY</b>	<b>HI</b>
<b>28</b>	52.5	71	93.5	7.4	10.7	45.8	19.4	69.17	42.1	3.310	9.940	0.335
<b>29</b>	36.835	73	91.1	7.7	10.2	49	18.3	58.33	44.19	3.342	9.600	0.348
<b>30</b>	44	66	85.6	7.2	10.1	53.6	19.4	61.5	41.64	3.288	10.020	0.329
<b>31</b>	36	58	81.2	6.5	9.4	46.6	19.5	48.17	36.81	2.824	7.610	0.368
<b>32</b>	33.5	60	81.4	6.7	9.9	51.4	18.7	47.5	36.47	3.200	6.330	0.553
<b>33</b>	34.665	66	81.2	7.2	9.9	53.3	18.3	65.165	41.84	2.666	6.770	0.407
<b>34</b>	37.17	72	93.3	8.4	10.4	51.9	19	59.665	43.88	2.912	8.370	0.359
<b>35</b>	33.5	65	92.2	6.8	11.3	41.7	18.4	51.165	42.39	2.300	8.380	0.270
<b>36</b>	47.665	76.5	87.8	6.8	11.5	53.8	20.5	63.83	41.12	3.434	9.900	0.344
<b>37</b>	43.33	65	80.7	1.8	10.3	53.8	20.9	55.335	42.23	2.632	7.110	0.373
<b>38</b>	42.835	66	86.6	6.8	10	36	17.1	59.335	34.09	3.818	8.190	0.474
<b>39</b>	41.665	74	87.1	7.7	11.6	49.4	18.4	60.335	32.265	3.198	8.910	0.364
<b>40</b>	40.665	65	87.5	6.8	9.8	39.2	18.2	65.5	34.695	2.562	6.880	0.371
<b>41</b>	36	65.5	82.4	7.9	10.7	36.8	17.2	57.665	42.585	2.834	6.720	0.421
<b>42</b>	40.335	67	85.9	6.9	9.8	46.6	18.6	64.17	40.1	3.170	8.130	0.389
<b>43</b>	49.665	65	87.2	7.7	9.3	50.5	17.3	54	41.47	3.624	9.010	0.407
<b>44</b>	48.165	67	87.8	6.1	9.3	45.3	19.8	59	44.985	3.080	8.680	0.352
<b>45</b>	44.835	70	81.1	5.8	8.8	50.1	19.3	65.665	44.57	2.482	6.560	0.379
<b>46</b>	49.665	69.5	83.3	6.4	9.4	54.7	19.3	62.165	38.01	2.438	7.570	0.318
<b>47</b>	42	66	85.9	7.1	9.8	42.5	17.7	51.5	36.045	2.982	8.770	0.341
<b>48</b>	49.835	63	80	7.4	8.5	44.2	16.1	76.335	34.675	2.950	7.300	0.414
<b>49</b>	53	60.5	83.9	7.5	10	43.5	18.9	71.5	38.665	2.846	8.910	0.333
<b>50</b>	46	62	85.5	8.9	9.6	50.5	17.9	61	40.37	3.776	10.136	0.376
<b>mean</b>	<b>44.05</b>	<b>66.62</b>	<b>86.62</b>	<b>7.12</b>	<b>10.17</b>	<b>48.50</b>	<b>18.61</b>	<b>61.18</b>	<b>39.71</b>	<b>3.24</b>	<b>8.76</b>	<b>0.38</b>
<b>C.D.</b>	<b>17.49</b>	<b>6.33</b>	<b>8.691</b>	<b>0.96</b>	<b>3.079</b>	<b>13.13</b>	<b>2.41</b>	<b>16.28</b>	<b>5.789</b>	<b>1.33</b>	<b>2.78</b>	<b>0.17</b>
<b>SE(m)</b>	<b>6.152</b>	<b>2.221</b>	<b>3.049</b>	<b>0.337</b>	<b>1.08</b>	<b>4.619</b>	<b>0.846</b>	<b>5.729</b>	<b>2.031</b>	<b>0.47</b>	<b>0.98</b>	<b>0.06</b>
<b>C.V.</b>	<b>19.753</b>	<b>4.714</b>	<b>4.978</b>	<b>6.684</b>	<b>15.019</b>	<b>13.467</b>	<b>6.425</b>	<b>13.243</b>	<b>7.232</b>	<b>20.38</b>	<b>15.81</b>	<b>21.78</b>

**GER**= Germination per meter, **DF**= 50% Heading, **PH**= Plant Height (cm), **AL**= Awn Length (cm), **SL**= Spike Length (cm), **GPS**= Grain Per Spike, **SPS**= Spikelet Per Spike, **TM**= Tiller per Meter, **TGW**= 1000 Grain Weight (g), **GY**= Grain Yield (t<sup>ha</sup>), **BY**=Biological Yield (t<sup>ha</sup>), **HI**= Harvest Index.

### **Plant height (cm)**

The best performance was exhibited by ENTRY 1 (98.3), ENTRY 27 (95.4), ENTRY 4 (93.8), ENTRY 28 (93.5) and ENTRY 34 (93.3). The shortest was ENTRY 17 (79.2), ENTRY 48 (80), ENTRY 37 (80.7) and ENTRY 23 (80.8). CV is found to be low (4.98%) while heritability (0.354) is medium and genetic advance (4.52%) is poor for this trait.

### **Awn length (Cm)**

The best performance was shown by ENTRY 50 (8.9), ENTRY 14 (8.5), ENTRY 34 (8.4), and ENTRY 4 (8.3). The shortest was ENTRY 37 (1.8). CV is found to be low (6.68%) while heritability (0.792) is quite high and genetic advance (23.95%) is high for this trait.

### **Spike length (Cm)**

The best performance was given by ENTRY 19 (17.7). The shortest was ENTRY 48 (8.5), ENTRY 45 (8.8), ENTRY 11 (8.9) and ENTRY 14 (8.9). CV is found to be high (15.02%) while heritability (0.233) is low and genetic advance (8.23%) is low for this trait.

### **Grain per spike**

The genotypes showing highest grain per spike are ENTRY 1 (61.5) and ENTRY 3 (60.6). The lowest grain per spike is shown by ENTRY 38 (36), ENTRY 41 (36.8) and ENTRY 40 (39.2). CV is found to be high (13.47%) while heritability (0.177) is low and genetic advance (5.4%) is low for this trait.

### **Spikelet per spike**

The highest spikelet per spike was shown by ENTRY 2 (21.6), ENTRY 10 (21), ENTRY 37 (20.9), ENTRY 1 (20.7), ENTRY 36 (20.5) and ENTRY 9 (20.1). The lowest spikelet per spike is shown by ENTRY 18 (15.4), ENTRY 14 (15.6) and ENTRY 48 (16.1). CV is found to be low (6.43%) while heritability (0.406) is medium and genetic advance (6.98%) is low for the trait.

### **Tiller per meter**

The maximum tiller per meter was shown by ENTRY 48 (76.34), ENTRY 17 (72.34), ENTRY 20 (72.17) and ENTRY 25 (72.17). The minimum tiller per meter was shown by ENTRY 32 (47.5) and ENTRY 31 (48.17). CV is found to be high (13.24%) while heritability (0.094) and genetic advance (2.69%) is low for the trait.

### **1000 grain weight (g)**

The highest 1000 grain weight was given by ENTRY 12 (46.81), ENTRY 16 (45.84), ENTRY 44 (44.99), ENTRY 45 (44.57), ENTRY 8 (44.49), ENTRY 9 (44.38) and ENTRY 29 (44.19). The lowest 1000 grain weight was given by ENTRY 39 (32.27). CV is found to be low (7.23%) while heritability (0.49) is medium and genetic advance (10.24%) is moderate for the trait.

### **Grain yield (t/ha)**

The highest grain yield was given by ENTRY 19 (4.302) and ENTRY 2 (4.036). The lowest grain yield was provided by ENTRY 35 (2.3), ENTRY 46 (2.438) and ENTRY 45 (2.482). CV is found to be quite high (20.38%) while heritability (0.044) is low and genetic advance (1.82%) is low for this trait.

### **Biological yield (t/ha)**

The highest biological yield was provided by ENTRY 1 (12.77), ENTRY 4 (12.07) and ENTRY 2 (12.03). The lowest biological yield was provided by ENTRY 32 (6.33), ENTRY 45 (6.56), ENTRY 41 (6.72), ENTRY 33 (6.77) and ENTRY 40 (6.88). CV is found to be high (15.81%) while heritability (0.309) is medium and genetic advance (12.08%) is moderate for the trait.

### **Harvest index**

The highest harvest index was provided by ENTRY 32 (0.553) and ENTRY 19 (0.503). The lowest harvest index was provided by ENTRY 1 (0.267), ENTRY 35 (0.270) and ENTRY 21 (0.299). CV is quite high (21.78%) while heritability (0.077) is low and genetic advance (3.36%) is low for this trait.



**Table 4.3. : Genetic parameters for different morpho-phenetic characters of 50 genotypes of wheat**

S. No.	Character	Mean	Range	C.V	P.C.V	G.C.V	H <sup>2</sup> <sub>bs</sub>	GA %
1	GER	44.05	33.50-65.00	19.75	20.69	6.16	0.089	3.78
2	DF	66.62	58.00-78.00	4.71	7.91	6.35	0.645	10.5
3	PH	86.62	79.20-98.30	4.98	6.19	3.68	0.354	4.52
4	AL	7.12	1.80-8.90	6.68	14.68	13.06	0.792	23.95
5	SL	10.17	8.50-17.70	15.02	17.15	8.28	0.233	8.23
6	GPS	48.5	36.00-61.50	13.47	14.84	6.24	0.177	5.4
7	SPS	18.61	15.40-21.60	6.43	8.34	5.31	0.406	6.98
8	TM	61.18	47.50-76.34	13.24	13.91	4.26	0.094	2.69
9	TGW	39.71	32.27-46.81	7.23	10.13	7.1	0.49	10.24
10	GY	3.24	2.30-4.30	20.38	19.97	4.2	0.044	1.82
11	BY	8.76	6.33-12.77	15.81	19.01	10.56	0.309	12.08
12	HI	0.38	0.27-0.55	21.78	21.22	5.88	0.077	3.36

GER= Germination per meter, DF= 50% Heading, PH= Plant Height (cm), AL= Awn Length (cm), SL= Spike Length (cm), GPS= Grain Per Spike, SPS= Spikelet Per Spike, TM= Tiller per Meter, TGW= 1000 Grain Weight (g), GY= Grain Yield (t<sup>ha</sup>), BY= Biological yield (t<sup>ha</sup>), HI= Harvest Index. CV= Coefficient of Variance, PCV= Phenotypic correlation Coefficient, GCV= Genotypic Correlation coefficient, H<sup>2</sup><sub>bs</sub>= Heritability (broad sense), GA%= Genetic advance as % of mean.

## 4.2. Study on physiological characters

### 4.2.a. Chlorophyll Index (CI)

The Chlorophyll index studies at four different crop growth stages (88 DAS, 95 DAS, 102 DAS and 109 DAS) showed significant difference among the genotypes as well as growth stages. However, genotype x growth stages interaction was found non-significant (Table 4.4a). The mean value of CI indicated gradual decline of chlorophyll index with advances in growth stages (Table 4.4c). This might be due to gradual decay in chlorophyll pigmentation with the maturity of crop. However, the rate of decrease in CI value was found highly variable among the 50 genotypes under study.

**Table 4.4a: Two-way Analysis of variance of Chlorophyll Index values**

Source of variation	df	Mean sum of square	F pr.
Genotype	49	1629.00*	0.013
Growth stage	3	112427.00***	<.001
Genotype x Growth stage	147	962.00	0.636
Error	199	1016.00	

**Table 4.4b: Statistical parameters of Chlorophyll Index values**

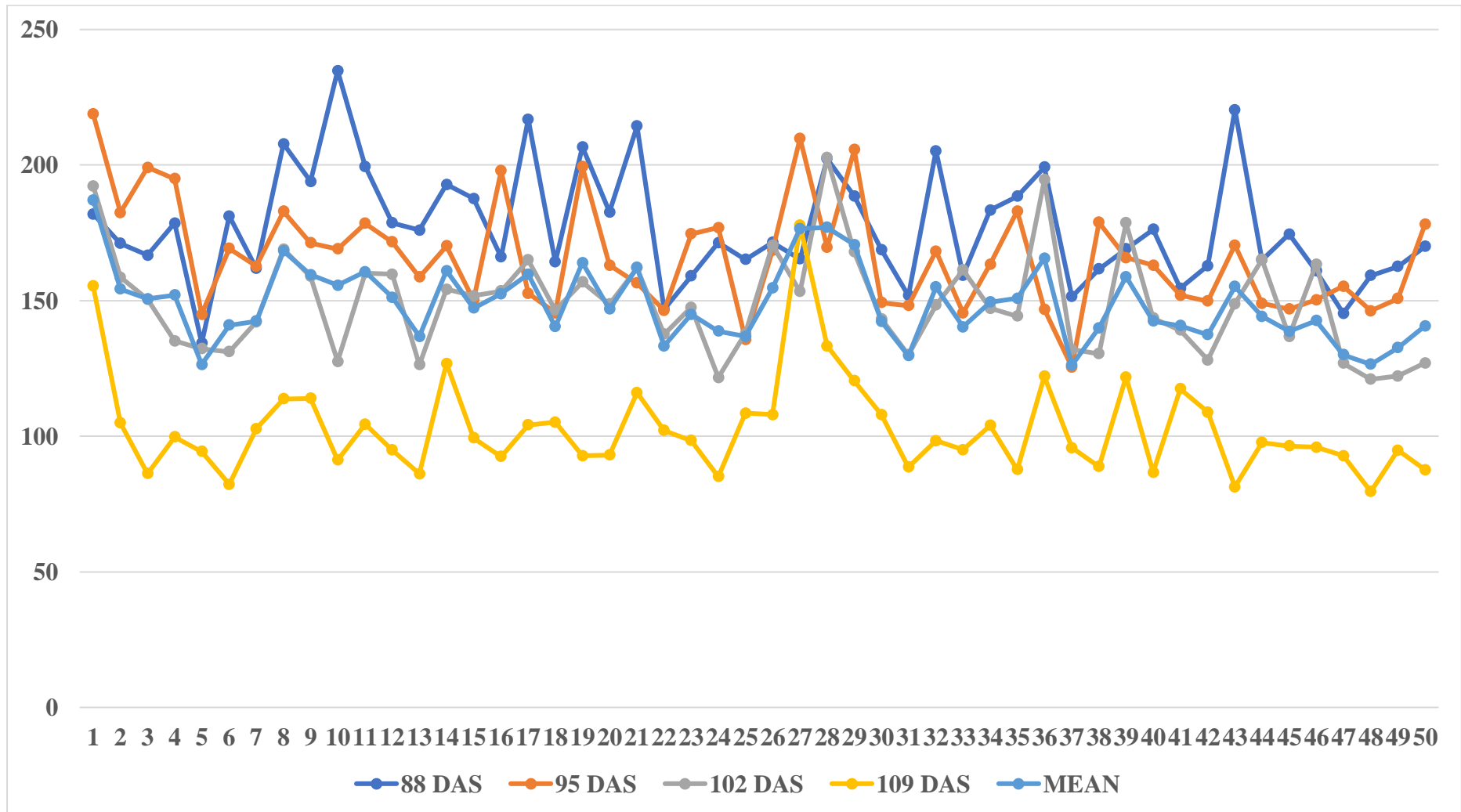
Factors	C.D.	SE(d)	SE(m)
Genotype	31.459	15.941	11.272
Growth Stage	8.898	4.509	3.188
Genotype X Growth stage	62.868	31.881	22.543

In the first growth stage the maximum CI value is shown by ENTRY 10 (234.800) whereas the minimum value is shown by ENTRY 5 (134.300). In the second growth stage the maximum value is shown by ENTRY 1 (218.800) whereas minimum by ENTRY 37 (125.5). In the third growth stage the maximum value shown by ENTRY 28 (202.700) whereas minimum vale by ENTRY 48 (121). In the fourth growth stage the maximum value is shown by ENTRY 27 (177.800) and minimum value by ENTRY 48 (79.6). Fig 1 is the graphical representation of the Chlorophyll Index along with the growth stages.

