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Estrogens, Neuroinflammation and Neurodegeneration

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1 **Abstract**

2

3 Inflammatory activation of microglia is a hallmark of several disorders of the CNS. In addition to protecting the
4 brain against inflammatory insults, microglia are neuroprotective and play a significant role in maintaining
5 neuronal connectivity; therefore, the prolongation and inflammatory status may limit the beneficial functions of
6 these immune cells. The findings that estrogen receptors are present in monocyte-derived cells and that
7 estrogens prevent and control the inflammatory response raise the question of the role that this sex hormone
8 plays in the manifestation and progression of pathologies that have a clear sex difference in prevalence, such as
9 multiple sclerosis, Parkinson's disease, and Alzheimer's disease. The present review aims to provide a critical
10 review of the current literature on the actions of estrogen in microglia and on the involvement of estrogen
11 receptors in the manifestation of selected neurological disorders. This current understanding highlights a
12 research area that should be expanded to identify appropriate replacement therapies to slow the progression of
13 such diseases.

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Introduction

The nervous system is not readily accessible to peripheral immune cells, but evolution has favored the selection of microglia as the resident immune cells in the central nervous system for the first line of protection against noxious stimuli, such as stress and pathogenic insults. To adapt to the needs of their environment, microglia are extremely plastic cells able to show an array of diversified phenotypes. Indeed, in response to a potential danger, microglia perform the following: *i.)* synthesize and release inflammatory molecules (e.g., TNFα, reactive oxidative species, inflammatory cytokines and chemokines); *ii.)* alert the brain and other immune cells, *iii.)* clear all debris in the parenchyma and *vi.)* provide nutrients to repair the damage induced in the cells surrounding the inflammatory battlefield; in addition, mounting evidence indicates that *v.)* microglia play a major supporting role in neurogenesis and neuronal activity.

In the case of major injury, microglia attract peripheral immune cells to form an integrative network (with astroglia, neutrophils, lymphocytes, plasma cells, and macrophages) that provides the brain with a strong defensive system. This functional complex is finely regulated by a well-timed synthesis of inflammatory and anti-inflammatory molecules for the transient inception of the inflammatory response in the presence of insults and to return to a surveying stage as the immune emergency is resolved. Failure of such homeostatic mechanisms may have severe pathological consequences, as an excessive, prolonged or asynchronous immune activation plays a very active role in the onset and progression of pathologies ranging from chronic pain and epilepsy to neurodegeneration and psychiatric disorders (1-4).

An emerging theme in the study of microglia function is the sex-related differences highlighted by a growing number of studies in male and female vertebrates. The precise roles played by genetic, hormonal or environmental cues in determining this sexual dimorphism remain to be clarified. Certainly, estrogens play a major role in controlling microglia activity. In this review, we will discuss recent advances in the understanding of microglial biology with a particular focus on the influence of estrogens on their function and on the physio-pathological relevance of this regulation. Furthermore, we will highlight the areas that need to be explored to

1 verify the potential for estrogen receptor ligands in the attenuation of neuroinflammation in specific neuronal
2 disorders.

3 **Microglia: the immune cells of the central nervous system**

4 **a. Microglia and brain development**

5 i. Microglia and structural organization of the developing brain

6 The existence of microglia was first described by Nissl in 1880. In the first decades of the twentieth century, the
7 seminal work of Santiago Ramon y Cajal and his student Pio Del Rio Hortega formed the basis for determining
8 the morphological and functional differences between microglia and other neural cells (5,6). Microglia, unique
9 among the major cell types in the CNS, are not derived from the ectoderm. In fact, during early fetal
10 development, a major wave of myeloid precursors migrate from the embryonic yolk sac to the brain to become
11 the resident microglia. Accordingly, genetic and cell lineage studies show that microglia originate from Pu.1
12 positive cells in both the mouse (7) and zebrafish (8), and fate-mapping experiments show colonization of the
13 brain by CSF1R^{+} erytro-myeloid progenitors at embryonic day 8.5 (9,10). The number of microglia precursors
14 that migrate to the brain around embryonic day 8.0 (10) is finite and relatively small (8) but sufficient to
15 proliferate, populate the entire brain and self-maintain for an entire life span. The factors required for brain
16 colonization have yet to be completely identified. In mice, colony stimulating factor 1 (CSF1) is involved
17 because mouse embryonic microglia express CSF1 receptor and *Csf1* gene deletion results in a significant loss of
18 microglia in adults (10,11). It is conceivable that macrophages migrate in response to an inflammatory stimulus,
19 as indicated by genetic studies conducted in zebrafish (12) and by the fact that in the developing brain, microglia
20 are generally large round amoeboid cells that produce elevated levels of cytokines and chemokines.

21 Microglia phagocytic activity contributes to the structural organization of the developing brain by eliminating
22 redundant neurons and synaptic connections (13) (Figure 1.). Microglia-dependent synaptic pruning is well-
23 documented in the developing hippocampus and thalamus, where in the presence of the complement system,
24 these cells engulf PSD-95-containing post-synaptic dendritic spines driven by the fractalkine system (14). The

phagocytic actions of microglia play a central role in the removal of apoptotic neurons (15), as well as inducing death in selected populations of viable neurons through a process called phagoptosis (16). The chemokine fractalkine (CX3CL1) released by the dying cells attracts phagocytic microglia, and the neurons to be engulfed are recognized by the phosphatidylserine (PS) exposed on the external surface. In addition, time-lapse microscopy in the brains of rodents and monkeys and in organotypic cortical slices showed that microglia phagocytose neural precursors in the cortex (17) and cerebellum (18). The *criteria* for the selection of the neurons to be eliminated during development require further investigation. However, the fact that microglia sense synaptic activity may suggest that the neurons eliminated are those not actively establishing synaptic contacts with their peers (19,20). Finally, microglia promote the survival of neurons and the growth of their axons (21) through the secretion of neurotrophic factors, such as IGF-1, IL-1β and IFNγ (22). Such a function is maintained in the adult brain, as discussed later.

ii. Microglia colonization of the brain is sex dependent

The colonization of the developing mouse brain by microglia appears to occur differently in the two sexes because males were shown to have overall more microglia early in postnatal development (P3-P4). This is in contrast with the fact that females have more microglia in selected brain areas later in development and in adulthood (23). The developing male hippocampus and cortex have a nearly 200-fold greater expression of the chemokine ligand CCL20 and 50-fold higher expression of the chemokine ligand CCL4 than those of females. Conceivably, these two chemokines play critical roles in driving the dimorphic perinatal colonization of brain regions relevant for cognition and memory, as well as a role in the highly sexual dimorphic POA region (23,24). The cause of the elevated levels of chemokines in the developing male brain remains unknown, but the temporal correlation between microglial brain colonization and the surge of testicular activity (at day E17) suggests the involvement of sex hormones (25) (Figure 2.). Additional investigations during brain maturation are necessary to learn whether sex hormone receptors have a sexual dimorphic expression. Microglia from P3 mice express ERα (26), and the mRNA content for this receptor subtype further increases in adult mice, suggesting that microglia sensitivity to estrogens increases with age (26). So far, no sexual differences were observed in ERα mRNA

content at any age (26,27). ERβ mRNA was detected in microglia primary cultures from P0 newborns (28), while the levels of this receptor are undetectable in microglia sorted *ex vivo* from mice at P3 and from adults (26,27). The data provided so far on the expression of the progesterone receptor (PR) and androgen receptor (AR) indicate that these receptors are not expressed in microglia in adult mice (27).

A sexually dimorphic behavior of microglia during development may be important because pre- or peri-natal infections may induce permanent neurological consequences. Indeed, excessive microglia activation during the developmental programming has been implicated in altered sexual behavior (29); dopamine-mediated functions and cognitive abilities (25,30); a predisposition to mental disorders, such as schizophrenia and autism (31); and in neurological alterations that have a different occurrence in males and females (31). This argues for the necessity of further investigation on the role played by the endocrine-microglia communication in the structural organization of the brain.

b. Microglia in the adult, healthy brain

In the adult nervous system, microglia are distributed throughout the brain with slight changes in concentrations and activity in each region (32,33). In the human brain, microglia account for up to 16% of all non-neuronal CNS cells and reside mainly in the white matter. In the rodent brain, microglia are found more often in the grey matter, and their content is lower (5-12% of all glial cells) (32,34). Observations in transgenic mice with a GFP reporter located under the control of the promoter for the fractalkine receptor gene (*Cx3cr1)* (35) demonstrated that microglia cells never rest and are constantly patrolling the brain parenchyma. Even in the absence of inflammatory stimuli, the processes extending from the cell somata are in continuous motion and growth, retracting and protruding in filopodia-like membranes with a bulbous ending (34). In this "surveillance" state aimed at detecting acute or chronic injuries, microglia movements are regulated internally by K^+/Cl^- co-transporters that mediate process swelling (36) and by de-polymerization and re-polymerization of actin filaments. These changes are induced by environmental cues, such as glutamate acting through microglia 2-amino-3-(5-methyl-3-oxo-1,2-oxazol-4-yl propanoic acid (AMPA) receptors (37), or purinergic molecules (38) and components of the complement system (5,39). Through this activity, microglia monitor three-dimensionally

1 the micro-environment (34) and fulfill their large number of housekeeping functions (e.g., removal of cellular
2 waste products and cell debris, remodeling of extracellular matrix, reshaping of synapses and neuronal
3 connectivity) (40). In addition, microglia secrete growth factors (transforming growth factor (TGF)β, fibroblast
4 growth factor (FGF), nerve growth factor (NGF) (41)) and lipoproteins, thus participating in stem cell
5 proliferation (42), neuronal dynamics and maintenance of neuronal membranes.