**Table 4.4c. : Two-way mean table for Chlorophyll Index under different growth stages**

<b>GENOTYPE</b>	<b>88 DAS</b>	<b>95 DAS</b>	<b>102 DAS</b>	<b>109 DAS</b>
1	181.900	218.800	192.300	155.400
2	171.100	182.500	158.600	104.900
3	166.800	199.100	150.500	86.200
4	178.600	195.000	135.100	99.700
5	134.300	144.900	132.300	94.400
6	181.200	169.300	131.200	82.300
7	161.900	162.700	142.200	102.800
8	207.800	182.900	169.000	113.800
9	193.900	171.300	158.900	114.000
10	234.800	169.100	127.600	91.200
11	199.500	178.500	160.200	104.400
12	178.700	171.700	159.700	94.900
13	176.000	158.700	126.400	86.100
14	192.800	170.200	154.200	126.800
15	187.700	150.200	151.800	99.400
16	166.100	197.900	153.500	92.500
17	216.900	152.700	165.000	104.200
18	164.300	145.500	146.500	105.200
19	206.700	199.400	156.900	92.800
20	182.700	163.100	148.700	93.100
21	214.400	156.500	162.000	116.100
22	146.900	146.400	137.600	102.100
23	159.100	174.600	147.500	98.400
24	171.400	176.900	121.600	85.100
25	165.200	135.700	137.900	108.500
26	171.500	168.500	170.500	108.000
27	165.400	209.800	153.400	177.800
28	202.400	169.600	202.700	133.300
29	188.600	205.700	168.000	120.400
30	168.800	149.300	143.300	108.000
31	151.900	148.200	130.100	88.600
32	205.200	168.200	148.400	98.300
33	159.400	145.500	161.300	95.000
34	183.400	163.400	147.200	104.000
35	188.500	183.000	144.300	87.700
36	199.200	146.800	194.600	122.100
37	151.500	125.500	132.000	95.800
38	161.700	178.900	130.400	88.800
39	169.100	165.800	178.700	121.800
40	176.300	163.100	143.600	86.700
41	154.600	152.000	139.100	117.500
42	162.900	149.800	128.100	108.900
43	220.400	170.500	148.800	81.300
44	165.000	149.000	165.200	97.700
45	174.500	146.900	136.800	96.400
46	160.900	150.300	163.400	95.900
47	145.300	155.300	126.900	92.800
48	159.300	146.200	121.000	79.600
49	162.600	150.800	122.200	94.800
50	170.000	178.200	127.000	87.600
<b>Mean</b>	<b>177.182</b>	<b>166.278</b>	<b>149.084</b>	<b>102.862</b>

**Fig 1. Graphical representation of Chlorophyll Index**



To quantify the rate in decrease of CI value Area Under Chlorophyll Index Progress Curve (AUCIPC) was estimated as per formula given by Rosyara *et al.*, (2007). Highest value of AUCIPC was shown by the genotype ENTRY 1 (2828.75) while the lowest value was produced by genotype ENTRY 37 (1,867.57) [Table 4.5]. High AUCIPC indicates higher retention of chlorophyll at maturity. Thus, genotypes such as ENTRY 1, 8, 19, 27, 28 and 29 were having high chlorophyll efficiency at maturity stage. Similar result was found by Rosyara *et al.*, (2010) where chlorophyll content was measured by SPAD reading and AUSDC (Area Under SPAD Decline Curve) value was found significant among genotypes after anthesis.

**Table 4.5. : AUCIPC values pertaining to different genotypes under study**

<b>Genotype</b>	<b>AUCIPC 1</b>	<b>AUCIPC 2</b>	<b>AUCIPC 3</b>	<b>Mean AUCIPC</b>
1	3,806.65	3,288.80	1,390.80	2,828.75
2	3,359.20	2,728.80	1,054.00	2,380.67
3	3,476.05	2,796.80	946.8	2,406.55
4	3,549.20	2,640.80	939.2	2,376.40
5	2,652.40	2,217.60	906.8	1,925.60
6	3,329.75	2,404.00	854	2,195.92
7	3,083.70	2,439.20	980	2,167.64
8	3,711.65	2,815.20	1,131.20	2,552.68
9	3,469.40	2,641.60	1,091.60	2,400.87
10	3,837.05	2,373.60	875.2	2,361.95
11	3,591.00	2,709.60	1,058.40	2,453.00
12	3,328.80	2,651.20	1,018.40	2,332.80
13	3,179.65	2,280.80	850	2,103.49
14	3,448.50	2,595.20	1,124.00	2,389.24
15	3,210.05	2,416.00	1,004.80	2,210.28
16	3,458.00	2,811.20	984	2,417.74
17	3,511.20	2,541.60	1,076.80	2,376.54
18	2,943.10	2,336.00	1,006.80	2,095.30
19	3,857.95	2,850.40	998.8	2,569.05
20	3,285.10	2,494.40	967.2	2,248.90
21	3,523.55	2,548.00	1,112.40	2,394.65
22	2,786.35	2,272.00	958.8	2,005.72
23	3,170.15	2,576.80	983.6	2,243.52
24	3,308.85	2,388.00	826.8	2,174.55
25	2,858.55	2,188.80	985.6	2,010.98
26	3,230.00	2,712.00	1,114.00	2,352.00
27	3,564.40	2,905.60	1,324.80	2,598.27
28	3,534.00	2,978.40	1,344.00	2,618.80
29	3,745.85	2,989.60	1,153.60	2,629.68
30	3,021.95	2,340.80	1,005.20	2,122.65
31	2,850.95	2,226.40	874.8	1,984.05
32	3,547.30	2,532.80	986.8	2,355.64
33	2,896.55	2,454.40	1,025.20	2,125.38
34	3,294.60	2,484.80	1,004.80	2,261.40
35	3,529.25	2,618.40	928	2,358.55
36	3,287.00	2,731.20	1,266.80	2,428.34
37	2,631.50	2,060.00	911.2	1,867.57
38	3,235.70	2,474.40	876.8	2,195.64
39	3,181.55	2,756.00	1,202.00	2,379.85
40	3,224.30	2,453.60	921.2	2,199.70
41	2,912.70	2,328.80	1,026.40	2,089.30
42	2,970.65	2,223.20	948	2,047.28
43	3,713.55	2,554.40	920.4	2,396.12
44	2,983.00	2,513.60	1,051.60	2,182.73
45	3,053.30	2,269.60	932.8	2,085.24
46	2,956.40	2,509.60	1,037.20	2,167.73
47	2,855.70	2,257.60	878.8	1,997.37
48	2,902.25	2,137.60	802.4	1,947.42
49	2,977.30	2,184.00	868	2,009.77
50	3,307.90	2,441.60	858.4	2,202.63
<b>SE(m)</b>	<b>382.2</b>	<b>338.3</b>	<b>101.5</b>	<b>245.4</b>
<b>L.S.D. (0.05)</b>	<b>1086.2</b>	<b>961.4</b>	<b>288.3</b>	<b>697.3</b>

#### 4.2.b. Canopy Temperature Depression (CTD)

Canopy Temperature Depression studied at two different growth stages (68 DAS and 93 DAS) showed significant difference among the growth stages. However, genotypes and genotype x growth stage interaction were found non-significant (Table 4.6a). The mean values of CTD (Table 4.6c) indicated gradual decline of CTD with advance in growth stages in most of the genotypes except 13 genotypes such as ENTRY 5, 8, 14, 18, 20, 21, 24, 28, 34, 35, 46, 48 and 49 where the CTD increased at later growth stages. This indicated high physiological efficiency for these genotypes under present environmental condition. In some genotypes such as ENTRY 1, 2, 6, 10, 15, 17, 22, 37, 39, 41, 42, 43 and 45 the CTD values even moved to negative one at later stages. One genotype (ENTRY 39) had negative values in both the growth stages (-0.050 in 68 DAS and -0.270 in 93 DAS). This might be due to high canopy temperature due to either poor plant water status or less physiological efficiency. Fig 2 is the graphical representation of CTD along with its growth stages. Similar results were found by earlier workers such as Guendouz *et al.*, 2012 using ten durum wheat varieties to show CTD as an indicator to drought tolerance in semi-arid conditions.

**Table 4.6a. : Two-way Analysis of variance for Canopy Temperature Depression under different growth stages**

Source of variation	df	Mean sum of square	F pr.
Genotype	49	1.064	0.598
Growth stage	1	20.840***	<.001
Genotype x Growth stage	49	0.929	0.784
Error	199	1.140	

**Table 4.6b. : Statistical parameters for Canopy Temperature Depression**

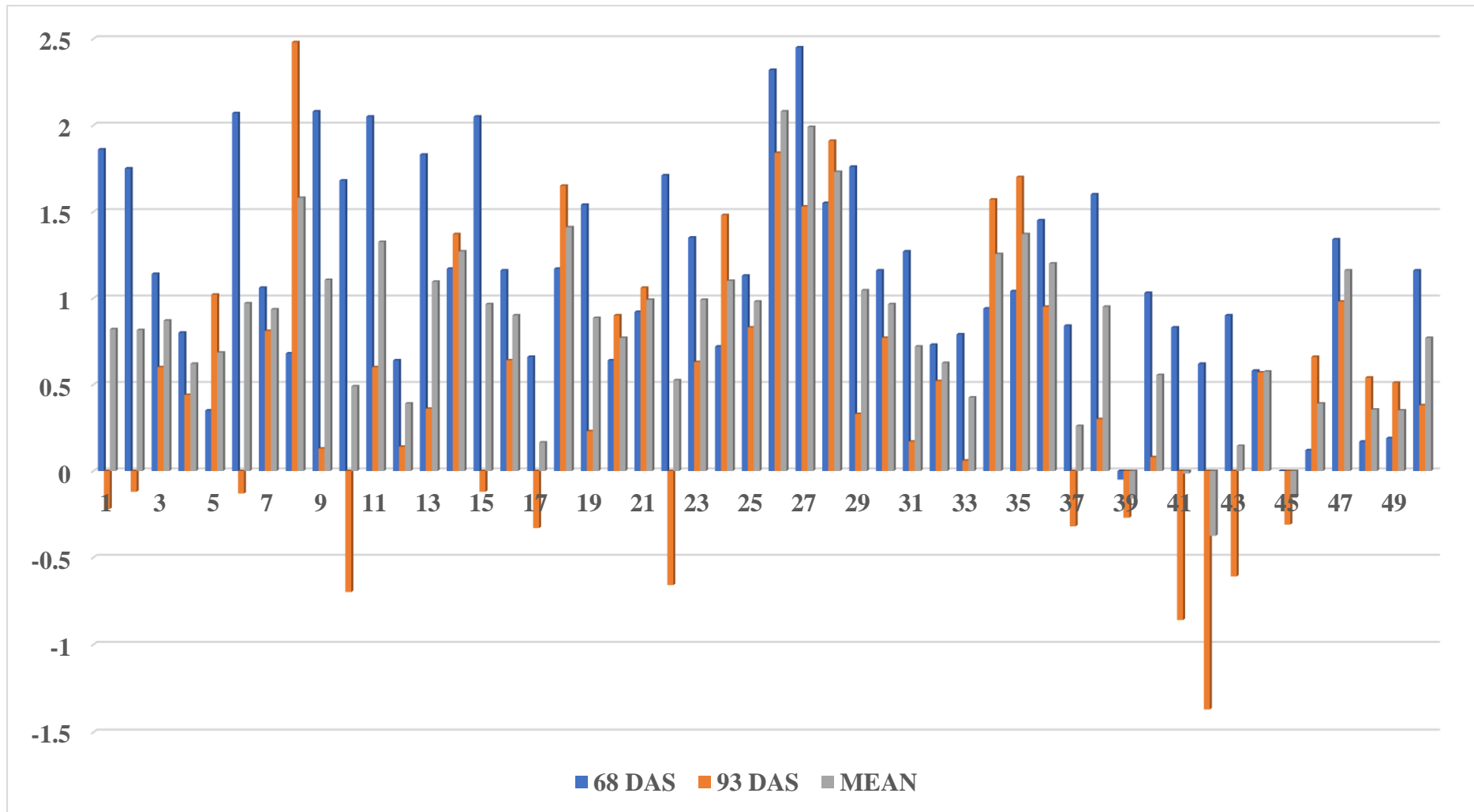
Factors	C.D.	SE(d)	SE(m)
Genotype	1.498	0.755	0.534
Growth Stage	0.300	0.151	0.107
Genotype X Growth stage	2.118	1.067	0.755

**Table 4.6c. : Two-way mean table for Canopy Temperature Depression**

<b>GENOTYPE</b>	<b>68 DAS</b>	<b>93 DAS</b>
1	1.860	-0.220
2	1.750	-0.120
3	1.140	0.600
4	0.800	0.440
5	0.350	1.020
6	2.070	-0.130
7	1.060	0.810
8	0.680	2.480
9	2.080	0.130
10	1.680	-0.700
11	2.050	0.600
12	0.640	0.140
13	1.830	0.360
14	1.170	1.370
15	2.050	-0.120
16	1.160	0.640
17	0.660	-0.330
18	1.170	1.650
19	1.540	0.230
20	0.640	0.900
21	0.920	1.060
22	1.710	-0.660
23	1.350	0.630
24	0.720	1.480
25	1.130	0.830
26	2.320	1.840
27	2.450	1.530
28	1.550	1.910
29	1.760	0.330
30	1.160	0.770
31	1.270	0.170
32	0.730	0.520
33	0.790	0.060
34	0.940	1.570
35	1.040	1.700
36	1.450	0.950
37	0.840	-0.320
38	1.600	0.300
39	-0.050	-0.270
40	1.030	0.080
41	0.830	-0.860
42	0.620	-1.370
43	0.900	-0.610
44	0.580	0.570
45	0.000	-0.310
46	0.120	0.660
47	1.340	0.980
48	0.170	0.540
49	0.190	0.510
50	1.160	0.380
<b>Mean</b>	<b>1.140</b>	<b>0.494</b>



**Fig 2. Graphical representation of Canopy Temperature Depression**



### Corelation between Canopy Temperature, Canopy Temperature Depression, Grain Yield and Biological Yield.

The correlation analysis as shown in Table 4.7 exhibits a positive correlation of grain yield and biological yield with CTD at 68 DAS indicating cooler canopy temperature leading to more grain yield and increase in biomass. Definitely, cooler canopy is an indicator of good plant water status and functions. Similar inference was provided by Karimizadeh *et al.*, 2011, Talebi *et al.*, 2011 and Guendouz *et al.*, 2012, Mondal *et al.*, 2013. Thus, CTD is more important parameter in selecting tolerant genotypes as concluded by Blum *et al.*, 1988 and Balota *et al.*, 2007.

**Table 4.7.: Correlation Matrix between CT, CTD, GY and BY**

Characters	CT <sub>1</sub>	CT <sub>2</sub>	CTD <sub>1</sub>	CTD <sub>2</sub>	GY	BY
CT <sub>1</sub>	1					
CT <sub>2</sub>	-0.012	1				
CTD <sub>1</sub>	-0.529**	-0.014	1			
CTD <sub>2</sub>	0.119	-0.738**	0.021	1		
GY	-0.004	0.008	0.289**	-0.035	1	
BY	-0.109	-0.026	0.362**	0.029	0.477**	1

\*significant at 5% probability level, \*\*significant at 1% probability level, CT<sub>1</sub>= Canopy Temperature at 68 DAS, CT<sub>2</sub>= Canopy Temperature at 93 DAS, CTD<sub>1</sub>= Canopy Temperature Depression at 68 DAS, CTD<sub>2</sub>= Canopy Temperature Depression at 93 DAS, GY= Grain Yield (t<sup>ha</sup>) and BY= Biological Yield (t<sup>ha</sup>).

### 4.3. Study on Spot blotch) resistance

Spot blotch (*Bipolaris sorokiniana* (Sacc.) Shoem) scoring was done at four crop growth stages in double-digit scale (00-99) developed by Saari and Prescott, 1975. Disease severity and AUDPC (Area Under Disease Progress Curve) was calculated as per Wilcoxson *et al.*, (1975).

Analysis of variance clearly revealed highly significant effect of genotype as well as growth stages on disease severity % (Table 4.8a). Interaction between genotype and growth stages was not found significant. Mean values of severity among different growth stages indicated gradual increase in severity along with increase in growth stages (Table 4.8c). This is obvious in spot blotch resistance where disease progress rapidly with the advances in maturity of crop especially in susceptible genotypes (Joshi *et al.*, 2007; Duveiller, 2005).

**Table 4.8a.: Two-way Analysis of variance for Disease severity (%) under different growth stages**

Source of variation	df	Mean sum of square	F pr.
Genotype	49	405.65***	<.001
Growth stage	3	85734.72***	<.001
Genotype x Growth stage	147	66.88	0.301
Error	199	61.81	

**Table 4.8b. : Statistical parameters for Disease severity (%)**

Factors	C.D.	SE(d)	SE(m)
Genotype	7.758	3.931	2.780
Growth Stage	2.194	1.112	0.786
Genotype X Growth stage	15.503	7.862	5.559

**Table 4.8c. : Two-way mean table for Disease severity (%) under different growth stages**

<b>GENOTYPE</b>	<b>88 DAS</b>	<b>95 DAS</b>	<b>102 DAS</b>	<b>109 DAS</b>
1	19.753	28.951	50.556	63.889
2	25.185	42.963	72.901	97.222
3	36.790	41.049	69.877	88.333
4	35.123	42.469	67.901	95.556
5	29.630	44.938	67.099	96.111
6	28.704	42.160	81.790	96.667
7	34.753	38.025	62.531	95.000
8	19.691	33.704	77.284	94.444
9	18.086	28.889	61.790	88.333
10	27.778	47.531	71.481	92.778
11	19.321	34.506	71.358	93.889
12	19.691	27.407	58.765	86.111
13	24.383	41.358	56.358	94.444
14	34.198	40.864	58.704	87.778
15	26.235	31.728	76.667	95.556
16	32.003	48.951	73.889	92.222
17	31.790	48.889	91.728	93.889
18	34.877	42.840	63.210	93.333
19	32.099	47.531	85.494	96.667
20	28.333	45.370	85.000	97.222
21	25.988	24.074	60.432	77.222
22	35.494	54.444	83.889	97.222
23	26.235	35.309	71.605	95.556
24	22.901	19.753	62.531	88.889
25	27.716	29.198	77.963	88.889
26	38.642	41.605	77.346	93.333
27	26.914	39.383	71.667	94.444
28	22.222	32.037	70.864	88.333
29	21.296	28.148	51.667	78.333
30	30.741	48.889	88.333	99.444
31	30.309	56.790	79.444	96.667
32	33.395	49.506	81.420	95.000
33	25.556	47.901	73.333	97.222
34	33.457	41.790	81.111	95.556
35	40.247	57.284	95.556	99.444
36	21.420	26.358	75.617	85.000
37	36.728	48.148	91.111	98.889
38	44.259	49.630	81.667	95.000
39	19.938	30.617	56.481	79.444
40	27.716	41.235	90.000	98.333
41	36.728	43.086	78.704	93.333
42	37.840	41.790	80.247	93.889
43	24.012	44.444	98.889	100.000
44	27.160	41.975	61.235	84.444
45	30.926	37.346	70.741	92.778
46	33.889	46.173	84.444	98.333
47	25.617	34.383	73.889	90.000
48	33.889	52.037	71.111	96.667
49	35.432	44.630	88.765	95.000
50	30.247	59.136	93.025	99.444
<b>Mean</b>	<b>29.307</b>	<b>40.944</b>	<b>74.549</b>	<b>92.511</b>

To identify the progress of disease along with growth stages AUDPC values was calculated for each genotype. Table 4.9 shows the mean values of AUDPC. In AUDPC 1 values the maximum value is shown by ENTRY 35 (341.36) and the minimum value is shown by ENTRY 24 (149.29). In AUDPC 2 the maximum value is exhibited by ENTRY 35 (534.94) whereas minimum by ENTRY 1 (278.27). In AUDPC 3 the maximum value is shown by ENTRY 43 (696.11) whereas minimum by ENTRY 1 (400.555). In the Mean AUDPC values the maximum is shown by ENTRY 35 (519.595) whereas the minimum value by ENTRY 1 (283.095).

Genotypes were classified into resistant (R), moderately resistant (MR), moderately resistant to moderately susceptible (MR-MS), moderately susceptible (MS), moderately susceptible to susceptible (MS-S), susceptible (S), susceptible to highly susceptible (S-HS) and highly susceptible (HS) as per AUDPC values suggested by Liatukas and Ruzgas, 2012. Among the genotypes 29 were highly susceptible (HS), 14 were susceptible to highly susceptible (S-HS) and 06 were susceptible (S). Only one genotype i.e., ENTRY 1 was found moderately susceptible to susceptible (MS-S) which was a local check variety i.e., DBW 187. It indicated that all the germplasm of the present nursery were either highly susceptible or susceptible category. Thus, no genotype could be selected in terms of disease resistance parameter.

**Table 4.9. : AUDPC values pertaining to different genotypes under study**

Genotype	AUDPC 1	AUDPC 2	AUDPC 3	Mean AUDPC	Resistance category
1	170.465	278.270	400.555	283.095	MS-S
2	238.515	405.525	595.430	413.160	HS
3	272.440	388.240	553.735	404.805	HS
4	271.575	386.300	572.100	409.990	HS
5	260.985	392.130	571.235	408.115	HS
6	248.025	433.825	624.595	435.485	HS
7	254.720	351.945	551.355	386.010	S-HS
8	186.885	388.455	601.050	392.130	S-HS
9	164.415	317.380	525.430	335.740	S
10	263.580	416.545	574.905	418.340	HS
11	188.395	370.525	578.360	379.095	S-HS
12	164.845	301.605	507.065	324.510	S
13	230.090	342.005	527.810	366.635	S-HS
14	262.715	348.485	512.685	374.630	S-HS
15	202.870	379.380	602.780	395.010	S-HS
16	283.335	429.940	581.390	431.555	HS
17	282.375	492.160	649.660	474.735	HS
18	272.005	371.170	547.905	397.025	S-HS
19	278.705	465.585	637.560	460.615	HS
20	257.965	456.295	637.780	450.680	HS
21	175.215	295.770	481.790	317.590	S
22	314.785	484.170	633.890	477.615	HS
23	215.405	374.200	585.060	391.555	S-HS
24	149.290	287.995	529.970	322.415	S
25	199.195	375.060	583.985	386.080	S-HS
26	280.865	416.330	597.375	431.525	HS
27	232.035	388.670	581.385	400.700	HS
28	189.905	360.155	557.190	369.085	S-HS
29	173.055	279.350	455.000	302.470	S
30	278.700	480.275	657.220	472.065	HS
31	304.845	476.820	616.390	466.020	HS
32	290.155	458.240	617.465	455.290	HS
33	257.095	424.320	596.945	426.120	HS
34	263.365	430.150	618.335	437.285	HS
35	341.360	534.940	682.500	519.595	HS
36	167.220	356.915	562.160	362.100	S-HS
37	297.070	487.405	665.000	483.155	HS
38	328.610	459.535	618.335	468.830	HS
39	176.945	304.850	475.740	319.175	S
40	241.325	459.320	659.170	453.270	HS
41	279.350	426.265	602.130	435.915	HS
42	278.705	427.130	609.475	438.440	HS
43	239.600	501.665	696.110	479.125	HS
44	241.975	361.235	509.875	371.030	S-HS
45	238.950	378.300	572.315	396.525	S-HS
46	280.215	457.160	639.720	459.035	HS
47	210.000	378.950	573.610	387.520	S-HS
48	300.740	431.020	587.225	439.660	HS
49	280.215	466.885	643.180	463.425	HS
50	312.840	532.565	673.645	506.345	HS
<b>S.Em (±)</b>	<b>26.634</b>	<b>38.896</b>	<b>35.310</b>	<b>29.865</b>	
<b>L.S.D. (0.05)</b>	<b>75.693</b>	<b>110.541</b>	<b>100.349</b>	<b>84.871</b>	

#### 4.4. Genetic divergence analysis (D<sup>2</sup> analysis)

Chi-square test indicated that there was sufficient divergence in the genotypes and hence D<sup>2</sup> analysis (Mahalanobis, P.C., 1936) was done. The distribution of genotypes into 5 different clusters was made on the basis of D<sup>2</sup> analysis based on 15 characters namely GER= Germination per meter, DF= 50% Heading, PH= Plant Height (cm), AL= Awn Length (cm), SL= Spike Length (cm), GPS= Grain per Spike, SPS= Spikelet per spike, TM= Tiller per Meter, AUCIPC= area under chlorophyll index progress curve, CTD= Canopy Temperature Depression, AUDPC=area under disease progression curve, TGW= 1000 Grain Weight (g), BM= Biological yield (t<sup>ha</sup>), GY= Grain Yield (t<sup>ha</sup>), and HI= Harvest Index. Highest no. of genotypes (17) was accommodated in Cluster I and II (Table 4.10) while cluster III contained 12 genotypes. Smallest cluster was cluster IV with only single genotype while Cluster V had 3 only.

The average intra (diagonal) and inter (off-diagonal) cluster D<sup>2</sup> were presented in Table 4.11. The inter-cluster distances were found higher than the intra-cluster distances which indicated the wider genetic diversity among the tested genotypes of different groups. Similar findings were reported by Samal and Jagadev (1996), Ahmed *et al.* (2002) and Zaman *et al.*, 2014. Maximum inter cluster D<sup>2</sup> was exhibited by the Cluster I and V (1243.168) closely followed by Cluster I and IV (1138.777). The other inter cluster differences such as Cluster I and III (877.550), Cluster II and IV (472.026) and Cluster II and V (420.085) was found moderate to low. High intra cluster distance was exhibited by cluster II (101.703) closely followed by Cluster I (101.444) as they contained highest number of genotypes. From the intra and inter cluster D<sup>2</sup> values, it was revealed that the genotypes from the most divergent clusters like I and V could be selected as parents for hybridisation to get wide spectrum of variation within the segregating material.

**Table 4.10. : Distribution of 50 wheat genotypes into different clusters.**

Cluster No.	Number of genotypes	Name of genotypes
<b>I</b>	17	ENTRY 1, ENTRY 2, ENTRY 3, ENTRY 4, ENTRY 5, ENTRY 6, ENTRY 7, ENTRY 8, ENTRY 9, ENTRY 10, ENTRY 11, ENTRY 12, ENTRY 13, ENTRY 14, ENTRY 15, ENTRY 16, ENTRY 17
<b>II</b>	17	ENTRY 18, ENTRY 19, ENTRY 20, ENTRY 21, ENTRY 22, ENTRY 23, ENTRY 24, ENTRY 25, ENTRY 26, ENTRY 27, ENTRY 28, ENTRY 29, ENTRY 30, ENTRY 31, ENTRY 33, ENTRY 34, ENTRY 35
<b>III</b>	12	ENTRY 32, ENTRY 36, ENTRY 38, ENTRY 39, ENTRY 40, ENTRY 41, ENTRY 42, ENTRY 43, ENTRY 44, ENTRY 45, ENTRY 47, ENTRY 48
<b>IV</b>	1	ENTRY 37
<b>V</b>	3	ENTRY 46, ENTRY 49, ENTRY 50

**Table 4.11. : Average intra (diagonal) and inter (off diagonal) cluster D<sup>2</sup> values of 50 genotypes of wheat**

Cluster No.	I	II	III	IV	V
I	<b>101.444</b>	343.788	877.550	1138.777	1243.168
II		<b>101.703</b>	245.427	472.026	420.085
III			<b>74.069</b>	226.630	119.553
IV				<b>0</b>	322.125
V					<b>75.990</b>

The cluster mean values for 15 characters along with their contribution to divergence have been given in Table 4.12. Highest grain yield as well as biological yield was produced by cluster I (3.399 and 9.277 ton<sup>-ha</sup>. respectively). At the same time, highest chlorophyll retention value was exhibited by cluster I having highest AUCIPC value (2345.88). In terms of disease infestation, it was also found that lowest AUDPC value had been shown by cluster I (390.179) which indicated less disease infestation for this cluster. Thus, it could be concluded that most promising genotypes in terms of yield, physiological efficiency and disease susceptibility have been grouped in cluster I.

The maximum contribution to the genetic divergence was exhibited by the character CTD (80.571%), followed by HI (10.286%), GER (8.163%) and GPS (8.163%). Highest CTD value was shown by Cluster V (12.712) which indicated that promising genotypes for this trait had been accommodated in this cluster.