6 When in the surveying state, which is characterized by a highly ramified phenotype, healthy microglia are not
7 believed to secrete inflammatory molecules. Once challenged by physical or chemical insults (infection, trauma,
8 oxygen deprivation), microglia are "activated" and acquire an amoeboid, macrophage-like, morphology,
9 becoming phagocytic and able to secrete a large variety of inflammatory molecules. Activated microglia move
10 more rapidly and may cover relatively large distances in the brain parenchyma. Several other morphological
11 features have been described for microglia. For instance, in chronic disorders, microglia might acquire a rod-like
12 shape or become multinucleated and increase their dimensions in the presence of indigestible material. Finally,
13 microglia might present processes that are short and stocky. This latter morphology is often observed in aging
14 brains and is considered dystrophic (43).

15 All these phenotypes reflect a differential functional status with the expression of biochemical markers that are
16 very useful for a more objective definition of the microglia functional status (Table I). In their surveying stage,
17 these cells express low levels of myeloid-monocytic markers, such as Fc receptor-cluster of differentiation (CD)
18 32 and CD64, integrins (CD11b and CD11c), major histocompatibility complex (MHC) classes I and II and
19 CD45 (44). In the presence of specific stimuli, such as tissue damage, the so-called 'damage-associated
20 molecular pattern' (DAMPs) released by the injured cells induce microglia to transition to a pro-inflammatory
21 state. In this "classical activation", or M1 state, microglia retract their ramifications, potentiate phagocytic
22 activity, and increase the expression of cell surface proteins relevant for the innate immune response, such as
23 Toll-like receptors (TLR), inflammasome, phagocytic and scavenger receptors, and receptors for advanced
24 glycation of end products (RAGE) (45-54). The M1 state is also associated with the production of cytokines and
25 chemokines, in particular CCL2 (also named monocyte chemotactic protein-1, MCP-1), which is responsible for

the recruitment and migration of additional microglia to the insult site (55). In the M1 state, microglia also show increased expression of phagocytic oxidase (PHOX) (56) and inducible nitric oxide synthase (iNOS) (57), as well as the increased generation of nitric oxide, the main cytotoxic mediator in acute and chronic inflammatory responses (58) (Table I).

This host defense mechanism, which is very effective in taming inflammation, may cause local collateral damage. Thus, upon removal of the inflammatory stimuli, an elaborate and organized response is required to replace lost and damaged cells and to restructure the damaged extracellular matrix (ECM), with the final aim to restore tissue homeostasis. At this point, microglia change their phenotype and promote the blockade of the immune response and the commencement of specific programs aimed at repairing the damaged tissue (59). This activity is carried out in concert with glia and neurons and includes the synthesis and secretion of specific anti-inflammatory cytokines, which are responsible for the transition of microglia to other functional phenotypes (60) (Table I). In particular, IL-4 and IL-13 induce the "alternative activation" (or M2a phenotype) responsible for the resolution of the inflammatory phase by indirectly repressing the production of pro-inflammatory cytokines and the expression of iNOS (61,62). Ligation of immunoglobulin Fc gamma receptors (FcγRs) (CD16, CD32 or CD64) by immune complexes on LPS or IL-1β primed microglia results in the "type II alternative activation" (M2b phenotype), leading to downregulated expression of IL-12, increased IL-10 secretion and increased MHC-II expression (Table I). The dampening of the inflammatory response is also associated with the "acquired deactivation" phenotype (or M2c phenotype), characterized by the production of IL-10 and TGFβ, and a strong repression of MHC-II (Table I). These cytokines account for trophic effects and tissue remodeling functions, including remodeling of the ECM (63), angiogenesis (64), and, in neurotrophic niches, neurogenesis (65). The acquired deactivation stage can also be induced by the presence of apoptotic cells as microglia recognize the PS exposed on the surface of apoptotic neurons (66). Soluble bridging molecules, such as the adapter protein growth arrest-specific 6 (Gas6) (67), bind to PS through their GLA domain (N-terminal 11 γ-carboxyglutamic acid residue), thus serving as eat-me signals that are recognized by receptor tyrosine kinases (68) on the microglial membrane (Tyro3, Axl, and Mer - TAM). In chronic inflammatory diseases associated with aging, this trophic function appears to be impaired (69), possibly in relation to a decline in Gas6 expression. These different stages

of activation do not have well-defined boundaries, but they represent a continuum among each other, and similar to peripheral macrophages, they are classified on the basis of the genes the preferentially express (Table I).

i. Microglia - astrocyte interactions

The anatomical changes induced by injury and disease in astrocytes were described more than 100 years ago. We now know that reactive astrocytes protect neural cells and tissues by several means that include the following: *i.*) the modulation of synaptic activity through the uptake of potentially toxic molecules, such as glutamate (70), or the blockage of transporters for inhibitory peptides, such as γ-aminobutyric acid (GABA) (71); *ii.)* the release of glutathione and adenosine to control oxidative damage (72,73) and *iii.)* the degradation of protein aggregates in the brain parenchyma (74). Microglia activation and astrogliosis are commonly observed in the case of brain injury, infection and neurodegenerative diseases; however, we lack the necessary insight into the functions of the bidirectional cross-talk occurring between these two cell types. The seminal work by Gan and colleagues based on *in vivo* transcranial time-lapse, two-photon imaging demonstrated that after a small laser insult, microglia near the site of injury responded within a few minutes, and microglial processes converged to the site of injury driven by the ATP released from astrocytes (75), recognized by the $P2Y_{12}$ G-coupled receptors expressed by the surveying microglia (76). Prior *in vitro* studies led to the hypothesis that astroglia may attenuate microglia reactivity or facilitate the resolution of the inflammatory response. This would occur through the synthesis and release of GABA, which decreases microglial production of inflammatory cytokines (77) and microglia expression of antioxidant molecules (e.g., hemeoxygenase-1) regulated by the erythroid 2-related factor, Nrf2 (78). In turn, initial studies point to a microglia-mediated modulation of astrocytes through the release of purines (79) or other inflammatory molecules, such as prostaglandin D_2 (PgD_2), that are known to induce astrogliosis (80). The development of methodologies for the isolation and culture of pure populations of microglia or macroglia demonstrated that astrocytes are insensitive to inflammatory stimuli, and their ability to produce regulators of the proteolytic balance (tissue inhibitors of metalloproteases or TIMPs) in response to molecules, such as LPS is mediated by microglia (81). This function was shown to be necessary for the survival of neurons after ischemic insult and in demyelinating diseases (82,83). The finding of a reciprocal regulation between

microglia and astrocytes in the control of neuroinflammation demands additional studies to better understand the extent to which impairments of this two-way communication is associated with the onset of CNS disorders.

ii. Microglia - interactions with other immune cells

Another important function of microglia is the presentation of foreign antigens to T lymphocytes. In the healthy brain, antigen-presenting cells (APC) are represented by macrophages and dendritic cells in the meninges, choroid plexus and perivascular spaces. Activated microglia upregulate the expression of the molecules that are needed for an optimal APC function (84). It is still controversial whether monocytes contribute to the adult microglial population. The current belief is that although monocytes may penetrate the adult brain and differentiate into microglia, these cell are short-lived and an unlikely source for maintaining the microglia population in steady-state conditions. However, during certain neuroinflammatory pathologies (e.g., multiple sclerosis or Alzheimer's disease), the recruitment of circulating bone marrow progenitors can supplement, to some extent, the microglial population (85,86).

i. Microglia - neuron interactions

The intimate relationship between neurons and microglia in the adult brain is believed to be the recapitulation of what was already described for microglial development. In the mature nervous system, the major form of communication between neurons and microglia is the fractalkine receptor, CX3CR1, but other proteins may be involved (e.g., CD200 and receptors for neurotransmitters and neuropeptides). Microglia are the only brain cells that can express high levels of the CX3CR1, a G-protein coupled activated by the CX3CL1 ligand, a trans-membrane glycoprotein that may be released by neurons after proteolytic cleavage as a consequence of cytotoxic or other stimuli. The activation of the fractalkine receptor serves two main purposes: *i.)* modulation of synaptic pruning (14) and *ii.)* constraint of microglia activation.

Recent molecular imaging studies with two-photon laser microscopy showed that surveying microglia stop and regularly interact with all synaptic elements (the presynaptic terminal, perisynaptic process and synaptic cleft, but not the dendritic shaft); these contacts last for a highly variable period of time (20). It is important to note that the frequency of these contacts is relative to neuronal activity. For instance, in the case of ischemia induced

1 by transient occlusion of the middle cerebral artery, the durations of microglia-neuron contacts are significantly
2 prolonged (from minutes to an hour). During that period, several presynaptic terminals disappear, clearly
3 suggesting that microglia may control spine densities in relation to neuronal activity (14,20). The study of mice
4 deficient in complement C1q or C3 showed defects in the elimination of CNS synapses (87), supporting the
5 hypothesis of an involvement of the complement in microglia regulation of synaptic functions (Figure 1). In
6 addition to the CX3CR1, microglia are equipped with a *plethora* of receptors for neurotransmitters and
7 neuropeptides, such as GABA, glutamate and Substance P (88), enabling microglia to sense neuronal activity,
8 synthesize and secrete inflammatory mediators, neurotrophic factors or modulators of its own phagocytic activity
9 (89). When damaged, neurons may attract microglia by releasing neurotransmitters, such as glutamate (90) or
10 other signals such as growth factors (e.g., FGF-2) (91). Once in the vicinity of neurons, microglia receptors sense
11 neuronal activity and lead microglia to participate in plastic changes at the synapse through several mechanisms
12 (41,89). In the case of neuronal death, microglia are rapidly activated to clear the apoptotic cell debris, which
13 could be harmful for the bystander neurons (51,92).

14 Neurons, in turn, have the means to control the transition of microglia into the inflammatory phenotype and may
15 increase the threshold of microglial sensitivity and reactions to neurotoxic stimuli. For instance, in CX3CR1-
16 deficient mice (93,94), the state of microglia activation in response to stimuli, such as LPS or 1-methyl-4-
17 phenyl-1,2,3,6-tetrahydropyridine (MPTP), was much higher than that in wt animals. Depending on the
18 circumstances, neurons may facilitate the resolution of the inflammatory status and induce microglia to
19 synthesize trophic factors relevant for neuronal health. Thus, the neuron-microglia reciprocal modulation must
20 be kept under a tight balance as microglia may be relevant for a homeostatic neuronal signaling, by its excessive
21 activity may damage neurons. This was noted by several *in vitro* studies that emphasized the damage-
22 exacerbating effects of microglia-derived NO after prolonged stimulation with LPS (95-97), glucose stimulation
23 or ischemia (98,99). High NO levels inhibit neuronal respiration, causing the release of glutamate (96,97), and
24 NMDA receptor-mediated neurotoxicity is potentiated by the presence of activated microglia. This mechanism
25 may recapitulate physiological events necessary for brain development, which in the adult healthy brain are kept

under control because neurons may limit microglia negative influences by releasing substances able to induce apoptosis of activated microglia (100,101).

All together, these observations support the vision that microglia are a very significant complement to astrocytes in the regulation of neuronal synaptogenesis, transmission and survival (40). However, at the same time, they underline the necessity of a continuous control of microglia functions for the activity of neurons.

c. Energy metabolism and neuroinflammation

Diet-induced obesity is associated with neuroinflammation (102), heightened cytokine levels in several brain regions (103,104), hippocampal synaptic malfunctioning (105), altered neurogenesis (106) and cognitive impairment (103,105). In rodents, within the first week of consuming a high-fat diet, markers of neuronal injury were observed in the arcuate nucleus of the hypothalamus and in the adjacent median eminence that were associated with reactive gliosis involving both microglia and astroglia (107). This effect was reversible and was generated by saturated, but not unsaturated fatty acids (107), indicating the existence of a selective mechanism. However, a continued exposure to a high-fat diet (HFD) led to a permanent activation of microglia (but not astroglia) in the mediobasal hypothalamus that was observed also in humans by means of magnetic resonance imaging (108). Exercise reduces neuroinflammation, and this could be due to an improved glucose tolerance (109). However, considering that the effects of HFD were circumscribed to the brain regions responsible for the control of energy homeostasis and that the peptides αMSH and NPY induce cytokine and nitric oxide production by microglia, it is tempting to speculate that the activity of anorexigenic and orexigenic neurons in response to an unbalanced diet may be the trigger for microglia activation (78,95,110,111). A better understanding of this phenomenon is relevant for human health because obesity and metabolic syndrome are important risk factors for the development of Alzheimer's disease (112-115), and obese patients show deficits in learning, memory and executive functioning (116).

Most interestingly, in animal studies, young females were shown to be refractory to diet-induced neuroinflammation; ERα KO female mice fed a chronic HFD diet behaved similarly to wt males. This suggests

1 that estrogens and their receptors may regulate the neuroinflammatory response, and in females, circulating
2 estrogens may play a protective, anti-inflammatory role. What remains to be established are the mechanisms
3 through which estrogens may modulate microglial inflammatory responses: estrogens may directly regulate the
4 inflammatory genes, but may also regulate the production of compounds (such as NPY) triggering inflammation
5 in the hypothalamic peptidergic neurons, which are known to express ERα and to be susceptible to the actions of
6 estrogen (117,118).

7 **d. Microglia and aging**

8 Aging is the major risk factor for the development of neurodegenerative diseases (ND), and several large-scale
9 genetic studies have implicated microglial molecules in sporadic forms of ND, thus giving strength to the
10 hypotheses of a prominent role of neuroinflammation in the onset and progression of ND. The general consensus
11 is that, in the adult brain, microglia protect and defend other neural cells from pathological insults, but we know
12 very little about microglia efficiency in a senescent brain. Therefore, the compelling question is whether
13 senescent microglia are able to maintain their proliferative and brain patrolling capacity together with their
14 responsiveness to pathological insults? As previously mentioned, the microglia migrated from the embryonic
15 yolk sac proliferate and self-renew throughout the entire life span. This suggests that microglia live for long
16 periods and may somewhat undergo senescence by losing their efficiency and depriving the brain of its natural
17 defense (119,120). The finding that with aging there is an increased density of microglia in several brain regions
18 (121,122), suggests that the capacity to proliferate is maintained in time. This poses the question as to whether
19 such a life-long process leads to telomere shortening and the loss or gain of functions associated with the
20 replicative senescence. In fact, we still know very little about the existence of the stem-like microglia progenitors
21 proposed by Elmore et al. (11), even if the heterogeneous distribution of microglia in the aging brain might
22 support the view of subpopulations of cells throughout the parenchyma (122,123). The few studies addressing
23 the analysis of microglia morphology in aging brains show that aged human and rodent microglia are less
24 ramified with more tortuous processes carrying some bulbous swellings (43,124). The motility of microglial
25 processes appears to be diminished by age, as was indicated by *in vivo* imaging studies (125,126) and by

1 transcriptomic analysis, which found that young microglia express more motility genes than old microglia (127).
2 Immuno-phenotyping and biochemical studies established that aging is associated with a general upregulation of
3 markers that are typical of the pro-inflammatory state, and microglia are more readily responsive to toxic insults
4 (128-131). On the other hand, at least in rodents, markers of the microglia anti-inflammatory state were shown to
5 be increased with age (124,128). The studies conducted so far may at times appear contradictory because
6 microglia might respond very differently in relation to the pathological context in which they are studied.
7 Therefore, conclusive results could be drawn only from studies carried out in healthy aged brains (Figure 3.).
8 However, the ability to engulf and degrade the extracellular material resulting from the phagocytosis process
9 remains under-investigated in senescent microglia. Histological studies in the brains of aged, healthy, humans or
10 animals have shown accumulation of protein aggregates in the parenchyma. However, it is unclear whether this
11 is the consequence of an age-dependent abnormal production of aberrant proteins or their lack of clearance due
12 to a decreased microglia phagocytosis. It is likely that both mechanisms are occurring, as it is conceivable that
13 aging correlates with a decreased ability of microglia to proteolytically digest the engulfed protein aggregates
14 and debris from the surrounding space with a consequent impairment of the phagocytotic process (132). In
15 addition to brain local events triggering microglia activity, the generalized, systemic, inflammation that
16 accompanies aging in mammals may provide a significant contribution to inducing a pro-inflammatory,
17 dysfunctional state of these cells that is also facilitated by the increased permeability of the brain–blood barrier
18 described in aged organisms (84,133-135).

19 ## Mechanisms of estrogen actions in microglia

20 ### a. Estrogens may modulate target cell activity by interacting with several receptors

21

22 #### *i.* Intracellular receptors ERα and ERβ - structure

23 In mammals, two isoforms of the ER have been described. They are referred to as α and β, with each encoded by
24 a separate gene (*ESR1* and *ESR2*, respectively). The structure of the two receptors is very similar; however, the
25 functions of the two may differ considerably in different cell systems (136). (Figure 4.)

ii. Intracellular receptors ERα and ERβ - functions

Inhibitory proteins: In the absence of the cognate ligand, ERs are in a complex with proteins that prevent the receptor binding to the DNA (heat shock proteins, Hsp90, Hsp70 and other chaperons). The complex is mainly, but not exclusively, localized in the cell nucleus (137,138). Upon binding the cognate ligands, these receptors undergo conformational changes that lead to the release of inhibitory proteins, thus unmasking the DBD.

Post-translational modifications: ER activity, prior to and after DNA binding, is regulated by a constellation of post-translational modifications (PTM). These modifications include the following: phosphorylation, acetylation, methylation, sumoylation, and palmitoylation (139). This large variety of PTM regulates the half-life of the receptor proteins, as well as their cellular localization (140), and their ability to interact with DNA and other signaling proteins (141). Thus, PTM are necessary to tune the receptor functions in relation to cues present in the host cell and in the whole organism. Moreover, initial studies have demonstrated that the state of PTM of ERs is highly plastic and significantly regulated by the hormonal *milieu*. To provide an example of the multiple consequences of PTM, ERα phosphorylation (which may occur at 10 different serine/threonine/tyrosine residues) is necessary for the receptor dimerization and the recruitment of specific transcription factors, such as p160 co-regulators with chromatin remodeling enzymes (142).

ER activation in the absence of natural or synthetic ligands: Ligands are generally required to activate sex hormone receptor transcriptional activities. However, it is now well established that these receptors may also be activated in the absence of a ligand (unliganded activation). This phenomenon, initially proposed by O'Malley's group for the progesterone receptor (143,144), was then supported by a large series of observations in other nuclear receptors including ERs. It is now well accepted that unliganded ERs can be activated by growth factors (such as epidermal and the insulin-like growth factors) (145-147) through the involvement of selected kinases (MAPK, PKA and p21 ras/ERK) (148-151). A series of biochemical and genetic studies suggested the existence of cell-specific phosphorylation sites required for unliganded receptor activation (*e.g.,* Ser 118 in COS-1 cells and Tyr 537 in neuroblastoma cells) (152).