**Table 4.12. : Cluster mean for 15 characters of wheat genotypes**

Cluster No.	GER	DF	PH	AL	SL	GPS	SPS	TM	TGW	GY	BY	AUCIDC	CTD	AUDPC	HI
<b>I</b>	46.491	65.412	87.735	7.318	9.971	50.182	18.812	61.314	40.712	3.399	9.277	2345.888	3.069	390.179	0.273
<b>II</b>	41.343	68.059	87.365	7.265	10.635	48.447	18.518	61.274	39.469	3.261	8.888	2281.967	7.487	413.438	0.269
<b>III</b>	43.097	67.083	85.058	6.975	9.917	45.483	18.242	60.403	38.589	3.112	7.948	2289.273	10.893	417.240	0.283
<b>IV</b>	43.330	65.000	80.700	1.800	10.300	53.800	20.900	55.335	42.230	2.630	7.110	1867.565	9.755	483.155	0.270
<b>V</b>	49.555	64.000	84.233	7.600	9.667	49.567	18.700	64.888	39.015	3.018	8.872	2126.708	12.712	476.268	0.250
<b>Mean</b>	<b>44.763</b>	<b>65.911</b>	<b>85.018</b>	<b>6.191</b>	<b>10.098</b>	<b>49.496</b>	<b>19.034</b>	<b>60.643</b>	<b>40.003</b>	<b>3.084</b>	<b>8.419</b>	<b>2182.280</b>	<b>8.783</b>	<b>436.056</b>	<b>0.269</b>
<b>Percent contribution to Divergence</b>	<b>8.163</b>	<b>1.141</b>	<b>0.163</b>	<b>2.612</b>	<b>0.489</b>	<b>8.163</b>	<b>0.979</b>	<b>0.327</b>	<b>2.041</b>	<b>0</b>	<b>0</b>	<b>0.163</b>	<b>80.571</b>	<b>1.061</b>	<b>10.286</b>

**GER**= Germination per meter, **DF**= 50% Heading, **PH**= Plant Height (cm), **AL**= Awn Length (cm), **SL**= Spike Length (cm), **GPS**= Grain Per Spike, **SPS**= Spikelet Per Spike, **TM**= Tiller per Meter, **TGW**= 1000 Grain Weight (g), **GY**= Grain Yield (t<sup>ha</sup>), **BY**= Biological Yield (t<sup>ha</sup>), **AUCIPC**= Area Under Chlorophyll Index Progress Curve, **CTD**= Canopy Temperature Depression, **AUDPC**= Area Under Disease Progression Curve, **HI**= Harvest Index

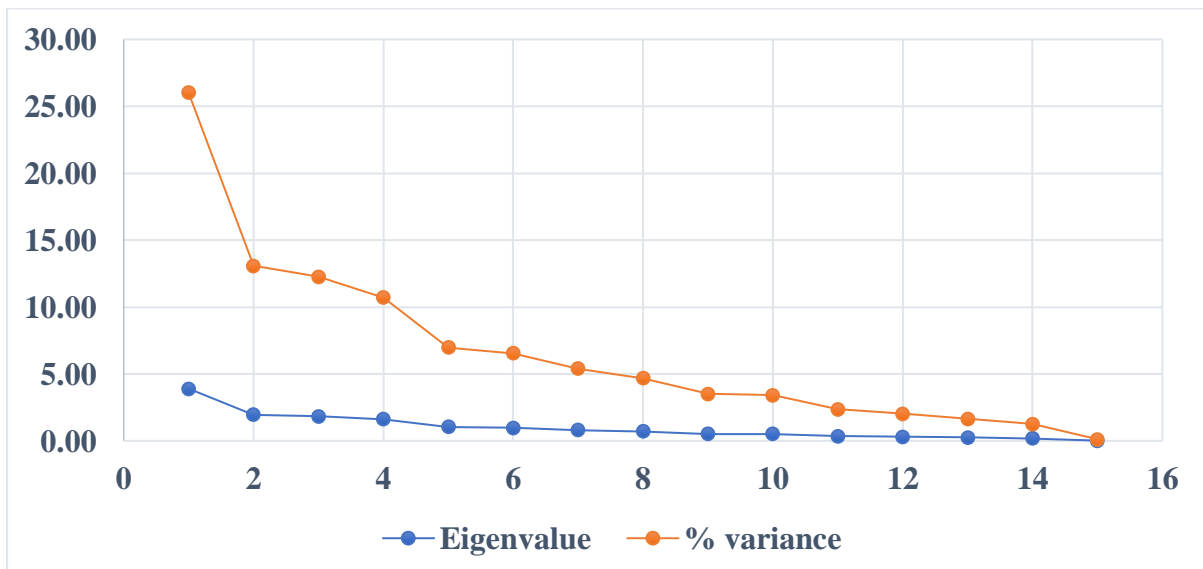
#### 4.5. Principal Component Analysis

Principal component analysis (PCA) is important for the reflection of the highest contributor to the total variation at each axis of differentiation (Sharma *et al.*, 1998). PCA was done by using all the fifteen characters that were used for D<sup>2</sup> analysis. Among the fifteen principal components (PCs) only five components (PCs) had showed Eigen value >1.00, which accounted for 69.03% of cumulative proportion of variance (Table 4.13). Among them PC 1 and PC 2 accounted for 26.01% and 13.08% variance respectively.

**Table 4.13. : Summary of the contribution of the different principal components to variability**

PC	Eigen value	% Variance
1	3.90	26.01
2	1.96	13.08
3	1.84	12.26
4	1.61	10.71
5	1.05	6.97
6	0.98	6.54
7	0.81	5.39
8	0.70	4.68
9	0.53	3.52
10	0.51	3.41
11	0.36	2.37
12	0.30	2.03
13	0.25	1.65
14	0.19	1.27
15	0.02	0.12

**Fig 3: Scree plot analysis for PCA**



Loading value of different characters are presented in Table 4.14. These showed both negative and positive loadings which indicated the presence of positive and negative correlation trends between the components and the variables. Therefore, the characters which loaded high positively and negatively contributed more to the diversity and they were the ones that were responsible for creating difference between clusters. In PC 1, characters such as BY, PH, GPS, AUCIPC had high positive loadings and thus contributed positively while AUDPC and CTD had negative loadings which contributed negatively. In PC 2, GY and HI had high positive loadings while DF had negative loading. The findings of PCA revealed that these effective contributing traits in PC 1 and PC 2 had the significant role in diversification of genotypes and selection may be possible based on this trait for future breeding programmes.

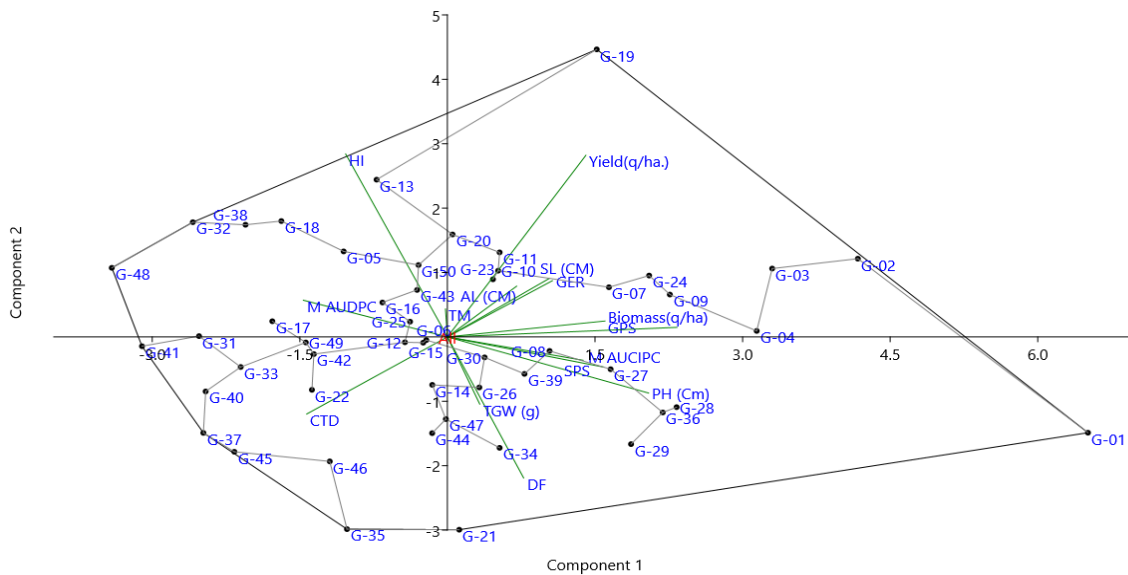
**Table 4.14. : Five principal components along with their factor loadings**

Characters	PC 1	PC 2	PC 3	PC 4	PC 5
GER	0.2044	0.1674	0.4115	0.3483	0.1114
DF	0.1483	-0.4193	-0.1469	0.2245	0.0209
PH	0.3916	-0.1670	-0.1830	-0.0124	0.1806
AL	0.1347	0.1509	-0.5108	0.2437	0.0162
SL	0.1982	0.1738	-0.1084	-0.4096	0.1561
GPS	0.3066	0.0473	0.3282	-0.1541	0.0201
SPS	0.2640	-0.0695	0.4692	-0.2545	0.1169
TM	-0.0038	0.0833	0.2169	0.6022	-0.1689
TGW	0.0629	-0.1999	0.1255	-0.2352	-0.6361
GY	0.2695	0.5402	-0.0723	0.0612	0.0481
BY	0.4469	0.0284	-0.0060	0.1480	0.1758
AUCIPC	0.3043	-0.0897	-0.2653	-0.1880	0.0271
CTD	-0.2739	-0.2293	-0.0084	0.0197	0.4909
AUDPC	-0.2804	0.1092	0.1714	-0.1157	0.4324
HI	-0.1981	0.5442	-0.0849	-0.1403	-0.1441

GER= Germination per meter, DF= 50% Heading, PH= Plant Height (cm), AL= Awn Length (cm), SL= Spike Length (cm), GPS= Grain Per Spike, SPS= Spikelet Per Spike, TM= Tiller per Meter, TGW= 1000 Grain Weight (g), GY= Grain Yield (t<sup>ha</sup>), BY= Biological Yield (t<sup>ha</sup>), AUCIPC= Area Under Chlorophyll Index Progress Curve, CTD= Canopy Temperature Depression, AUDPC= Area Under Disease Progression Curve, HI= Harvest Index

PCA biplot graph (Fig 4) also showed the divergence of the present wheat population with respect to 15 quantitative traits. Strong positive correlation was found between GY, BY, GPS while negative association between GY and DF, CTD and PH. These findings would be helpful for making selection criteria of individual genotypes under present environmental situation.

**Fig 4. PCA Biplot graph showing 50 genotypes by points (G-1 to G-50) and trait vectors as green arrow**



#### 4.6 Hierarchical clustering of 50 genotypes based on Euclidean distance

Hierarchical clustering of 50 genotypes of wheat was done based on Euclidean distance (Fig 5.). It revealed 5 clusters as shown in Table 4.15.

**Table 4.15.: Hierarchical clustering of 50 genotypes of wheat.**

Cluster No.	Number of genotypes	Name of genotypes
I	18	ENTRY 6, ENTRY 7, ENTRY 13, ENTRY 15, ENTRY 18, ENTRY 20, ENTRY 23, ENTRY 24, ENTRY 30, ENTRY 33, ENTRY 34, ENTRY 38, ENTRY 40, ENTRY 41, ENTRY 44, ENTRY 45, ENTRY 46, ENTRY 50
II	8	ENTRY 5, ENTRY 22, ENTRY 25, ENTRY 31, ENTRY 37, ENTRY 42, ENTRY 48, ENTRY 49
III	5	ENTRY 8, ENTRY 19, ENTRY 27, ENTRY 28, ENTRY 29
IV	17	ENTRY 2, ENTRY 3, ENTRY 4, ENTRY 9, ENTRY 10, ENTRY 11, ENTRY 12, ENTRY 14, ENTRY 16, ENTRY 17, ENTRY 21, ENTRY 26, ENTRY 32, ENTRY 35, ENTRY 36, ENTRY 39, ENTRY 43
V	2	ENTRY 1, ENTRY 47

If we compare the both clusters (Table 4.10 and Table 4.15), we found lot of dissimilarity between them. Clustering based on Euclidean distance, grouped the 50 genotypes

into 5 Clusters within which Cluster I had the most genotypes (18), followed by Cluster IV (17). Moreover, Cluster II, III, and V comprised of 8, 5, and 2 genotypes respectively. The probable reason for this dissimilarity is that the Mahalanobis distance is a metric that takes into account both inter-variable correlations and variance differences. Variables with high variation and strongly linked variables are given less weight by the Mahalanobis distance, ensuring that all qualities are treated equally (Mimmack *et al.*, 2001). Whereas the Euclidean distance assigns equal weight to each variable and it gives excess weight to correlated variables (Jolliffe 1986).



## 4.7. Correlation analysis

### Correlation between 15 characters of wheat

The correlation analysis (Table 4.16) revealed that among the 15 characters only eight of them were positively associated with grain yield. Those traits included GER, PH, AL, SL, GPS, AUCIPC, BY and HI. GER was also positively correlated with GPS, SPS, TM, BY and GY. DF was positively correlated with PH and negatively correlated with AUDPC and HI. PH was positively correlated with DF, AL, BY and GY, whereas, negatively correlated with HI. AL was positively correlated with PH, CTD, BY and GY, whereas negatively correlated with SPS. SL was positively correlated with GY only. GPS is positively correlated with GER, SPS, BY and GY. SPS was positively correlated with GER, GPS and BY whereas negatively correlated with AL only. TM was positively correlated with GER only.

Among the physiological traits, AUCIPC was found positively correlated with PH, CTD, BY and GY. Similarly, CTD was positively correlated with AUCIPC and BY. This indicated that with higher AUCIPC value the biomass of the genotypes also increased as well as canopy temperature decreased due to high physiological efficiency. Similar result was found in wheat Khakwani *et al.*, 2012 and Abdul *et al.*, 2018.

AUDPC was found to be positively correlated with HI whereas negatively correlated with DF, PH, GPS, AUCIPC and BY. This indicated that higher disease severity was negatively associated with high chlorophyll index value. This might be due to loss of greenness during high disease infestation. Similar finding was found by Rosyara *et al.*, 2007, Rosyara *et al.*, 2010.

**Table 4.16. : Correlation Matrix between 15 characteristics of wheat.**

Characters	GER	DF	PH	AL	SL	GPS	SPS	TM	AUCIPC	CTD	AUDPC	TGW	BY	GY	HI
<b>GER</b>	1														
<b>DF</b>	-0.044	1													
<b>PH</b>	0.124	0.218*	1												
<b>AL</b>	0.008	0.096	0.236*	1											
<b>SL</b>	-0.055	-0.038	0.118	0.069	1										
<b>GPS</b>	0.396**	0.122	0.115	0.010	0.190	1									
<b>SPS</b>	0.356**	0.107	0.130	-0.262**	0.178	0.642**	1								
<b>TM</b>	0.403**	0.122	-0.152	0.046	-0.124	0.123	0.109	1							
<b>AUCIPC</b>	0.116	0.046	0.442**	0.179	0.190	0.187	0.083	-0.186	1						
<b>CTD</b>	-0.017	0.100	0.312**	0.220*	-0.009	0.004	0.023	0.001	0.294**	1					
<b>AUDPC</b>	-0.088	-0.309**	-0.394**	-0.156	-0.053	-0.199*	-0.104	0.023	-0.262**	-0.021	1				
<b>TGW</b>	0.058	0.002	-0.151	-0.111	0.079	0.151	0.189	0.133	0.040	-0.074	0.029	1			
<b>BY</b>	0.328**	0.115	0.723**	0.249*	0.156	0.357**	0.267**	0.011	0.402**	0.239*	-0.382**	-0.165	1		
<b>GY</b>	0.276**	-0.136	0.286**	0.240*	0.316**	0.230*	0.049	0.090	0.217*	0.144	-0.166	-0.103	0.477**	1	
<b>HI</b>	-0.095	-0.258**	-0.394**	0.006	0.169	-0.088	-0.170	0.052	-0.173	-0.090	0.202*	0.053	-0.473**	0.515**	1

\*significant at 5% probability level, \*\*significant at 1% probability level.

**GER**= Germination per meter, **DF**= 50% Heading, **PH**= Plant Height (cm), **AL**= Awn Length (cm), **SL**= Spike Length (cm), **GPS**= Grain Per Spike, **SPS**= Spikelet Per Spike, **TM**= Tiller per Meter, **AUCIPC**= Area Under Chlorophyll Index Progress Curve, **CTD**= Canopy Temperature Depression, **AUDPC**=Area Under Disease Progression Curve, **TGW**= 1000 Grain Weight (g), **BY**= Biological Yield (t<sup>ha</sup>), **GY**= Grain Yield (t<sup>ha</sup>), **HI**= Harvest Index



*Chapter 5*

***Summary and  
Conclusion***

### SUMMARY AND CONCLUSION.

---

The ANOVA for the 12 characters indicated significant differences between the genotypes for 50% Heading, Plant height, Awn length, Spike length, Spikelet per spike, 1000 grain weight and biological yield.

Mean germination per meter was 44.05 which ranged between 33.5-65 and had a moderate CV (19.75%) and poor heritability (0.089) and genetic advance (3.78%). Range of days to flowering varied between 58 to 78 days with a mean value of 66.62 and it had low CV (4.71%) and high heritability (0.645) and genetic advance (10.5%) is moderate for this trait. Plant height varied between 79.20 cm to 98.30 cm with a mean value of 86.62 cm and low CV (4.98%) and moderate heritability (0.354) and poor genetic advance (4.52%). Mean awn length was 7.12 cm and it varied between 1.80 cm-8.90 cm with low CV (6.68%) and high heritability (0.792), genetic advance (23.95%). The mean of spike length was 10.17 cm which ranged between 8.50 cm-17.70 cm with high CV (15.02%) and low heritability (0.233). The grain per spike had a mean of 48.50 which ranged between 36.00-61.50 with moderate CV (13.47%) and low heritability (0.177). The mean of spikelet per spike was 18.61 which ranged between 15.40-21.60 with low CV (6.43%) and moderate heritability (0.406) and low genetic advance (6.98%). Mean tiller per meter was 61.18 with a range of 47.50-76.34 with moderate CV (13.24%) and very low heritability (0.092) and genetic advance (2.69%). Mean 1000 grain weight was 39.71g and it ranged between 32.27g-46.81g with low CV (7.23%) and moderate heritability (0.490) and genetic advance (10.24%). Mean grain yield was 3.24 t<sup>ha</sup> and it ranged between 2.3-4.3 t<sup>ha</sup> with a high CV (20.38%) and poor heritability (0.044). However, mean biological yield was 8.76 t<sup>ha</sup> and it ranged between 6.33-12.77 t<sup>ha</sup> and heritability (0.309) and genetic advance (12.08%) is moderate for the trait. Harvest index ranged between 0.27-0.55 with a mean value of 0.38, the CV of this trait was high (21.78%) whereas heritability was very poor (0.077).

Among the physiological traits Chlorophyll Index (CI) studied at four different crop growth stages (88 DAS, 95 DAS, 102 DAS and 109 DAS) showed significant difference among the genotypes as well as growth stages. The mean value of CI indicated gradual decline of chlorophyll index with advances in growth stages. However, the rate of decrease in CI value was found highly variable among the 50 genotypes under study. Area Under Chlorophyll Index

Progress Curve (AUCIPC) showed highest value in genotype ENTRY 1 which indicated higher retention of chlorophyll at maturity in this genotype.

Canopy Temperature Depression studied at two different growth stages (68 DAS and 93 DAS) showed significant difference among the growth stages only. 13 genotypes (ENTRY 5, 8, 14, 18, 20, 21, 24, 28, 34, 35, 46, 48 and 49) showed higher CTD value at later stages which indicating high physiological efficiency for these genotypes. Correlation study between CT, CTD, GY and BY revealed positive association of grain yield and biological yield with CTD value at 68 DAS indicating cooler canopy temperature leading to higher grain yield and increased biomass. Definitely, cooler canopy is an indicator of good plant water status and functions. Similar results were shown by Karimizadeh *et al.*, 2011, Talebi *et al.*, 2011 and Guendouz *et al.*, 2012.

Spot blotch (*Bipolaris sorokiniana* (Sacc.) Shoem) scoring was done at four crop growth stages in double-digit scale (00-99). Disease severity and AUDPC (Area Under Disease Progress Curve) was calculated. Analysis of variance clearly revealed highly significant effect of genotype as well as growth stages on disease severity %. Mean disease severity under different growth stages indicated gradual increase in severity with the advancement in maturity. This is obvious in spot blotch resistance where disease progress rapidly with the advances in maturity of crop especially in susceptible genotypes (Joshi *et al.*, 2007).

Genotypes were classified into resistant (R), moderately resistant (MR), moderately resistant to moderately susceptible (MR-MS), moderately susceptible (MS), moderately susceptible to susceptible (MS-S), susceptible (S), susceptible to highly susceptible (S-HS) and highly susceptible (HS) as per AUDPC values (Liatukas and Ruzgas, 2012). The grouping separates the 50 genotypes into 6 as susceptible, 14 as susceptible to highly susceptible and 29 as highly susceptible. The only moderately susceptible to susceptible is ENTRY 1. No genotype was found either moderately resistant (MR) or resistant (R) category.

The 50 genotypes were classified into five clusters as per  $D^2$  analysis. The distribution of genotypes into 5 different clusters on the basis of  $D^2$  analysis based on 15 characters indicated substantial genetic diversity. The maximum inter cluster  $D^2$  value was exhibited by the Cluster I and V (1243.168) closely followed by Cluster I and IV (1138.777). The maximum intra cluster  $D^2$  was exhibited by Cluster II (101.703) closely followed by Cluster I (101.444). From this intra and inter cluster  $D^2$  value analysis it indicated that the genotypes from the most divergent Clusters viz. I and V could be selected as parents

for hybridisation to get wide spectrum of variation. Maximum contribution to the genetic divergence was attributed by the character CTD, followed by HI, GER and GPS.

PCA was done by using all the fifteen characters that were used for D<sup>2</sup> analysis. Among the fifteen principal components (PCs) only five components (PCs) had showed Eigen value >1.00, which accounted for 69.03% of cumulative proportion of variance. Biplot curve showed, in PC 1 characters such as BY, PH, GPS, AUCIPC had high positive loadings and thus contributed positively while AUDPC and CTD had negative loadings which contributed negatively. In PC 2, GY and HI had high positive loadings while DF had negative loading. Also, strong positive correlation was found between GY, BY, GPS while negative association between GY and DF, CTD and PH. These findings led to the conclusion that these effective contributing traits in PC 1 and PC 2 had the significant role in diversification of genotypes and selection might be possible based on this trait for future breeding programmes. Hierarchical clustering of 50 genotypes of wheat based on Euclidean distance exhibited dissimilarity with clustering using Mahalanobis distance due to difference in weightage provided to correlated variables by both the methods.

The correlation analysis among the 15 characters showed that only eight of them were positively associated with grain yield. Those traits included GER, PH, AL, SL, GPS, AUCIPC, BY and HI. Among the physiological traits, AUCIPC was found positively correlated with PH, CTD, BY and GY. Similarly, CTD was positively correlated with AUCIPC and BY. This indicated that with higher AUCIPC value the biomass of the genotypes also increased as well as canopy temperature decreased due to high physiological efficiency. Similar result was found in wheat Khakwani *et al.*, 2012 and Abdul *et al.*, 2018.

AUDPC was found to be positively correlated with HI whereas negatively correlated with DF, PH, GPS, AUCIPC and BY. This indicated that higher disease severity was negatively associated with high chlorophyll index value. This might be due to loss of greenness during high disease infestation. Similar finding was found by Rosyara *et al.*, 2007, Rosyara *et al.*, 2010.



**Plate No 1: Germination per meter calculation.**



**Plate No 2: Measuring chlorophyll index using chlorophyll meter**



**Plate No 3: Infrared Thermometer to measure Canopy Temperature.**



**Plate No 4: 19 HTWYT nursery at 100% heading stage**



**Plate No 5: 19 HTWYT nursery prior maturity stage.**



**Plate No 6: Spot Blotch in lower leaves of wheat.**

*Chapter 6*  
***References***

## Chapter 6

### REFERENCE.

---

- Abdul Latief A. Al-Ghzawi , Yahya Bani Khalaf, Zakaria I. Al-Ajlouni, Nisreen A. AL-Quraan, Iyad Musallam and Nabeel Bani Hani. (2018). The Effect of Supplemental Irrigation on Canopy Temperature Depression, Chlorophyll Content, and Water Use Efficiency in Three Wheat (*Triticum aestivum* L. and *T. durum* Desf.) Varieties Grown in Dry Regions of Jordan. *Agriculture*, 8, 67.
- Acharya K, Duta A K and Pradhan P (2011). *Bipolaris sorotintiana* (Sacc) Shoem the most destructive wheat fungal pathogen in the warmer areas *Australian. Journal of Crop Scienc* **5**: 1064-71
- Ahmed, A. U., M. A. Sarker, J. A. Choudhury, N. Ara and M. M. Ali. (2002). Genetic divergence analysis in chickpea. *Bangladesh J. Agril. Res.* **27**(1): 47-50.
- Allard, R. W., (1960). Principles of Plant Breeding. *John Wiley and Sonc Inc.*, New York, USA.
- Amani I. Fischer R A and Reynolds M P (1996). Canopy temperature depression association with yield of irrigated spring wheat cultivars in a hot climate. *J. Agronomy and Crop Science* **176**: 1 19-129.
- Amey D C (1951) Inheritance of resistance to Spot blotch in barley Seedling. *Phytopatho* **41**: 691-698.
- Aryal L, Shrestha S M, Khatri G, Chhetri B and Bhandari D (2015) Chlorophyll Content measurement of drought tolerant wheat genotypes sown at normal and late conditions as an indicator of spot blotch resistance. *J. Inst. Agric. Anim. Sci* **33-34**: 65-72.
- Balota, M., Payne, W.A., Evett, S.R., Lazar, M.D. (2007). Canopy temperature depression sampling to assess grain yield variation and genotypic differentiation in winter wheat. *Crop Sci.*, **47**:1518-1529.
- Bergale, S., Billore, M., Holkar, A.S., Ruwali, K.N., Prasad,S.V. and Mridulla, B. (2001). Genetic variability, diversity and association of quantitative traits with grain yield in bread wheat (*Triticum aestivum* L.). *Madras Agric. J.*, **88**(7-9): 457-461.



- Bhushan B. Singh K, Kaur S and Nanda G S (2002) Inheritance and allelic relationship of leaf blight resistance genes in three bread wheat varieties in the adult plant stage. *Journal of Genetics and Plant Breeding* **56**: 69-76
- Blum, A. (1988). Plant breeding for stress environments. CRC Press, Boca Raton, FL.
- Braun H J, Atlin G and Payne T (2010) Multi-location testing a tool to identify plant response to global climatic change. *Climatic change and crop production* **1**:115-38
- Braun H J, Rajaram S and Ginkel M (1996) CIMMYT's approach to breeding for wide adaptation. In Adaptation in plant breeding *Euphytica* **92**(1-2):197-183
- Burton G W, Devane E W (1953) Estimating heritability in tall fescue (*Festuca arundinaceae*) from replicated clonal material. *Agron J* **45** : 478—481
- Chaurasia S, Joshi A K, Dhari R and Chand R (1999) Resistance to foliar blight of wheat: A search. *Genet. Resource Crop Evol* **46**: 469-475
- Dotlacil, L, Hermuth, Stehno, Z. And Maner, M., (2000), Diversity in European winter wheat landraces and obsolete cultivars. *Czech-J. Genet. And Pl. Breed.*, **36**(2): 29-36.
- Dubey R, Pathak H, Chakraborty B, Singh S, Gupta D K, Harit R C. (2020). Impact of terminal heat stress on wheat yield in India and options for adoption. *Agricultural System* 181 102826.
- Dubin H J, van Ginkel M V (1991) The status of wheat disease and disease research in the warm areas. In: Sauders DA (ed.), Proceeding of the Wheat for the Non-traditional, Warm Areas Conference, 1990, Foz do Iguacu, u, Brazil, Mexico, DF, Mexico, CIMMYT, 125-145.
- Dublin. H.J and Rajaram. S. (1996). Breeding disease-resistant wheats for tropical highlands and lowlands. *Annu. Rev. Phytopathol* **34**:503–26
- Dwivedi V.K and Singh, S.P. (2002). Genetic divergence in wheat (*Triticum aestivum* L.). *New Agriculturist*, **13**(1-2): 5-7.
- Fikre, G. Alamerew, S. And Tadesse, Z. (2015). Genetic Variability Studies in Bread Wheat (*Triticum aestivum* L) Genotypes at Kulumsa Agricultural Research Center, South East Ethiopia, *J. Biol Agric. And Healthcare*, **5**(7):2224-2229.