Protein-protein interactions: ER PTM involves their interaction with other proteins, such as calmodulin, cyclin D1, BRCA-1 and transcription factors (*e.g., c-fos, c-jun)* that may modify receptor functions (153,154). The interaction with co-regulators is essential for the modulation of target gene transcription because it enables the recruitment of general transcription factors to the TATA box and histone modification to facilitate RNA pol II transcription of the target genes (155,156).

ER intracellular signaling: Once activated by ligands or by PTM, ERs regulate the activity of their target cells by several mechanisms that include the following: a.) dimerization that enables recognition and binding of EREs in the promoter of target genes and interaction with co-activator and co-repressors to promote/repress transcription; b.) binding to other nuclear transcription factors (e.g., AP-1, NF-κB) interfering with their transcriptional capacity; c.) binding to cytoplasmic molecules involved in signal transduction (e.g., Src, PI3K, STATs) and alterations of their signaling (157).

iii. Intracellular ERα and ERβ may translocate to the cell membrane to regulate specific cell functions

Monomers of the classical intracellular ERα and ERβ may be induced to migrate to the cell membrane and associate with caveolae (158) by serine palmitoylation (C451/447 for mouse/human ERα) (140). In the cell membrane, estrogen binding induces dimerization of the ER and rapid signaling through Gα and Gβγ proteins (159). The association with G proteins was shown to occur in a cell- and context-specific mode to provide the appropriate cell response to various stimuli. A rapid process of depalmitoylation regulates the length and extent of this signaling (160). The number of intracellular receptors that are palmitoylated and transported to the membrane is a fraction of the total (approximately 5-6% of all ERs), yet it is sufficient to have a significant impact on glucose and lipid metabolism in different cell types (161,162). The recent generation of a mouse with a mutation at aa 451 (C451A) finally demonstrated that this receptor localization is essential at least for ovarian functions (163). Nevertheless, little is known so far about its functions in the CNS.

iv. The pharmacology of intracellular ERα and ERβ

In the last 50 years, a number of synthetic ER ligands were generated and developed for clinical use. These ligands include the following: clomiphene, tamoxifen, toremifene, raloxifene, bazedoxifene and ospemifene. The

common characteristic of these ER modulators is that they were selected to circumvent the use of natural estrogens due to their potential effects on endometrial and breast cancers. These compounds were named selective ER modulators (SERMs) because they bind the ER, but their agonist-antagonist effect is tissue dependent. Thus, most of the SERMs have antagonistic actions on ER in reproductive tissues. The first and second generation SERMs, tamoxifen and raloxifene, are used to treat ER-positive breast cancer and postmenopausal osteoporosis, respectively. The third-generation SERM, bazedoxifene (BZA), effectively prevents osteoporosis while blocking estrogenic stimulation in breast and uterine tissues. Unfortunately, the specific estrogenic vs. anti-estrogenic effects of SERMs on neuronal, glial or microglial cells have not been fully determined. The ability of these ligands to exert a tissue-specific action is attributed to the fact that by binding the ligand pocket of the ER, they induce conformational changes that are quite different than those of the E2-ER complex, this possibly limits their ability to interact like natural estrogens with the co-regulators (164). Selective ER down-regulators (SERDs, or pure antiestrogens), an alternative to SERMs, are characterized by a different activity because they cause downregulation and degradation of ERs. The prototype of a SERD is fulvestrant (ICI 182,780). More recently, the identification of microRNAs that can directly target ERs raised major interest particularly in the cancer field. The mechanisms involved in the miRNA-dependent modulation of ERs varies among the miRNAs isolated; most regulate the content of ERα indirectly, others, such as miR221-222, were shown to target ERα 3'-UTR and decrease ERα protein, but not mRNAs (165). A major advance in the field of synthetic ER ligands occurred recently with the identification of compounds that can discriminate the two ER isoforms and selectively bind to ERα (as its agonist PPT (1,3,5-tris (4-hydroxyphenyl)-4-propyl-1H-pyrazole) and antagonist side-chain pyrazoles) (166) or ERβ, e.g., the selective agonist DPN (2,3-bis (4-hydroxyphenyl) propionitrile). Unfortunately, the use of SERMs to target neural cells for clinical application is quite limited to date, due to still poor knowledge of the precise molecular targets of these molecules in the CNS, and the lack of pharmacokinetic data reporting the levels of permeability across the BBB (167).

v. Membrane receptors - GPR30

With a structure completely different from the intracellular receptors previously described, another molecule, an orphan 7-transmembrane receptor, GPR30, was found to be able to recognize and bind estrogens. GPR30 is a G-

protein-coupled receptor that can bind 17β-estradiol in the nanomolar range, thus with an affinity for the hormone with an approximate ten-fold lower than the intracellular receptors (168). GPR30 is present in the cell membrane and in the endoplasmic reticulum. In different model systems, the activation of GPR30 is mediated by 17β-estradiol and has been associated with several functions, including Ca^{++} mobilization (169-171), cAMP production (168), activation of protein kinases (171,172), activation of specific ion channels (173) and modulation of gene expression (174). Yet, the mechanisms underlying the intracellular activities of GPR30 are still unclear. A recent study showed that upon activation, the GPR30 forms hetero-oligomer complexes with Ca^{++}-ATPase and inhibits its activity through tyrosine phosphorylation of the pump (175). It is likely that the receptor may interact with several intracellular signal transducers through mechanisms that may change depending on the cell type. GPR30 is expressed in macrophages and in microglia (both primary cultures from neonatal rat brain and BV-2 cells), where it was shown to inhibit the production of cytokines and oxidative stress-related genes following stimulation by LPS (176) or hypoxia (177). GPR30 pharmacology has been the subject of several studies. In breast cancer cells, GPR30 is activated by the ER antagonist ICI 182,780 (178), Tamoxifen (179), and selective ligands for GPR30 have been identified and tested in several systems. The most investigated synthetic ligands for GPR30 are currently two steroids known as G-1 (agonist) (180) and G-15 (antagonist) (181,182); both of these ligands can readily cross the BBB.

b. Which ERs are expressed in microglia of the adult, mature brain?

The presence of ER in microglia was reported by several authors, and a large number of reports highlighted the anti-inflammatory action of estrogens in microglia cells, cultures and living animals. However, the abundance and type of ER expressed in microglia remains an object of discussion because of the large discrepancies in reports from different laboratories. Three major elements contribute to the inconsistencies in the literature:

Source of microglia and culture conditions: Microglia studies are carried out in retroviral immortalized cells; primary cell cultures from the neonatal or adult brain; pluripotent stem cells differentiated *in vitro*; and cells directly dissociated from embryonic, neonatal or mature brains with a variety of methodologies. Functional (183) as well as more recent genome-wide transcriptomic studies (184) demonstrated substantial changes in the

activity and transcriptome of FACS-sorted or cultured microglia and in microglia in the different stages of activation. This is not surprising in view of the plasticity of these cells and suggests that the expression of ERs is likely to change depending on the model system utilized. Indeed, studies carried out in microglia cell lines showed that ERα and ERβ mRNA content changes significantly with the number of passages (185). Considering that the growth factors present in the serum and the estrogenic activity of phenol red may activate ERs, it is conceivable that the culture media may represent a further element affecting the expression of these receptors.

Microglia sex: Generally, when primary cultures are established, both sexes are utilized, and even for the available cell lines, the sex of origin (N9 and M4T.4 are male and BV-2 and C8-B4 are female) (185) is not taken into consideration.

Current technology for the quantitative analysis of ER gene expression: Our ability to verify the expression of ERs by immuno-histochemical methodologies has been hampered by the minute dimensions of these cells, the low concentration of these receptors and the lack of a reliable and constant source of high-affinity antibodies.

The analysis of the literature on whole genome direct sequencing of microglia from adult brains clearly shows the presence of ERα, but not ERβ mRNA (Table II). The relative concentration of this mRNA is comparable to that for the mineralocorticoid receptor (MR) and is considerably lower than that for the mRNAs encoding the glucocorticoid receptor (GR), while no AR was found (Table II). These data appear to be consistent with regard to the relative abundance of the different receptors possibly because all studies utilized microglia dissociated from adult brains. Unfortunately, at present no data are available from the neonatal brain and primary cultures. In keeping with these findings are the results of the studies by Sierra, who first published a systematic analysis on the presence of selected steroid receptors in microglia isolated from the adult brain by cell sorting. Sierra reported the presence of ERα, GR and MR (27), but no detectable expression of ERβ, AR and PR. Staining with ERα antibodies revealed that this receptor is expressed mostly in the cytoplasm near the nucleus and in the cell processes (27,186). Interestingly, not all microglia cells were stained, and the use of electron microscopy enabled visualization of labeling for ERα in microglia processes in close apposition to neuronal dendritic spines. Immuno-detection of ERs in neonatal cultures of microglia showed both ERα and ERβ (138). The finding that ERα is low at P3 and increases rapidly to reach adult levels at day P21 (26) suggested that the expression of

1 these receptors may change in response to environmental cues. After treatment with LPS, ERα (together with
2 GR and MR) is downregulated, providing evidence that the functional status of microglia may influence the
3 expression of these genes, which in adult animals does not appear to be influenced by the hormonal status or sex
4 (27,187). The high reproducibility of these results in different laboratories sheds light from earlier reports on the
5 presence of ERβ or AR expression in microglia from healthy adult brains (188,189). Nevertheless, ERβ may be
6 expressed in microglia isolated from the spinal cord (190). Transformed cell lines (e.g., the murine BV-2 and N9
7 cells) express ERβ, and ERα mRNA is not present (BV-2) (191) or present at low levels (N9) (28,192). Direct
8 RNA sequencing of BV-2 transcriptome failed to detect mRNAs for all ERs (193); however, in this latter report,
9 the low expression of GR suggests that the sensitivity of the assay used was not optimal when compared to other
10 whole-genome RNA sequences.
11 With regard to GPR30, the results of several whole-genome RNA sequencing data (194,195) point to its
12 expression in the adult brain microglia, and immunostaining experiments carried out in rat microglia from the
13 neonatal brain showed its presence (176).
14