- Fischer R A, Rees D, Sayre K D and Lu Z M (1998) Wheat yield progress is associated with higher stomatal conductance and photosynthesis rate and cooler canopies. *Crop science* **38**: 1467-1475
- Gashaw A, Mohammed H and Singh H (2007) Genetic diversity in selected durum wheat genotypes of Ethiopian plasm. *African crop science journal* **15**(2):67-72
- Goel P, Swati S P, Srivastava K and Jaiswal J P (2006) Assessment of losses by spot blotch (*Bipolaris sorokiniana*) with reference to resistance in wheat in Tarai region of Uttaranchal. *Indian Phytopathology* **59**(1):36-40.
- Gowda D S, Singh G P and Singh A M (2011) Relationship between canopy temperature depression, membrane stability, relative water content and grain yield in bread wheat (*Triticum aestivum*) under heat-stress environments. *Ind. Jour. Of Agri.Sc* **81**(3): 197-202.
- Griffe F (1925) Correlated inheritance of botanical characters in barley and the manner of reaction to *Helminthosporium sativum*. *J. Agric. Res* **30**: 915-933.
- Guendouz A, Maamri K, Benidir M and Hafsi M. (2012). Canopy Temperature Efficiency as Indicators for Drought Tolerance in Durum Wheat (*Triticum Durum* Desf.) in Semi-Arid Conditions. *Journal of Agriculture and Sustainability*. **1**: 23-38
- Guendouza A and Maamari K (2012) Grain-filling Chlorophyll Content Relation with Grain Yield Component of Durum Wheat in Mediterranean Environment. *Afr. Crop Sci. J* **20**(1): 31-37.
- Gupta D K, Bhatia A, Kumar A, Das T K, Jain N, Tomar R, Malayan S K, Fagodia R K, Dubey R, Pathak H. (2016). Mitigation of greenhouse gas emission from rice-wheat system of the Indo-Gangetic plains: through tillage, irrigation and fertilizer management. *Agric. Ecosyst. Environ.* **230**, 1-9.
- Hailegiorgis, D, Mesfin, M, Genet, T. (2011). Genetic Divergence Analysis on some bread wheat genotypes grown in Ethiopia. *J. Central European Agric.*, **12**(2):344-352.
- IPCC, 2007. Summary for policymakers. In: Parry M, Canziani O, Palutikof J, Vander Linden P, Hanson C (Eds.), *Climate Change 2007: Impacts, Adaptation and Vulnerability. Contribution of Working Group II to the Forth Assessment Report of the*

- Intergovernmental Panel on Climate Change. Cambridge University Press, Cambridge, pp. 7-22.
- IPCC, 2014. Climate Change 2014. Fifth Assessment Synthesis Report (Longer Report) of Intergovernmental Panel on Climate Change. Cambridge University Press, Cambridge, United Kingdom and New York, USA.
- Jaiswal, J.P., Arva, M., Kumar, A., Swati and Rawat, R.S. (2010) Assessing Genetic Diversity for yield and quality traits in indigenous bread wheat germplasm. *Electronic J. Pl. Breed.*, **1**(4): 202-209.
- Jangid K K and Srivastava J P (2018) Canopy Temperature Depression and Stay Green: Major Components for identifying Terminal Heat Stress Resistant Genotypes of Wheat, *Int. J. Pure App. Biosci* **6**(3): 374-381.
- Jangid K K and Srivastava J P (2018) Canopy Temperature Depression and Stay Green: Major Components for Identifying Terminal Heat Stress Resistant Genotypes of Wheat, *Int. J. Pure App. Biosci* **6**(3): 374-381
- Johnson HW, Robinson HF, Comstock RE (1955). Estimates of genetic and environmental variability in soybean. *Agron. J.*, **47**: 314-318
- Jolliffe, I. T., (1986): Principal Component Analysis. *Springer-Verlag*, 271 pp
- Joshi A K, Kumar S, Chand R and Ortiz-Ferrara G (2004) Inheritance of resistance to spot blotch caused by *B. Sorokiniana* in spring wheat. *Plant Breeding* **123**: 213-9.
- Joshi A K and Chand R (2002) Variation and inheritance of leaf angle, and its association with spot blotch (*B. Sorokiniana*) severity in wheat (*Triticum aestivum*). *Euphytica* **124**: 283-90.
- Joshi A K, Ortiz-Ferrara G, Crossa J, Singh G, Sharma R C, Chand R and Parsad R (2007) Combining superior agronomic performance and terminal heat tolerance with resistance to spot blotch (*Bipolaris sorokiniana*) of wheat in the warm humid Gangetic Plains of South Asia. *Field crops research* **103**(1):53-61.
- Karimizadeh R, Mohammadi M, Ghaffaripour S, Karimpour F, Shefazadeh M K. (2011) Evaluation of physiological screening techniques for drought-resistant breeding of durum wheat genotypes in Iran. *African Journal of Biotechnology* Vol. **10**(56), pp. 12107-12117

- Khakwani, A.; Dennett, M.D.; Munir, M.; Abid, M. (2012) Growth and yield response of wheat varieties to water stress at booting and anthesis stages of development. *Pak. J. Bot.* **44**, 879–886.
- Kumar S, Archal S, Tyagi R K, Kumar J, Vk V, Jacob S R, Srinivasan K, Rashamani J, Parimalan R, Sivaswamy M and Tyagi S (2016) Evaluation of 19,460 wheat accessions conserved in the Indian national gene bank to identify new sources of resistance to rust and spot blotch diseases. *Plos one.* **11**(12):e0167702.
- Kumar, B., Lal, G. M., Ruchi and Upadhyay, A. (2009). Genetic Variability, Diversity and Association of Quantitative Traits with Grain Yield in Bread Wheat (*Triticum aestivum*). *Asian J. Agric. Sci*, **1**(1): 4-6.
- Leilah A.A and Al-Khateeb S.A. (2005). Statistical analysis of wheat yield under drought conditions. *Journal of Arid Environments* **61**:483–496
- Liatukas Z and Ruzagas V (2012) Spot blotch resistance in derivatives of European winter wheat. *Turkish Journal of Agriculture and Forestry.* **36**(3):341-351.
- Lush JL (1949). Animal breeding plants (2<sup>nd</sup> Ed.). IOWA State college press, Ames, IOWA.
- Mahalanobis, PC. 1936. On the generalised distance in statistics. *Proc Nat. Inst Sci, India*, **2**: 49-55
- Mehta Y R (1985) Breeding wheat for resistance to *Helminthosporium* spot blotch. In: Wheat for the more tropical environments. *A proceeding of Mexico, D.F. CIMMYT.* pp 135-140.
- Mimmack G M, Mason S J, Galpin J S. (2001). Choice Of Distance Matrices In Cluster Analysis: Defining Regions. *Journal of Climate*, **14**(12), 2790-2797.
- Moll, R. H., Sathuana, w. S. and Robinson, H. F. 1962. Heterosis and genetic diversity in variety crosses of maize. *Crop Sci*, **2**: 197-199.
- Mondal S, Singh R P, Crossa J, Huerta-Espino J, Sharma I, Chatrath R Kalappanavar I K (2013) Earliness in wheat: A key to adaptation under terminal and continual high temperature stress in South Asia. *Field crops research* **151**: 19-26.

- Murdock L, Call D and James J (2004) Comparison and Use of Chlorophyll Meters On Wheat (Reflectance vs. Transmittance/Absorbance). Cooperative extension service, University of Kentucky-College of agriculture, UK.
- Nair, K. R. And H. K. Mukherjee. (1960). Classification of natural and plantation teak (*Tectona grandis*) grown at different location of India and Burma with respect to its mechanical and physiological properties. *Sankhya.*, **22**: 11-20.
- Neupane R B, Sharma R C and Duveiller E (2007) Major gene controls of field resistance to spot blotch in wheat genotypes “Milan/Shanghai #7, and ‘Chirya 3’. *Plant Disease* **91**: 692-7.
- Niles M T, Luhell M, Brown M. (2015). How limiting factors drive agricultural adaptation to climate change. *Agric. Ecosyst. Environ.* **200**: 178-185.
- Nimbalkar, C.A, Navale, P.A. and Biradar, A.B. (2002). Generalized D2 and genetic diversity in wheat. *J. Maharashtra agric. Univ*, **27**(1): 43-45
- Panigrahy S, Ray S S, Manjunath K R, Pandey P S, Sharma S K, Sood A, ... Parihar J S. (2011). A spatial database of cropping system and its characteristics to aid climate change impact assessment studies. *J. Indian Soc. Remote Sensing* **39** (3), 355-364.
- Panse, V. G., & Sukhatme, P. V. (1989). *Statistical methods for agricultural workers*. New Delhi: Indian Council of Agricultural Research.
- Parry M L, Rosenzweig C, Iglesias, Livermore, Fisher A M G. (2004). Effect of climate change on global food production under SRES emission and socioeconomic scenario. *Glob. Environ. Chang.* **14**, 53-67.
- Pathak H, Ladha J K, Aggarwal P K, Peng S, Das S, Singh Y, Singh B, Kamra S K, Mishra B, Sastri A S R A S, Aggarwal H P. (2003). Trends of climate potential and on farm yield of rice and wheat in the Indo-Gangetic plains. *Field Crop Res*, **80** (3), 223-234.
- Rai A, Thapa A, Bhattacharya PM, Chowdhury A K and Dev RM (2016) Evaluation of physiological trait for spot blotch (*Bipolaris sorokiniana*) resistance in wheat genotype. (*Triticum aestivum*). *The bioscan* **11**(4): 2727-2731.
- Rajaram, S., van Ginkel, M. & Fischer, R.A. (1995) CIMMYT’s wheat breeding mega-environment (ME). In Proceedings of the 8<sup>th</sup> International Wheat Genetics Symposium, Jul. 19-24 1993. Beijing, China.

- Rao, C.R. (1952). *Advanced Statistical Methods in Biometrical Research*. John Wiley and Sons, New York.
- Ray J and Ahmed J J (2015) Canopy temperature effects on yield and grain growth of different wheat genotypes. *IOSR Journal of Agriculture and Veterinary Science* **8**: 48-55.
- Redhu, A.S., Solanki, Y.P.S., Sethi, S.K. and Singh, I. (1995). Genetic diversity in some Indian exotic wheat varieties. *Crop Improv.* **22**(2):214-217
- Reynolds M P, Singh R P and Ibrahim A (1998) Evaluating physiological traits to complement empirical selection for wheat in warm environments. *Euphytica* **100**: 84-95.
- Rosenzweig C, Elliott J, Deryng D, Ruane A C, Muller C, Arneth A, Boote K J, Folberth C, Glotter M, Khabarov N, Neumann K. (2014), assessing agricultural risk of climate change in the 21<sup>st</sup> century in a global gridded crop model inter-comparison. *Proc. Natl. Acad. Sci.* **111** (9), 3268-3273.
- Rosyara U R, Duveiller E, Pan K and Sharma R C (2007) Variation in chlorophyll content, anatomical traits and agronomic performance of wheat genotypes differing in spot blotch resistance under natural epiphytotic conditions. *Australasian Plant Pathology* **36**(3): 245-251.
- Rosyara U R, Subedi S, Duveiller E and Sharma R C (2010) The effect of spot blotch and heat stress on variation of canopy temperature depression, i chlorophyll fluorescence and chlorophyll content of hexaploid wheat genotypes. *Euphytica* **174**(3): 377-390.
- Saari EE (1998) Leaf blight disease and associated soil borne fungal pathogens of wheat in South and South East Asia. In: Duveiller E, Dubin HJ, Reeves J, McNab A (eds), *Proceeding of the Helminthosporium Blight of Wheat: Spot Blotch and Tan spot Workshop, 1997, El Bata'n, Mexico, Mexico, DF, Mexico, CIMMYT*, 37-51.
- Saari EE and Prescott JM (1975). A scale for appraising the foliar intensity of wheat disease. *Plant Disease Reporter* **59**:377-380.
- Salman, S., Khan, Sd,khan, d., Khan, R.U. and Khan, (2014). Genetic variability studies in bread wheat (*Triticum aestivum* L.) accessions. *Pak. J. Agric. Res.*, **27**(1): 214-230

- Samal, K. M. and N. Jagadeb. (1996). Genetic divergence among chickpea cultivars. *Indian J. Genet.* **56**(1):86-88.
- Samale M, Reynolds M P, Warburton M, Skovmand B, Trethowan R, Singh R P, Ortiz-Monasterio I and Crossa J (2002) Dimensions of diversity in modern spring bred wheat in developing countries from 1965. *Crop Science* **42**(6):1766-1779.
- Searle SR (1961) Phenotypic, genetic and environmental correlation. *Biometrics* **17**(3):474-480.
- Sharma R C and Duveiller E (2006) Spot Blotch Continues to Cause Substantial Grain Yield Reductions under Resource-limited Farming Conditions. *J. Phytopathology* **154**: 482-488.
- Sharma, P. K., Gupta, P. K. And Balyan, H.S. 1998. Genetic diversity in large collection wheat (*Triticum Sp.*) *Indian J. Genet.*, **58**:271-278.
- Singh K, Sharma S N, Sharma Y. (2011). Effect of high temperature on yield attributing traits in bread wheat. *Bangladesh J. Agric. Res.* **36** (3), 415-426.
- Singh, P., Singh, A. K. Sharma, M. And Salgotra, S. K. 2014. Genetic divergence study in improved bread wheat varieties (*Triticum aestivum*). *African J. Agric. Res.*, **9**(4): 507-512.
- Singh, R.K. and B.D. Chaudhary. (1977). Biometrical methods in quantitative genetic analysis. Kalyani Publishers, New Delhi. pp. 39-68.
- Steffenson B J and Fetch T G (1995-1996) Evaluation of barley for resistance to Spot blotch, *Biol. Cult. Tests control Plant Disease* **11**:79.
- Talebi R (2011) Evaluation of chlorophyll content and canopy temperature as indicators for drought tolerance in durum wheat (*Triticum durum* Desf.). *Australian Journal of Basic and Applied Sciences* **5**(11): 1457-1462.
- Teixeira E L, Fischer G, Van Veltuizen H, Walter C, Ewert F. (2013). Global hotspot of heat stress on agricultural crops due to climate change. *Agric. For. Meteorol.* **170**: 206-215.
- Van Ginkel M and Rajaram S (1998) Breeding for resistance to spot blotch in wheat. *Global perspective*. In: Duveiller E, Dubin HJ, Reeves J, McNab A (eds), *Helminthosporium Diseases of Wheat: Spot Blotch and Tan Spot*. I CIMMYT, Mexico, pp 162-170.