15 **c. Estrogen activity in microglia**

16 We are at an early stage in our understanding of estrogen influences on microglia activity, and the limited
17 number of studies available have concentrated on the anti-inflammatory actions of estrogen. Very little is known
18 with regard to the role of this hormone in the microglia functions reported above. For instance, considering the
19 relevance of estrogens in shaping neuronal circuitries during the sexual differentiation of the CNS, it is
20 surprising that the no investigator addressed the study of the role of estrogens in microglia-dependent synaptic
21 pruning. Similarly, very little attention has been given to the effects of these hormones on microglia trophic and
22 repair abilities despite the well-known neuronal protective actions of estrogen. The following paragraphs will
23 review the current literature on the anti-inflammatory effects that estrogen have in microglia.
24

25 i. Estrogen blockade of microglia activation after acute stimulation with inflammatory stimuli

There is a general consensus on the ability of estrogens to limit the microglia pro-inflammatory status after short exposure to bacterial lysates (196), viruses (197), unmethylated CpG oligonucleotides (198), or hypoxia (199,200). The hypothesis of the anti-inflammatory potential of estrogen was based initially on the *in vitro* observation that 17β-estradiol prevented the morphological changes induced by LPS and the concomitant synthesis of pro-inflammatory molecules (such as MMP9, prostaglandin E2, iNOS with ROS production) (196). These findings were subsequently reinforced by investigations on the anti-inflammatory potential of the synthetic ligands of ERs, such as tamoxifen and raloxifene, and natural estrogens, such as genistein, daidzein, and kaempferol, that were shown to attenuate the beta-amyloid peptide or LPS-induced microglia pro-inflammatory phenotype by inhibiting the synthesis of TNFα, IL-1β, MCP-1 or MIP2 in a dose-dependent manner (138) or the production of other inflammatory molecules, such as nitric oxide and ROS. These effects were blocked by prior treatment with ICI 182,780 (201-203). Most of these studies were performed in BV-2 cells or primary cultures of microglia either alone or mixed with astrocytes. Which of the two intracellular isoforms of ER is responsible for the anti-inflammatory properties of estradiol remains controversial, and both ERα and ERβ may trigger anti-inflammatory responses in the presence of high concentrations of ligand. The use of isoform-specific modulators, such as PPT or DPN, showed that PPT-activated ERα was more effective than DPN-induced ERβ for the inhibition of microglial production of IL-1α, IL-1β, TNF-α, and COX-2. This suggested that ERα plays a more significant role than ERβ in diminishing the inflammatory response of microglia. However, an increase in ERβ expression following treatment with 17β-estradiol or DPN in rat primary microglia provided greater attenuation of NO production, thus suggesting ERβ also plays a role (204).

ii. Estrogens and neuroinflammation – in vivo experiments

In vivo studies provided further, strong evidence on the capacity of estrogens to inhibit the neuroinflammatory processes. Ovariectomy (ovx) in rodents is clearly associated with an increased number of microglia with a pro-inflammatory morphology and up-regulation of a large number of markers of microglia reactivity (including the receptors for the recognition of inflammatory stimuli and for phagocytosis) (131,205). The fact that the administration of estradiol prior to ovx blocked microglia activation suggested that the neuroinflammatory reaction ensuing the surgery was mainly due to the lack of this hormone (131,205,206). Augmented expression

of inflammatory markers was also observed in women in post-menopause, particularly in areas of the brain functionally related to the regions shown to be most responsive to inflammatory stimuli in rodents.

The extent of microglia activation induced by ovx is exacerbated by aging. In fact, in ovx old mice, the expression of inflammatory mediators is much stronger than in intact mice of the same age. This indicates that low circulating estrogens increase the susceptibility of senescent microglia to inflammation (207). Most interestingly, studies with a mouse reporting on the transcriptional activity of ERs (ERE-Luc mouse (208) showed that the receptor transcriptional activity in the hippocampus diminished significantly with aging, in spite of the fact that the synthesis of ERα mRNA was increased and circulating levels of estrogens remained quite high (Figure 5). It is therefore plausible that with age a form of estrogen resistance is involved in the impaired ability of microglia to resolve inflammation, possibly leading to an ever increasing neuroinflammatory phenotype. Age is correlated with increased inflammation in males as well. Additional studies are necessary to understand the state of ER activity and local estrogen production to evaluate the contribution of these hormones to the activity of senescent microglia in males. The *in vivo* local or systemic administration of the endotoxin LPS is a commonly used model system for triggering a robust but transient inflammatory reaction in the rodent brain and is a valid tool to evaluate the effect of circulating estrogens in the prevention of acute microglia inflammatory activation. Intra-ventricular or intra-parenchymal injection of LPS demonstrated that the estrogen anti-inflammatory actions occurred in all brain areas studied and required the presence of ERα (209). The systemic administration of LPS enabled the authors to demonstrate that a peripheral inflammation may trigger a response also in the central nervous system with a rapid activation of microglia and expression of inflammatory cytokines. The same experimental setting performed in intact and in ERα or ERβ ko mice indicated that ERα was more effective than the ERβ isoform in dampening the local production of inflammatory molecules; ERβ was required for the suppression of BBB permeability (210). The lower number of immunoreactive microglia cells in mice treated with estrogens and LPS compared with mice treated with LPS alone was observed in both males and female mice (186). Similar to estrogens, the acute administration of tamoxifen and raloxifene to ovx, young and aged mice reduced microglia activation following LPS stimulation (186,198) or brain and spinal cord injury (211-213) pointed to the fact that these SERMs have an agonist activity on microglia ER. Long-term

1 treatment with 17β-estradiol or raloxifene in old ovx females significantly decreased the number of microglia
2 cells in the hippocampus (207) compared to placebo, suggesting that estrogens and SERMs may be considered
3 as protective treatments against age- and disease-related pathologies.

4

5 ### iii. Cellular mechanisms of ER-dependent anti-inflammatory activity

6 A detailed knowledge of how mammalian innate immunity is regulated has developed over the past 15 years.
7 Membrane and endosomal Toll-like receptors (TLRs) activated by a series of small molecules also derived from
8 parasites, bacteria, fungi, and viruses may dimerize to initiate a cytoplasmic response leading to the activation of
9 the transcription factors responsible for the induction of pro-inflammatory cytokines, and, in the case of
10 endosomal TLRs, the induction of type I interferon (IFN). Two important families of transcription factors
11 activated downstream of TLR signaling are the nuclear factor-κB (NF-κB) and interferon-regulatory factors
12 (IRFs). Other transcription factors, such as cyclic AMP-responsive element-binding protein (CREB) and
13 activator protein 1 (AP1), are also important (214). Currently, the understanding of the molecular mechanisms of
14 estrogen anti-inflammatory actions is incomplete because of the multiplicity of responses elicited by estrogens in
15 the neural, glial and immune cells in the brain and the variability in the microglial experimental systems used (as
16 explained above). Nevertheless, several lines of evidence indicate that estrogens and ERs control TLR signaling
17 in myeloid cells (215,216). ERE are present in the vicinity of genes encoding selected TLR (217), and studies in
18 microglia and macrophages obtained from wt and genetically modified mice have demonstrated that deletion of
19 the ERα DNA binding site blocks PAMPs/DAMPs-induced upregulation of TLR (218,219). Aside from this
20 activity, which suggests that estrogens increase the ability of microglia to respond to noxious stimuli, we also
21 know that estrogens inhibit the production of inflammatory cytokines by interfering with TLR signaling through
22 NF-kB and AP-1. Repeated studies have shown that p65 binding to its target genes is impaired by estrogens
23 through a non-genomic pathway involving modulation of the PI3K-dependent pathway (220). The hypothesis
24 that ERα inhibits NF-kB activity by inducing the synthesis of its inhibitory protein, IkBα, remains controversial.
25 Similarly, AP-1 may be involved in the actions of estrogen, as p85 PI3K signaling is involved in the estrogen-

dependent blockade of TLR4 in macrophages. Estrogens can block the activity of p38 by interfering with its phosphorylation (221), but whether this occurs through direct binding to the intracellular ER or through other mechanisms has not been investigated. These results support the hypothesis that estrogens act by reducing the inflammatory response. However, more recent findings indicate the possibility that estrogens exert a more widespread effect on macrophage activity by controlling their ability to transition among different activation stages. Using time-lapsed measurements of inflammatory cytokine production, one study demonstrated that estrogens may accelerate the resolution of LPS-induced inflammation by blocking IL-1β synthesis and increasing production of the anti-inflammatory IL-10 (222). The mechanism involved is of particular interest because in the absence of lymphoid cells that produce IL-4 to quench the inflammation, ERα would induce synthesis of SOCS3 through direct regulation of the *Socs3* gene promoter in microglia. SOCS3 is a transcription factor that is instrumental for the synthesis of IL-10, the main cytokine involved in the onset of the acquired deactivation status (223). Thus, through this action, estrogen would augment the intrinsic ability of macrophages to end the pro-inflammatory phase (Figure 6.). This anti-inflammatory activity of ERα would be even more valuable in the presence of other inflammatory cells that can terminate macrophage inflammation by secreting IL-4. In fact, this cytokine considerably increases the number of ERs in macrophages, therefore enhancing their anti-inflammatory potential. Thus, these studies indicate that the presence of the hormone estrogen and its ERα isoform facilitates both intrinsic and extrinsic programs for the resolution of inflammation and the direction of the LPS-stimulated immune cells toward the IL-10-dependent phenotype (acquired deactivation) responsible for tissue remodeling and restoration of homeostatic conditions (222).

This estrogen activity is particularly valuable in the case of chronic inflammation and in aging brains, where the maintenance of the microglial pro-inflammatory status may cause neuronal damage and could thus provide an explanation for the neuroprotective effects of estrogens demonstrated in models of neuronal injury and neurodegeneration (224,225).

Estrogens: protective or risk factors in brain injury and neurodegeneration?

Numerous studies using animal models in *in vitro* explant cultures or in observational studies and clinical trials involving humans have suggested that ovarian hormones play an important role providing women protection against stroke and neurodegenerative diseases (157,226-229). However, the mechanisms that enable such effects have not been fully elucidated. Dissecting the cell types targeted by estrogen has been slowed by the fact that ERs are expressed by all neural cells, and the neuroprotective effects likely result from receptor activation in more than one cell system. Furthermore, the cells involved may change depending on the nature of the disorder. This has broad implications for the selective targeting of ERs in the treatment of neurodegenerative conditions due to disease or injury, particularly in aging and in the post-menopause.

a. Estrogens and stroke or hypoxic neuronal death

In experimental models of stroke (middle cerebral artery occlusion, MCAO), 17β-estradiol attenuated cell death resulting from ischemic injury and promoted neuronal survival and tissue integrity (157,226). As stroke activates a significant microglial reaction, the question raised by these findings was whether the neuroprotective effects of estrogens are dependent on their anti-inflammatory activity and their ability to modulate the synthesis of neuroprotective factors, such as IGF-1, in microglia (230). In this experimental model, the protective effect was observed only when estradiol was administered immediately rather than weeks after ischemia. This may suggest that estrogens should be present at high levels when microglia become activated; therefore, its primary target should be microglia. However, the role that microglia play in MCAO was clearly demonstrated using the Cre-loxP system, which selectively deletes ERα in cells of myeloid lineage (Cre recombinase under the control of the lysozyme M promoter) or in neurons (Cre recombinase under CAMKII promoter). The neuroprotective role of 17β-estradiol in MCAO was maintained only in mice that possessed monocytes without ERα, thus leading to the conclusions that neuronal ERα mediates the neuroprotective role of estrogens and that microglia ERα is dispensable, at least in stroke. The study was performed in male and female mice showing superimposable outcomes (231). However, it is worth noting that the results obtained with this model have been the subject of discussion because the recombination in microglia does not appear to be very efficient (232).

b. Demyelinating diseases

1 Multiple sclerosis (MS) is a demyelinating disease characterized by a strong inflammatory component that is the
2 main contributor to myelin sheath destruction and an ensuing progressive paralysis. The fact that MS affects
3 women twice as often as men and that women may undergo clinical remission in the late stages of pregnancy
4 suggests that sex hormones a role in the development of this disorder (233). Indeed, clinical data and studies in
5 animal models of MS (e.g., experimental autoimmune encephalomyelitis [EAE]) support this hypothesis by
6 demonstrating that estrogens ameliorate EAE severity in both males and females (234,235). However, the
7 mechanisms through which estrogen exerts its beneficial effects in MS requires further investigation because
8 ERs are present in all neural cells affected by MS, including neurons, oligodendrocytes, Schwann cells (236),
9 and microglia. Considering the strong neuroinflammatory component of this disease, estrogen could act in
10 microglia by lessening its inflammatory reaction or minimizing the infiltration of circulating lymphocytes and
11 monocytes. This has been investigated in the EAE model using genetic and pharmacological approaches. Most
12 studies based on the administration of isoform-specific ligands of ERs (237) and ERα KO mice as myeloid cell
13 donors (238) indicated a key role for ERα in the protective effects of estrogen in EAE. However, ERβ also may
14 play a role in demyelinating disorders (239,240). The discrepancies in these previous results and conclusions
15 may be due to differences in the experimental model used (EAE; or demyelination induced by Theiler's virus or
16 cuprizone), the time at which the analysis was carried out and the fact that the two receptors may have different
17 functions. As suggested by the work of Brown *et al*. (210), the major involvement of ERβ might be relative to its
18 capacity to control the permeability of peripheral cells through the BBB. The activated ERβ might facilitate
19 peripheral lymphocyte migration into the CNS by secreting the interleukins that are necessary for dampening
20 neuroinflammation. Indeed, studies performed in B cell-deficient mice have shown that IL-10 administration
21 significantly improves the pathology (241). Finally, it is important to emphasize that not all MS animal models
22 are applicable for studying the effects of sex hormones. For instance, cuprizone administration disrupts the
23 estrous cycle, limiting the ability to establish sex differences (242).

24 ### c. Neurodegenerative diseases

25 Alzheimer's disease

Dementia is present in 16% of women and 11% of men aged over 71 years. This higher incidence in women was observed previously in age-matched groups, starting from 60–64 years up to 95 years of age. Therefore, it cannot be attributed to women having a longer life longevity. In women, β-amyloid accumulation is greater (243,244) than in man (245,246). This appears to be a characteristic feature of Alzheimer's Disease (AD) because no evidence of sex prevalence has been reported for mild cognitive impairment (MCI) or frontotemporal lobar degeneration (FTLD). Most animal models of AD (Tg2576, APPswe/PSEN1E9, APP23, APP_{swe}xPS1, and 3xTg-AD) reproduce the same sex specificity of Aβ accumulation and show a poorer behavioral performance than those reported in humans (247-251). It remains to be established whether the lack of ovarian functions plays a role in the sex-related differences in the incidence of AD. In the sporadic forms of AD, the association of homozygous single nucleotide polymorphisms (SNP) of the genes *ESR1* (rs9340799, rs2234693; rs2228480) and *ESR2* (rs4986938) with APOE4 (the best established genetic risk factor for AD) (112,252,253) conferred an increased risk of cognitive impairment in both sexes, with a higher prevalence in women. The explanation for the sex dimorphic effect of this association may reside in the fact that estrogen affects cholesterol and lipid transport, and in the brain, estrogen regulates the expression of low-density lipoprotein receptor-related protein (LRP), which has been implicated in Aβ processing. These observations suggest that an impaired ER signaling may constitute a predisposing factor to AD, but by itself, it is not sufficient to increase the risk of developing AD. Nevertheless, an understanding of how a lack of estrogens can modify the course of the disease would be extremely valuable from both therapeutic and social standpoints.

For many years, ApoE4 has been considered the best known risk factor for AD pathology and accounts for only 10%–20% of the sporadic AD risk. More recently, several independent genome-wide association studies (GWAS) have identified new common variants associated with sporadic AD (253). These findings have contributed to the diverging focus of the AD pathogenesis from the classical Aβ-centric view towards neuroinflammation (254-261). In fact, most of the genes associated with sporadic AD encode proteins relevant to immune cell functions (e.g., CD33 (257,258,262), CLU, BIN1, PICALM, CR1, CD2AP, EPHA1, ABCA7, MS4A4A/MS4A6E (254-258), and TREM2 (259,260)). For instance, the R47H variant of the TREM2 (Triggering Receptor Expressed on Myeloid cells 2) gene, was linked to the onset of AD with a probability

comparable to that for ApoE4 (259-261). Epidemiological studies further argue for a relevant role of neuroinflammation in AD because pathologies characterized by high levels of inflammation, such as vascular disorders and metabolic diseases, increase the risk and prevalence of AD (112,115,116). For example, the risk for AD is augmented by 60% in patients with diabetes mellitus (DM) (114). Thus, the influence of estrogens on microglial functions may play a role in AD. In animal models of AD, 17β-estradiol increased microglia viability *in vitro* and *in vivo*, whereas in humans, 17β-estradiol enhanced the uptake of Aβ in human cortical microglia (263), possibly by increasing the expression of the complement protein C3 (264), which plays a pivotal role in cytokine-induced activation of microglial phagocytosis (265). Finally, estrogens were shown to upregulate microglial proteasome activity through the p42/44 MAPK pathway, which is critical for a rapid and efficient turnover of oxidized or otherwise damaged proteins and therefore maintains microglial homeostasis in response to Aβ-induced activation and metabolic stress (266,267). In APP23 mice overexpressing human amyloid precursor protein with the Swedish mutation, ovary ablation increased microglia activation at Aβ deposits and facilitated the progression of these cells toward a highly reactive state (206). Long-term administration of 17β-estradiol blocked this effect and decreased microglia reactivity compared to control animals. In the same study, estrogens were shown to inhibit Aβ-induced expression of the scavenger receptor in macrophage cells. Considering that estrogen facilitates the resolution of the inflammatory process (222), it may also play a role in downregulating oxidative stress resulting from microglia hyperactivity. Despite the many lines of evidence that indicate estrogens have positive effects on the risk for AD, mixed results have been obtained with HRT when it was used to counteract the development and progression of AD. There are reports pointing to a beneficial role of long-term HRT on the risk of AD and the age at onset in postmenopausal women (268-270), whereas other reports question the overall benefits (271-273) of HRT. These discrepancies may be due to preexisting genetic and hormonal differences, the time at which HRT was started (274), the timing of the early onset of a neurodegenerative disease, and the type of HRT (e.g., presence/absence of progesterone). Progesterone treatment inhibited E2-mediated induction of neurotrophin expression and spatial memory performance (275). Similar effects were observed on Aβ accumulation after continuous progesterone treatment in adult female 3xTg-AD mice. However, the treatment had no effect by itself, but it counteracted the beneficial effect of E2 on Aβ

accumulation (276). Nevertheless, the observation that cholinergic activity is decreased by continuous treatments in the hippocampus of ovx female rats but enhanced by cyclic treatment with 17β-estradiol and progesterone (277) suggests that treatments designed to mimic the natural hormonal fluctuations that occur during the ovarian cycle might have beneficial effects on AD-related disorders. Indeed, it has been described that in female 3×Tg-AD mice, cyclic progesterone delivery counteracted the increase in Aβ resulting from ovx, and led to an enhancement of the Aβ-lowering effect of E2, along with significant improvements in working memory and visual attention (251). Moreover, progesterone treatment induced a reduction in tau hyperphosphorylation, thus suggesting a beneficial effect of cyclic P4 treatment in combination with E2 replacement in lowering the hallmarks of AD.