- Velazquez-Cruz C (1994) Genetica de la resistencia a *Bipolaris sorokiniana* en trigos harineros. Ph.D. Thesis.
- Verma, D N., Singh, B. N., Sihgh, G. Singh, M.K. and Setter, T.L. 2014. Genetic diversity analysis for yield and other agronomic traits in bread wheat under water logged sodic soil condition. *J. Wheat Res.*, **6**(1):51-58.
- Voidani, P., Meybodi, M. And Damania, A.B. (1993). Distribution and genetic diversity of primitive bread wheats in Iran. Biodiversity and Wheat Improvement, ICARDA, Aleppo, Syria, 409-415.
- Walia, D.P. and Garg, D.K., (1996), Evaluation of genetic divergence in wheat (*Triticum aestivum*) germplasm. *Indian J. Genet. And Pl. Breed.*, **56**(4):452-457.
- Wilcoxson R D, Rasmusson D C and Mives MR (1975) Development of barley resistant to spot blotch and genetics of resistance. *Plant Dis* **74**:201-210.
- Zaman M A, Siddique M N A, Mahbubur Rahaman M, Abida M Y and Islam M J (2014) Divergence analysis of drought tolerant genotypes of wheat (*Triticum astivum* L.). *Bangladesh J. Agril. Res.* **39**(3): 385-396

**ANNEXTURE**

**A. Table of Chlorophyll Index at four growth stages and Area Under Chlorophyll Index Progress Curve (AUCIPC)**

<b>GEN</b>	<b>REP</b>	<b>1<sup>ST</sup> CI</b>	<b>2<sup>ND</sup> CI</b>	<b>3<sup>RD</sup> CI</b>	<b>4<sup>TH</sup> CI</b>	<b>AUCIPC 1</b>	<b>AUCIPC 2</b>	<b>AUCIPC 3</b>	<b>MAUCIPC</b>
1	1	169	191.2	218	179	3421.9	3273.6	1588	2761.167
1	2	194.8	246.4	166.6	131.8	4191.4	3304	1193.6	2896.333
2	1	155.6	193.4	136.2	105.6	3315.5	2636.8	967.2	2306.5
2	2	186.6	171.6	181	104.2	3402.9	2820.8	1140.8	2454.833
3	1	171.2	166.8	154.6	94.8	3211	2571.2	997.6	2259.933
3	2	162.4	231.4	146.4	77.6	3741.1	3022.4	896	2553.167
4	1	180	225.2	154.6	110.8	3849.4	3038.4	1061.6	2649.8
4	2	177.2	164.8	115.6	88.6	3249	2243.2	816.8	2103
5	1	150.8	171.8	161.8	98	3064.7	2668.8	1039.2	2257.567
5	2	117.8	118	102.8	90.8	2240.1	1766.4	774.4	1593.633
6	1	207	189.6	147	89.6	3767.7	2692.8	946.4	2468.967
6	2	155.4	149	115.4	75	2891.8	2115.2	761.6	1922.867
7	1	185.4	148.4	165.2	109.6	3171.1	2508.8	1099.2	2259.7
7	2	138.4	177	119.2	96	2996.3	2369.6	860.8	2075.567
8	1	194	187.4	171	137.8	3623.3	2867.2	1235.2	2575.233
8	2	221.6	178.4	167	89.8	3800	2763.2	1027.2	2530.133
9	1	234.8	196.4	186.4	134	4096.4	3062.4	1281.6	2813.467
9	2	153	146.2	131.4	94	2842.4	2220.8	901.6	1988.267
10	1	218	174.2	142.4	100	3725.9	2532.8	969.6	2409.433
10	2	251.6	164	112.8	82.4	3948.2	2214.4	780.8	2314.467
11	1	213.6	187.6	139.8	86.2	3811.4	2619.2	904	2444.867
11	2	185.4	169.4	180.6	122.6	3370.6	2800	1212.8	2461.133
12	1	180.2	180.6	131.4	80.8	3427.6	2496	848.8	2257.467
12	2	177.2	162.8	188	109	3230	2806.4	1188	2408.133
13	1	198.8	159	138.2	98	3399.1	2377.6	944.8	2240.5
13	2	153.2	158.4	114.6	74.2	2960.2	2184	755.2	1966.467
14	1	202	180.2	163.8	123.4	3630.9	2752	1148.8	2510.567
14	2	183.6	160.2	144.6	130.2	3266.1	2438.4	1099.2	2267.9
15	1	214.4	165	172.2	111	3604.3	2697.6	1132.8	2478.233
15	2	161	135.4	131.4	87.8	2815.8	2134.4	876.8	1942.333
16	1	191.4	247	194.4	102.8	4164.8	3531.2	1188.8	2961.6



16	2	140.8	148.8	112.6	82.2	2751.2	2091.2	779.2	1873.867
17	1	260.4	155.4	150.2	91.8	3950.1	2444.8	968	2454.3
17	2	173.4	150	179.8	116.6	3072.3	2638.4	1185.6	2298.767
18	1	183.4	158.8	144.6	94.8	3250.9	2427.2	957.6	2211.9
18	2	145.2	132.2	148.4	115.6	2635.3	2244.8	1056	1978.7
19	1	224.4	205	148.2	84.8	4079.3	2825.6	932	2612.3
19	2	189	193.8	165.6	100.8	3636.6	2875.2	1065.6	2525.8
20	1	188.6	172	137	81.4	3425.7	2472	873.6	2257.1
20	2	176.8	154.2	160.4	104.8	3144.5	2516.8	1060.8	2240.7
21	1	214.8	151	182.4	119.2	3475.1	2667.2	1206.4	2449.567
21	2	214	162	141.6	113	3572	2428.8	1018.4	2339.733
22	1	153.6	170.4	162.4	94.8	3078	2662.4	1028.8	2256.4
22	2	140.2	122.4	112.8	109.4	2494.7	1881.6	888.8	1755.033
23	1	187.4	188.2	156.8	104.2	3568.2	2760	1044	2457.4
23	2	130.8	161	138.2	92.6	2772.1	2393.6	923.2	2029.633
24	1	182.8	164.2	120.6	96.8	3296.5	2278.4	869.6	2148.167
24	2	160	189.6	122.6	73.4	3321.2	2497.6	784	2200.933
25	1	153.6	148.4	124	95.6	2869	2179.2	878.4	1975.533
25	2	176.8	123	151.8	121.4	2848.1	2198.4	1092.8	2046.433
26	1	189.8	163.6	171	121	3357.3	2676.8	1168	2400.7
26	2	153.2	173.4	170	95	3102.7	2747.2	1060	2303.3
27	1	182.6	199.2	177.4	110.4	3627.1	3012.8	1151.2	2597.033
27	2	148.2	220.4	129.4	245.2	3501.7	2798.4	1498.4	2599.5
28	1	192.6	197	211.8	121.4	3701.2	3270.4	1332.8	2768.133
28	2	212.2	142.2	193.6	145.2	3366.8	2686.4	1355.2	2469.467
29	1	170.2	211.2	203.2	125	3623.3	3315.2	1312.8	2750.433
29	2	207	200.2	132.8	115.8	3868.4	2664	994.4	2508.933
30	1	170.4	169.4	156.2	108.8	3228.1	2604.8	1060	2297.633
30	2	167.2	129.2	130.4	107.2	2815.8	2076.8	950.4	1947.667
31	1	164.6	158.4	132.4	95.2	3068.5	2326.4	910.4	2101.767
31	2	139.2	138	127.8	82	2633.4	2126.4	839.2	1866.333
32	1	178.8	146	138	97.8	3085.6	2272	943.2	2100.267
32	2	231.6	190.4	158.8	98.8	4009	2793.6	1030.4	2611
33	1	163.6	173	187	86.4	3197.7	2880	1093.6	2390.433
33	2	155.2	118	135.6	103.6	2595.4	2028.8	956.8	1860.333
34	1	206.2	170.2	155	124.6	3575.8	2601.6	1118.4	2431.933
34	2	160.6	156.6	139.4	83.4	3013.4	2368	891.2	2090.867

35	1	198.8	190.6	162.8	87	3699.3	2827.2	999.2	2508.567
35	2	178.2	175.4	125.8	88.4	3359.2	2409.6	856.8	2208.533
36	1	198.4	136.6	185.8	96	3182.5	2579.2	1127.2	2296.3
36	2	200	157	203.4	148.2	3391.5	2883.2	1406.4	2560.367
37	1	144.8	128.2	121	87.2	2593.5	1993.6	832.8	1806.633
37	2	158.2	122.8	143	104.4	2669.5	2126.4	989.6	1928.5
38	1	139	174.2	131	80.6	2975.4	2441.6	846.4	2087.8
38	2	184.4	183.6	129.8	97	3496	2507.2	907.2	2303.467
39	1	133.8	155.6	158.4	112	2749.3	2512	1081.6	2114.3
39	2	204.4	176	199	131.6	3613.8	3000	1322.4	2645.4
40	1	177.8	149.6	126.6	85.8	3110.3	2209.6	849.6	2056.5
40	2	174.8	176.6	160.6	87.6	3338.3	2697.6	992.8	2342.9
41	1	152.4	163.4	153.4	109.6	3000.1	2534.4	1052	2195.5
41	2	156.8	140.6	124.8	125.4	2825.3	2123.2	1000.8	1983.1
42	1	162	135.8	109.8	98.6	2829.1	1964.8	833.6	1875.833
42	2	163.8	163.8	146.4	119.2	3112.2	2481.6	1062.4	2218.733
43	1	217.8	178.8	132.2	77	3767.7	2488	836.8	2364.167
43	2	223	162.2	165.4	85.6	3659.4	2620.8	1004	2428.067
44	1	163.8	135	153	91	2838.6	2304	976	2039.533
44	2	166.2	163	177.4	104.4	3127.4	2723.2	1127.2	2325.933
45	1	174.6	137.6	137.4	87	2965.9	2200	897.6	2021.167
45	2	174.4	156.2	136.2	105.8	3140.7	2339.2	968	2149.3
46	1	166.6	130	135.2	104.8	2817.7	2121.6	960	1966.433
46	2	155.2	170.6	191.6	87	3095.1	2897.6	1114.4	2369.033
47	1	161.8	145.8	122.8	101.4	2922.2	2148.8	896.8	1989.267
47	2	128.8	164.8	131	84.2	2789.2	2366.4	860.8	2005.47
48	1	185	168.8	111.6	85	3361.1	2243.2	786.4	2130.233
48	2	133.6	123.6	130.4	74.2	2443.4	2032	818.4	1764.6
49	1	206.4	172.8	120.6	101.2	3602.4	2347.2	887.2	2278.933
49	2	118.8	128.8	123.8	88.4	2352.2	2020.8	848.8	1740.6
50	1	182.2	158.2	125	77.8	3233.8	2265.6	811.2	2103.533
50	2	157.8	198.2	129	97.4	3382	2617.6	905.6	2301.733

**B. Table of Canopy Temperature Depression (CTD) at two growth stages.**

<b>GEN</b>	<b>REP</b>	<b>CTD1</b>	<b>CTD2</b>	<b>M CTD</b>
1	1	2.54	-1.32	0.61
1	2	1.18	0.88	1.03
2	1	0.98	0	0.49
2	2	2.52	-0.24	1.14
3	1	1.22	-0.46	0.38
3	2	1.06	1.66	1.36
4	1	1.74	0.46	1.1
4	2	-0.14	0.42	0.14
5	1	0.94	0.96	0.95
5	2	-0.24	1.08	0.42
6	1	1.28	-0.02	0.63
6	2	2.86	-0.24	1.31
7	1	-0.14	0.76	0.31
7	2	2.26	0.86	1.56
8	1	0.76	1.46	1.11
8	2	0.6	3.5	2.05
9	1	1.44	-1.42	0.01
9	2	2.72	1.68	2.2
10	1	2.32	-1.7	0.31
10	2	1.04	0.3	0.67
11	1	2.6	-0.7	0.95
11	2	1.5	1.9	1.7
12	1	0.84	-1.06	-0.11
12	2	0.44	1.34	0.89
13	1	1.8	0.12	0.96
13	2	1.86	0.6	1.23
14	1	2.3	1.16	1.73
14	2	0.04	1.58	0.81
15	1	2.18	-0.36	0.91
15	2	1.92	0.12	1.02
16	1	1.56	1.2	1.38
16	2	0.76	0.08	0.42

17	1	0.7	-0.2	0.25
17	2	0.62	-0.46	0.08
18	1	2	0.94	1.47
18	2	0.34	2.36	1.35
19	1	1.52	1.26	1.39
19	2	1.56	-0.8	0.38
20	1	1.64	0.6	1.12
20	2	-0.36	1.2	0.42
21	1	0.76	0.4	0.58
21	2	1.08	1.72	1.4
22	1	2.08	-0.84	0.62
22	2	1.34	-0.48	0.43
23	1	1.18	0.22	0.7
23	2	1.52	1.04	1.28
24	1	0.6	1.8	1.2
24	2	0.84	1.16	1
25	1	1.42	1.82	1.62
25	2	0.84	-0.16	0.34
26	1	3.26	3	3.13
26	2	1.38	0.68	1.03
27	1	2.78	2.04	2.41
27	2	2.12	1.02	1.57
28	1	2.14	2.68	2.41
28	2	0.96	1.14	1.05
29	1	2.18	0.04	1.11
29	2	1.34	0.62	0.98
30	1	2.84	-2.1E-15	1.42
30	2	-0.52	1.54	0.51
31	1	1.12	0.56	0.84
31	2	1.42	-0.22	0.6
32	1	0.44	-0.2	0.12
32	2	1.02	1.24	1.13
33	1	0.86	0.82	0.84
33	2	0.72	-0.7	0.01
34	1	0.78	1.18	0.98

34	2	1.1	1.96	1.53
35	1	1.4	0.04	0.72
35	2	0.68	3.36	2.02
36	1	1.54	1.16	1.35
36	2	1.36	0.74	1.05
37	1	1.54	-2.06	-0.26
37	2	0.14	1.42	0.78
38	1	0.5	-0.88	-0.19
38	2	2.7	1.48	2.09
39	1	-1.66	-0.86	-1.26
39	2	1.56	0.32	0.94
40	1	1.16	-0.7	0.23
40	2	0.9	0.86	0.88
41	1	0.3	-0.84	-0.27
41	2	1.36	-0.88	0.24
42	1	0.54	-2.62	-1.04
42	2	0.7	-0.12	0.29
43	1	1.66	0.28	0.97
43	2	0.14	-1.5	-0.68
44	1	0.1	0.3	0.2
44	2	1.06	0.84	0.95
45	1	0	0.96	0.48
45	2	0	-1.58	-0.79
46	1	-0.72	0.38	-0.17
46	2	0.96	0.94	0.95
47	1	1.22	0.22	0.72
47	2	1.46	1.74	1.6
48	1	0.28	-1.86	-0.79
48	2	0.06	2.94	1.5
49	1	-0.04	-0.16	-0.1
49	2	0.42	1.18	0.8
50	1	0.4	0.84	0.62
50	2	1.92	-0.08	0.92

**C. Table of Disease Severity % at four growth stages with Area Under Disease Progress Curve (AUDPC).**

<b>GEN</b>	<b>REP</b>	<b>DS% 1</b>	<b>DS% 2</b>	<b>DS% 3</b>	<b>DS% 4</b>	<b>AUDPC 1</b>	<b>AUDPC 2</b>	<b>AUDPC 3</b>	<b>M AUDPC</b>
1	1	17.40741	23.7037	48.2716	58.88889	143.8889	251.9136	375.0617	256.9547
1	2	22.09877	34.19753	52.83951	68.88889	197.037	304.6296	426.0494	309.2387
2	1	24.44444	32.22222	59.1358	95.55556	198.3333	319.7531	541.4198	353.1687
2	2	25.92593	53.7037	86.66667	98.88889	278.7037	491.2963	649.4444	473.1481
3	1	33.33333	30.74074	57.53086	81.11111	224.2593	308.9506	485.2469	339.4856
3	2	40.24691	51.35802	82.22222	95.55556	320.6173	467.5309	622.2222	470.1235
4	1	33.7037	40.98765	57.65432	97.77778	261.4198	345.2469	544.0123	383.5597
4	2	36.54321	43.95062	78.14815	93.33333	281.7284	427.3457	600.1852	436.4198
5	1	23.08642	37.65432	48.76543	97.77778	212.5926	302.4691	512.9012	342.6543
5	2	36.17284	52.22222	85.4321	94.44444	309.3827	481.7901	629.5679	473.5802
6	1	24.44444	33.95062	68.02469	93.33333	204.3827	356.9136	564.7531	375.3498
6	2	32.96296	50.37037	95.55556	100	291.6667	510.7407	684.4444	495.6173
7	1	37.65432	35.4321	54.69136	96.66667	255.8025	315.4321	529.7531	366.9959
7	2	31.85185	40.61728	70.37037	93.33333	253.642	388.4568	572.963	405.0206
8	1	14.07407	19.75309	57.90123	92.22222	118.3951	271.7901	525.4321	305.2058
8	2	25.30864	47.65432	96.66667	96.66667	255.3704	505.1235	676.6667	479.0535
9	1	18.14815	28.14815	51.11111	86.66667	162.037	277.4074	482.2222	307.2222
9	2	18.02469	29.62963	72.46914	90	166.7901	357.3457	568.642	364.2593
10	1	27.90123	42.83951	63.58025	90	247.5926	372.4691	537.5309	385.8642
10	2	27.65432	52.22222	79.38272	95.55556	279.5679	460.6173	612.284	450.823
11	1	23.08642	37.65432	64.44444	91.11111	212.5926	357.3457	544.4444	371.4609
11	2	15.55556	31.35802	78.2716	96.66667	164.1975	383.7037	612.284	386.7284
12	1	17.77778	24.19753	60	85.55556	146.9136	294.6914	509.4444	317.0165
12	2	21.60494	30.61728	57.53086	86.66667	182.7778	308.5185	504.6914	331.9959