Parkinson's disease

Sex is one of the strongest risk factors for Parkinson's disease (PD), as men have a 2–fold greater relative risk for developing PD than women of all ages. Furthermore, the phenotypic characteristics and symptomatology of the disease are also sexually dimorphic (278). Sex-specific differences have been reported in the gene expression profiles of neurons obtained from the substantia nigra (SN) of PD patients, which may indeed underlie the sexual dimorphism in the disease etiology, symptoms and responses to therapy (279). The male prevalence of PD is also observed in PD animal models. Injections of neurotoxins (MPTP or methamphetamine in mice and 6-OHDA in rats) reduced the number of dopaminergic neurons in the SN and the dopamine levels in the striatum, with a higher potency observed in males using low doses of neurotoxins, possibly mimicking the early stages of PD (280,281) (282). The sex prevalence in PD may be associated with intrinsic differences in the brain structures affected by the disease, as well as with sex-related environmental factors, as both events are related to estrogen. In fact, the organization of the SN-striatal (SNDA) dopaminergic system is sexually dimorphic, with males presenting a higher number of neuronal cells and regulatory networks, potentiated adaptive responses to psychomotor stimulants and upregulated expression of genes related to familial PD (279). Considering the role that estrogen plays in brain development, and particularly its influence on the dopaminergic system, the sex differences in the NSDA system may be determined by the actions of estrogen on brain cells and somehow

favors PD development in men. On the other hand, strong evidence supports the hypothesis that estrogens contribute to the sex-related prevalence of PD in adults. This evidence includes the following: the inverse correlation between circulating estrogens and the severity of PD symptoms in women (283), the higher risk of PD in women with early natural or surgical menopause (284,285) and the higher prevalence of PD in climacteric women in comparable age groups (283,286). Furthermore, in animal studies of PD, estrogens were consistently shown to reduce the toxin-induced depletion of DA in the female striatum (280,281).

So far, no firm hypothesis has been put forward to explain how estrogens affect the manifestation of PD in females. The current view suggests that accumulation of misfolded proteins, mitochondrial and endosomal dysfunction and oxidative stress are the major biochemical mechanisms underlying the pathogenesis PD (287). However, a genetic association between a neuroinflammatory deficit and the pathogenesis of PD is still missing. As in the case of AD, microglia activation is strongly involved in the manifestation of PD, as well as a consequence or a cause of the selective vulnerability of neurons that produce dopamine, a highly reactive chemical that generates oxidative species and adds to the malfunctioning mitochondria, lysosomal and protein aggregates toxicity. The following key features link microglia to the pathological hallmarks of PD: *i.*) higher microglial density in the midbrain relative to other brain areas (32,288), likely as a result of the local oxidative environment; *ii.)* microglia proliferation induced by dopaminergic toxicants specifically in the SN, which is general sign of brain damage that is also observed in AD (289); the inflammatory response in the SN is different from other brain regions, such as the striatum or olfactory bulb, as it occurs independently of IL-1β; *iii.)* the delivery of LPS in the SN induces microglia activation, which selectively kills dopaminergic neurons and causes stable motor deficits (interestingly, serotoninergic neurons of the SN as well as neurons in the cortex and striatum are spared by this inflammatory insult (290,291)); and *iv.)* damaged dopaminergic neurons specifically release alpha-synuclein and neuromelanin aggregates that are potent triggers of microglia activation (292). Thus, microglia derangement can be considered as an "environmental" factor that may participate, either alone or in association with genetic variants, in the predisposition to PD.

1 Considering the sexual dimorphism in neuronal structures and functions affected in PD and the effects of
2 estrogens on microglia, the logical question to be asked is whether estrogen may delay the onset and reduce the
3 symptomatology of PD in females by targeting microglia. The fact that ER was not localized to DA neurons of
4 the striatum, whereas ERα, ERβ and GPR30 were expressed by microglia and interneurons in this brain region
5 (293,294) may point to the involvement of cells other than dopaminergic neurons that mediate hormonal
6 neuroprotection, and microglia may certainly be taken into consideration. Although still limited, the available
7 literature provides two main lines of evidence that strongly support the link between estrogen actions in
8 microglia and neuroprotection in PD: the sexual diversity in microglia reactivity, which triggers neuronal death
9 in males while providing neuroprotection in females, and the ruinous effects of menopause/ovariectomy on
10 neuroinflammation in females. In fact, a single peritoneal injection of LPS induces the selective loss of
11 dopaminergic neurons in the SN and motor behavior deficits in male mice, while repeated injections of the
12 endotoxin are necessary to induce neurotoxicity in females (295) (296). In line with this finding, estrogens
13 reduce microglia activation following LPS administration through the inhibition of the Mac-1 receptor and
14 PHOX protein complex, which regulate intracellular and extracellular ROS production (209,297) (56).
15 Accordingly, the genetic ablation of ERα in mice increases Mac-1 expression in microglia (209). Thus, these
16 results strongly suggest a link between the activity of estrogen in microglia and the peculiar reactivity of these
17 cells in females (209). Other approaches that more closely mimic the pathology of PD showed that the
18 expression of inflammatory mediators, such as TNFα, IL-1β, IFNγ and iNOS, are increased in the male NSDA
19 system and it is associated with an earlier and greater reduction in the striatal dopamine content in male mice
20 compared to female mice (298). Accordingly, Morale et al. showed that the toxic potential of activated microglia
21 following MPTP injury is inversely proportional to the levels of circulating estrogens (299). Thus, it appears that
22 the link between estrogen actions in microglia and female microglia reactivity may prove beneficial for the
23 manifestation of PD. More recently, it was shown that the expression of Mac-1 and other neuroinflammatory
24 genes is increased by ovx in the forebrain of middle-aged female rats, and this effect was reverted by the chronic
25 administration of estrogens (300). In agreement with these findings, increased expression of neuroinflammatory
26 genes was observed in the forebrains of pre- and postmenopausal women (205). Finally, recent evidence

highlighted the role that the renin-angiotensin system plays in microglia and showed this system to be more potentiated in males, leading to a higher neuronal loss in the SN (301), while its downregulation by estrogens results in reduced oxidative stress, neuroinflammation and neurodegeneration in females (302), (303).

In summary, the above mentioned studies allow one to draw a tentative summary of the microglial pathways that may play a role in the beneficial effects of estrogens on neurodegeneration. These effects include the following: i) the reduction of intra and extra-microglial oxidative stress through the restoration of mitochondrial functions and potentiation of reductive enzymatic systems; ii) the modification of damage-activated intracellular signaling that provides a faster healing process through the adjustment of microglia reactivity; and iii) altogether, these observations support the view that communication between estrogen and microglia is one the mechanisms that reduce the risk of PD in premenopausal women.

Concluding remarks and future directions

Given the involvement of the inflammatory process in neurodegeneration and the anti-inflammatory potential, it seems reasonable to further evaluate how the actions of estrogen in microglia might influence the onset and progression of neurodegenerative diseases. However, in undertaking these studies, we should consider the numerous factors that can constitute a confounding element in the interpretation of the results. A summary of the mechanisms that aging and the lack of estrogens may perturb in microglia is shown in Figure 7.

The main factor to be taken into consideration is the model system that is used to study the effects of the hormones; the major limitation of primary cultures of neonatal microglia or transformed cells is represented by their bias in the ER expression that may not reproduce what is occurring in microglia of the mature brain. Cultures of microglia isolated from the adult brain may be a better model. However, the low recovery of the current isolation procedure represents a major limitation and in addition the culture would not provide these cells with the stimuli necessary for their continuous surveillance and response to the environment. Perhaps

biochemical studies aimed at studying microglia responsiveness to physio-pathological or pharmacological stimuli should be assessed in cells freshly isolated from the brain. The use of Fluorescence Activated Cell Sorting (FACS) and transgenic mice carrying appropriate reporters for microglia identification may help to overcome the low efficiency of current methodologies for microglia isolation. Alternatively, microglia may be studied in living organisms. In this case a rigorous characterization of the model used and of the experimental setting is necessary to obtain reproducible and meaningful results because our understanding of estrogen physiology is still in its infancy. Several authors study estrogen in intact males to avoid the influence of high levels of this hormone in the circulation. This, besides limiting the vision of the research to the sex likely less influenced by female sex hormones, may give variability in the results due to the circadian synthesis of testosterone and the presence/absence of aromatase converting the male sex hormone into estrogens. Studies in females rely on the ovx/hormone replacement paradigm, which limits the interpretation of the outcome because we know very little on the endocrine compensatory reactions induced by the removal of the ovaries. Indeed, we may find totally different effects of pharmacological or hormone replacement that is dependent on the time of ovx. A better understanding of the physiological functions of estrogens in intact males at different hours of the day, and in intact females in the different phases of the cycle, is mandatory to obtain reproducible and meaningful observations. The selection of the correct route, dosage and timing of estrogen administration is also challenging, as it is in all cells targeted by estrogen, including microglia, and the response may dramatically change, leading to conflicting and uninterpretable results. To this aim, the use of appropriate reporter animals (i.e., animals genetically modified to produce easily measurable proteins in response to a selective stimulus (304) should be encouraged, as these animals allow for the *spatio*-temporal analysis of specific biochemical pathways in single, living animals, thus facilitating the interpretations of the physiological changes occurring over time. For example, the available reporter animals could facilitate the identification of the phase of the cycle and the cells actively responding to estrogens, whereas others would facilitate the identification of microglia in the activated or deactivated status, and these reporter systems could be bred with each other to obtain the analysis of multiple end points at the same time.

The other confounding element in the study of the effects estrogen in microglia *in vivo* is represented by the fact that all neuronal cells are capable of expressing ERs. For instance, estrogens target neurons where they may exert a direct neuroprotective function. This represents a confounding element for defining the microglial contribution to neuronal health. Once more, the use of appropriate reporter animals would sharpen our vision and facilitate the study of microglial activity during the development of the pathology in a specific model of disease. These models would also be amenable to the study of the effect of age, nutritional cues and dietary interventions. Indeed, prior results have shown that age and nutritional status have major, sex-dependent effects on microglia activity, which must be taken into consideration in future studies (Box 1).

In conclusion, the large body of experimental evidence provided so far indicates that microglia represent another target for the neuroprotective actions of estrogen. Indeed, as a plausible factor driving microglia colonization of the nervous system, as well as a modulator of microglia reactivity in adult brain, estrogens appear to play a role in the neurodegenerative process, conditioning the incidence of these pathologies as well as the course of their progression. The lack of a direct, strong linkage between estrogen receptor mutations and neurodegenerative diseases suggests that these sex hormones do not play a primary role in promoting the progression of the neurodegenerative program, which, in the sporadic forms of these disorders, is highly multifactorial. However, the impairment of estrogenic signaling in combination with a lack of other elements relevant for neuronal health may facilitate the initiation of the neurodegenerative process as shown by the studies on the correlation between ER signaling and ApoE4 (305). The fact that estrogens are not primary contributors, but only participate in the complex combination of the events necessary to trigger the neurodegenerative process, represents the main obstacle for the study of the effects and the definition of adequate replacement therapies.

The identification of further correlations between estrogen deficiencies and pathologies of the CNS characterized by the significant neuroinflammatory component may provide a means for the study of the efficacy of replacement therapies, but time and cost factors may be unsuitable for what is needed. Such therapies, aimed at reconstituting the natural defenses of the brain against neuroinflammation might be less amenable to undesired collateral effect than exogenous molecules such as sodium thiosulfate (306), mito-apocynin (307) or kolaviron

(308) recently proposed for the reduction of neuroinflammation. In the near future, efforts should be mainly aimed at a better understanding of the physiology of estrogen actions in the microglia of males and females. This knowledge is vital for the design of appropriate hormone replacement therapies that can overcome the lack of the natural hormone in targets, sparing their potential negative effects in reproductive organs.

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Figure Legends

Figure 1. Microglia are a dynamic mediator of synaptic development and homeostasis. Microglia in its surveying state senses the state of activity of neurons, is attracted to the dendritic spines through proteins of the complement and fractalkine and participates in neuronal plasticity potentially through the release of proteases able to modulate the structure and functions of the synapses. Microglia possibly respond to the release of ATP which may induce the shedding of lipid-rich vescicles that were reportd to increase the frequency and amplitude of the excitatory post-synaptic potential (EPSC).

Figure 2. Brain development: the sexually dimorphic activity of microglia. Primitive macrophages exit the yolk sac blood islands at the onset of circulation and colonize the neuroepithelium from E8.5 to give rise to microglia. The blood brain barrier starts to form from E13.5 and may isolate the developing brain from the contribution of fetal liver hematopoiesis. Embryonic microglia expand and colonize the whole CNS until adulthood. The high concentration of chemokines in the male brain facilitates microglia proliferation.

Figure 3. Morphological and functional elements point to alterations of microglia activity with age.

1 **Figure 4. Structure and PTM sites of the nuclear estrogen receptors.**

2 Like all steroid receptors, the ERs belong to a family of hormone modulated-transcription factors characterized 3 by the presence of 6 functional domains: *A-B*. the N-terminal domain that contains the activation function 1 (AF-4 1) enabling the interaction with co-regulators also in the absence of the ligand; *C*. the highly conserved DNA 5 binding domain (DBD), responsible for the recognition of specific DNA sequences (named estrogen responsive 6 elements, ERE) through the two Zn fingers; *D*. the hinge region, a flexible domain that connects DBD with the 7 ligand binding domain (LBD) able to influence intracellular trafficking and subcellular distribution; *E.* the LBD 8 responsible for ligand recognition that contains the ligand-dependent activation function 2 (AF-2); the LBD, 9 contributes to the dimerization interface of the receptor in concert with the DBD; *F.* the C-terminal domain that 10 participates in the binding to ligands. Both ERs unrdego a large number of regulatory post-translational 11 modifications exemplified in the figure (human ERα and murine ERβ).

12 **Figure 5. Aging effects on circulating estrogens.** The uterus weight as a biomarker of circulating estrogens 13 shows that, in mice, the activity of the ovaries does not decrease with age: actually at 18 months when mice are 14 not cycling the plasma content of this hormone is higher than in young, fertile animals. Ovariectomy clearly 15 decreases the circulating levels of the hormone, showing that organs other than ovaries give a minimal 16 contribution to steroidogenesis.

17 **Figure 6. Estrogen and microglia functions.** Estrogens regulate microglia inflammatory potential by 18 interfering with the process of NFkB activation (a) and by facilitating the transition to the stages where 19 microglia exert neuroprotective functions (b), possibly including the maintenance and pruning of dysfunctional 20 synapses (c).

21 **Figure 7. Estrogen-dependent protective effects of microglia.**

22 Several biochemical processes promoted by microglia and regulated by estrogens protect neuronal functions: *i*.) 23 phagocytosis clears the debris and dysfunctional proteins (i.e., β-amyloid) in the parenchyma, *ii*.) the production 24 of antioxidant systems and enzymes (i.e. the renin-angiotensin axis) limits the oxidative stress; *iii.)* the healing 25 process is facilitated when damage-activated intracellular signaling pathways are activated, *iv.)* synaptic

1 maintenance participates in neuronal signaling. With aging misfolded proteins, cell debris and other
2 inflammatory stimuli accumulate in the brain parenchyma inducing a continuous stimulation of microglia that
3 with senescence has a decreased phagocitic potential and ability to return to the surveying state. This initiates a
4 vicious cycle with a progressive increase of the production of inflammatory products detrimental for neuronal
5 health.
6
7 Box 1. **Critical issues for study estrogen action in microglia and the definition of replacement therapies.**
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9
10

1 **TABLE I. Molecular characterization of microglia phenotypes**

	Inflammatory (M1)	Alternative Activation (M2a)	Type II alternative activation (M2b)	Acquired Deactivation (M2c)
Function	Killing of intracellular pathogens Pathogen phagocytosis Extracellular matrix degradation	Extinction of inflammatory response Killing of encapsulated parasites		Immunoregulation Engulfment of apoptotic/dead cells Extracellular matrix deposition Tissue remodeling
Markers	**↑ TLRs** **↑ CR1, CR3, CR4** **↑ CD36, CD91** **↑ RAGE** **↑ NF-κB** **↑ TNF-α** **↑ IL-1β, IL-6** **↑ IL-12, IL-23** **↑ CCL2** **↑ iNOS, PHOX** **↑ MHC-II** **↑ MMPs** **↑ TREM2** **↑ IL-6** **↑ CD14** **↑ CD40** **↑ CD74** **↑ CD68**	**↑ Polyamines** **↓ IL-12** **↓ iNOS** **↑ IL-1ra** **↑ CD163** **↑ CD206** **↑ MHC-II** **↑ *Arg-I, Ym-1, Fizz-1*** **↑ TREM-2** **↑CD 33**	**↑IL-10** **↓ IL-12** **↑CD16** **↑CD32** **↑CD64** **↑ MHC-II**	**↑ TGFβ** **↑ IL-10** **↓ IL-12** **↑ Versican** **↑ PTX3** **↑ MARCO,** **↓ MHC-II** **↑ TIMP1** **↑ CD163**
Stimulus	IL-1β; TNFα; (IL-6)	IL-4; IL-13	LPS; IL-1β	IL-10; TGFβ
References	(309-312)	(59,313,314)	(315)	(59,313)

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3

1 **Table II. Nuclear Receptor expression in microglia**

Source of microglia	Method of isolation	ERα	ERβ	GPR30	AR	PR	GR	MR	References
		mRNA content (RPKM*)							
BV-2	-	0.0	0.0	0.0	0.0	0.0	3.0	0.0	Crotti *et al. (193)*
Adult mouse whole brain	CD11b+ magnetic separation	1.5	0.0	1.5	0.1	0.0	42.2	3.5	Maggi *et al. Unpublished data*
Adult mouse whole brain	Percoll/FACS	1.4	0.0	0.3	0.0	0.0	196	0.0	Lavin Y *et al.* (195)
Mouse brain cortex	FACS	3.4	0.0	0.6	0.0	0.1	11.9	1.8	Zhang Ye *et al.* (194)
Mouse brain cortex	Single cell RNAseq	0.02	0.00	0.05	0.00	0.00	1.5	0.05	Zeisel *et al.* (316)
Spinal cord	Percoll/ CD11b- magnetic separation	0.1	0.0	n/a	0.0	0.0	2.9	0.6	Chiu *et al*. (317)

2 * Reads Per Kilobase of transcript per Million mapped reads.

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3 ### *References*

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