13	1	19.75309	41.11111	64.44444	90	213.0247	369.4444	540.5556	374.3416
13	2	29.01235	41.60494	48.2716	98.88889	247.1605	314.5679	515.0617	358.93
14	1	40	45.92593	66.66667	93.33333	300.7407	394.0741	560	418.2716
14	2	28.39506	35.80247	50.74074	82.22222	224.6914	302.9012	465.3704	330.9877
15	1	22.96296	32.71605	75.55556	94.44444	194.8765	378.9506	595	389.6091
15	2	29.50617	30.74074	77.77778	96.66667	210.8642	379.8148	610.5556	400.4115
16	1	25.24005	51.23457	63.33333	85.55556	267.6612	400.9877	521.1111	396.5866
16	2	38.76543	46.66667	84.44444	98.88889	299.0123	458.8889	641.6667	466.5226
17	1	24.07407	43.08642	91.23457	88.88889	235.0617	470.1235	630.4321	445.2058
17	2	39.50617	54.69136	92.22222	98.88889	329.6914	514.1975	668.8889	504.2593
18	1	36.91358	47.16049	70	93.33333	294.2593	410.0617	571.6667	425.3292
18	2	32.83951	38.51852	56.41975	93.33333	249.7531	332.284	524.1358	368.7243
19	1	20	45.06173	92.09877	96.66667	227.716	480.0617	660.679	456.1523
19	2	44.19753	50	78.88889	96.66667	329.6914	451.1111	614.4444	465.0823
20	1	31.48148	47.28395	85.55556	95.55556	275.679	464.9383	633.8889	458.1687
20	2	25.18519	43.45679	84.44444	98.88889	240.2469	447.6543	641.6667	443.1893
21	1	21.85185	24.07407	64.32099	80	160.7407	309.3827	505.1235	325.0823
21	2	30.12346	24.07407	56.54321	74.44444	189.6914	282.1605	458.4568	310.1029
22	1	29.1358	52.96296	82.22222	95.55556	287.3457	473.1481	622.2222	460.9053
22	2	41.85185	55.92593	85.55556	98.88889	342.2222	495.1852	645.5556	494.321
23	1	17.77778	30.24691	54.32099	92.22222	168.0864	295.9877	512.9012	325.6584
23	2	34.69136	40.37037	88.88889	98.88889	262.716	452.4074	657.2222	457.4486
24	1	23.33333	16.04938	61.60494	84.44444	137.8395	271.7901	511.1728	306.9342
24	2	22.46914	23.45679	63.45679	93.33333	160.7407	304.1975	548.7654	337.9012
25	1	26.2963	20.98765	70.37037	78.88889	165.4938	319.7531	522.4074	335.8848
25	2	29.1358	37.40741	85.55556	98.88889	232.9012	430.3704	645.5556	436.2757

26	1	37.77778	38.76543	64.69136	88.88889	267.9012	362.0988	537.5309	389.177
26	2	39.50617	44.44444	90	97.77778	293.8272	470.5556	657.2222	473.8683
27	1	28.2716	38.39506	70	96.66667	233.3333	379.3827	583.3333	398.6831
27	2	25.55556	40.37037	73.33333	92.22222	230.7407	397.963	579.4444	402.716
28	1	23.58025	34.69136	66.66667	90	203.9506	354.7531	548.3333	369.0123
28	2	20.8642	29.38272	75.06173	86.66667	175.8642	365.5556	566.0494	369.1564
29	1	18.51852	21.60494	43.7037	88.88889	140.4321	228.5802	464.0741	277.6955
29	2	24.07407	34.69136	59.62963	67.77778	205.679	330.1235	445.9259	327.2428
30	1	30.24691	44.69136	88.88889	100	262.284	467.5309	661.1111	463.642
30	2	31.23457	53.08642	87.77778	98.88889	295.1235	493.0247	653.3333	480.4938
31	1	30.24691	56.04938	72.22222	95.55556	302.037	448.9506	587.2222	446.07
31	2	30.37037	57.53086	86.66667	97.77778	307.6543	504.6914	645.5556	485.9671
32	1	34.32099	45.80247	80	96.66667	280.4321	440.3086	618.3333	446.358
32	2	32.46914	53.20988	82.83951	93.33333	299.8765	476.1728	616.6049	464.2181
33	1	28.88889	51.85185	90	98.88889	282.5926	496.4815	661.1111	480.0617
33	2	22.22222	43.95062	56.66667	95.55556	231.6049	352.1605	532.7778	372.1811
34	1	34.69136	44.32099	68.88889	92.22222	276.5432	396.2346	563.8889	412.2222
34	2	32.22222	39.25926	93.33333	98.88889	250.1852	464.0741	672.7778	462.3457
35	1	37.40741	54.07407	93.33333	98.88889	320.1852	515.9259	672.7778	502.963
35	2	43.08642	60.49383	97.77778	100	362.5309	553.9506	692.2222	536.2346
36	1	21.11111	21.48148	74.93827	87.77778	149.0741	337.4691	569.5062	352.0165
36	2	21.7284	31.23457	76.2963	82.22222	185.3704	376.358	554.8148	372.1811
37	1	37.28395	50.24691	92.22222	100	306.358	498.642	672.7778	492.5926
37	2	36.17284	46.04938	90	97.77778	287.7778	476.1728	657.2222	473.7243
38	1	53.45679	47.16049	70	94.44444	352.1605	410.0617	575.5556	445.9259
38	2	35.06173	52.09877	93.33333	95.55556	305.0617	509.0123	661.1111	491.7284



39	1	20.74074	30.49383	58.64198	80	179.321	311.9753	485.2469	325.5144
39	2	19.1358	30.74074	54.32099	78.88889	174.5679	297.716	466.2346	312.8395
40	1	26.66667	51.85185	86.66667	96.66667	274.8148	484.8148	641.6667	467.0988
40	2	28.76543	30.61728	93.33333	100	207.8395	433.8272	676.6667	439.4444
41	1	35.92593	45.4321	77.40741	93.33333	284.7531	429.9383	597.5926	437.428
41	2	37.53086	40.74074	80	93.33333	273.9506	422.5926	606.6667	434.4033
42	1	45.30864	44.69136	89.25926	94.44444	315	468.8272	642.963	475.5967
42	2	30.37037	38.88889	71.23457	93.33333	242.4074	385.4321	575.9877	401.2757
43	1	28.02469	42.34568	97.77778	100	246.2963	490.4321	692.2222	476.3169
43	2	20	46.54321	100	100	232.9012	512.9012	700	481.9342
44	1	28.14815	36.41975	40.24691	71.11111	225.9877	268.3333	389.7531	294.6914
44	2	26.17284	47.53086	82.22222	97.77778	257.963	454.1358	630	447.3663
45	1	35.4321	43.58025	72.46914	90	276.5432	406.1728	568.642	417.1193
45	2	26.41975	31.11111	69.01235	95.55556	201.358	350.4321	575.9877	375.9259
46	1	29.25926	44.44444	76.66667	98.88889	257.963	423.8889	614.4444	432.0988
46	2	38.51852	47.90123	92.22222	97.77778	302.4691	490.4321	665	485.9671
47	1	22.46914	36.54321	61.48148	82.22222	206.5432	343.0864	502.963	350.8642
47	2	28.76543	32.22222	86.2963	97.77778	213.4568	414.8148	644.2593	424.177
48	1	27.40741	45.55556	67.77778	95.55556	255.3704	396.6667	571.6667	407.9012
48	2	40.37037	58.51852	74.44444	97.77778	346.1111	465.3704	602.7778	471.4198
49	1	24.07407	43.82716	86.04938	95.55556	237.6543	454.5679	635.6173	442.6132
49	2	46.79012	45.4321	91.48148	94.44444	322.7778	479.1975	650.7407	484.2387
50	1	23.45679	54.69136	91.60494	98.88889	273.5185	512.037	666.7284	484.0947
50	2	37.03704	63.58025	94.44444	100	352.1605	553.0864	680.5556	528.6008

## VITA

<b>Name of the student</b>	Sourik Poddar
<b>Father's Name</b>	Gopal Poddar
<b>Mother's Name</b>	Shampa Poddar
<b>Nationality</b>	Indian
<b>Date of Birth</b>	29.10.1997
<b>Permanent Home Address</b>	R. N. Road, Near Gauri Mahal, Coochbehar-736101, West Bengal.

### EDUCATION QUALIFICATIONS

#### Bachelor's degree:

<b>Degree</b>	B.Sc. Agriculture
<b>Name of university</b>	Hemvati Nandan Bahuguna Garhwal University
<b>Year of award</b>	2019
<b>percentage</b>	84.45%






#### Master's degree:

<b>OGPA</b>	
<b>Title of Master's Thesis</b>	Study of Genetic Diversity of Wheat ( <i>Triticum aestivum</i> L.) Genotypes Under Terai Agroclimatic Conditions
<b>Awards/Distinctions/Scholarships</b>	University Merit Scholarship

## Document Information

<b>Analyzed document</b>	ST COPY 4.08.2021.docx (D110981363)
<b>Submitted</b>	8/4/2021 11:33:00 AM
<b>Submitted by</b>	SAIKAT DAS
<b>Submitter email</b>	saikat.ubkv@gmail.com
<b>Similarity</b>	3%
<b>Analysis address</b>	saikat.ubkv.ubkv@analysis.urkund.com

## Sources included in the report

<b>W</b>	URL: <a href="https://www.researchgate.net/publication/318024610_Evaluation_of_Elite_Spring_Wheat_Triticum_aestivum_L_Genotypes_for_Yield_and_Yield_Attributing_Traits_under_Irrigated_Condition">https://www.researchgate.net/publication/318024610_Evaluation_of_Elite_Spring_Wheat_Triticum_aestivum_L_Genotypes_for_Yield_and_Yield_Attributing_Traits_under_Irrigated_Condition</a> Fetched: 3/6/2021 5:07:08 AM		1
<b>W</b>	URL: <a href="http://www.saulibrary.edu.bd/daatj/public/index.php/getDownload/07-02504_11.pdf">http://www.saulibrary.edu.bd/daatj/public/index.php/getDownload/07-02504_11.pdf</a> Fetched: 7/29/2021 12:15:03 AM		2
<b>W</b>	URL: <a href="https://www.researchgate.net/publication/45724731_Correlation_and_Path_Coefficient_Analysis_in_Bread_Wheat_under_Drought_Stress_and_Normal_Conditions">https://www.researchgate.net/publication/45724731_Correlation_and_Path_Coefficient_Analysis_in_Bread_Wheat_under_Drought_Stress_and_Normal_Conditions</a> Fetched: 8/6/2020 11:53:30 AM		1
<b>W</b>	URL: <a href="https://www.researchgate.net/publication/45724793_Path_Coefficient_Analysis_for_Water_Use_Efficiency_Evapo-transpiration_Efficiency_and_Some_Yield_Related_Traits_in_Wheat">https://www.researchgate.net/publication/45724793_Path_Coefficient_Analysis_for_Water_Use_Efficiency_Evapo-transpiration_Efficiency_and_Some_Yield_Related_Traits_in_Wheat</a> Fetched: 9/1/2020 1:08:14 PM		1
<b>W</b>	URL: <a href="https://www.researchgate.net/publication/340477886_Assessment_of_genetic_diversity_in_promising_bread_wheat_Triticum_aestivumL_genotypes">https://www.researchgate.net/publication/340477886_Assessment_of_genetic_diversity_in_promising_bread_wheat_Triticum_aestivumL_genotypes</a> Fetched: 12/24/2020 10:37:45 AM		3
<b>W</b>	URL: <a href="https://www.researchgate.net/publication/226585313_Variation_in_chlorophyll_content_anatomical_traits_and_agronomic_performance_of_wheat_genotypes_differing_in_spot_blotch_resistance_under_natural_epiphytotic_conditions">https://www.researchgate.net/publication/226585313_Variation_in_chlorophyll_content_anatomical_traits_and_agronomic_performance_of_wheat_genotypes_differing_in_spot_blotch_resistance_under_natural_epiphytotic_conditions</a> Fetched: 7/23/2020 5:10:51 PM		6
<b>W</b>	URL: <a href="https://www.researchgate.net/publication/237751711_Canopy_temperature_depression_as_an_indication_of_correlative_measure_of_spot_blotch_resistance_and_heat_stress_tolerance_in_spring_wheat">https://www.researchgate.net/publication/237751711_Canopy_temperature_depression_as_an_indication_of_correlative_measure_of_spot_blotch_resistance_and_heat_stress_tolerance_in_spring_wheat</a> Fetched: 12/7/2020 6:27:29 AM		1
<b>W</b>	URL: <a href="https://repository.ju.edu.et/bitstream/handle/123456789/2793/Tefera%20%20Abebe%20MSc%20thesis%20on%20rice.pdf?sequence=1">https://repository.ju.edu.et/bitstream/handle/123456789/2793/Tefera%20%20Abebe%20MSc%20thesis%20on%20rice.pdf?sequence=1</a> Fetched: 5/1/2021 2:06:10 PM		3
<b>W</b>	URL: <a href="https://www.ijcmas.com/7-7-2018/C.K.%20Chethana,%20et%20al.pdf">https://www.ijcmas.com/7-7-2018/C.K.%20Chethana,%20et%20al.pdf</a> Fetched: 7/10/2020 10:13:10 AM		1
<b>W</b>	URL: <a href="https://pdffox.com/thesis-master-of-science-in-agriculture-pdf-free.html">https://pdffox.com/thesis-master-of-science-in-agriculture-pdf-free.html</a> Fetched: 7/27/2021 12:29:20 PM		2

URL: [https://www.thepharmajournal.com/archives/2010/vol8issue2/PartK/R-2\\_06\\_633.pdf](https://www.thepharmajournal.com/archives/2010/vol8issue2/PartK/R-2_06_633.pdf)

1/37

(SOURIK PODDAR)

Signature of student

(DR. SAIKAT DAS)

Chairman

Advisory committee