DNA extraction CTAB2016-02 03.05.2016 — Peter Schuchert

Tissue: alcohol fixed, remove all alcohol, some little residue may remain
Buffer 1x CTAB: 0.7 M NaCl, 10 mM EDTA, 50 mM Tris pH 8.0, 1% hexadecyltrimethylammonium bromide CTAB

DNA number	sample number	species		locality	dissolved in µl TE	~yield ng/µl	ABI reactions
1143	136b	***Aegina***	***citrea***	Friday Harbour	400	>10	
1144		***Ptychogastria***		Antarctica	400	>10	
1145	Vfr16.01	***Oceania***	***armata***	Villefranche	400	>10	
1146	Vfr16.02	***Oceania***	***armata***	Villefranche	400	>10	
1147	Vfr16.03	***Oceania***	***armata***	Villefranche	200	10	
1148	Vfr16.04	***Oceania***	***armata***	Villefranche	400	10	
1149	Vfr16.05	***Geryonia***	***proboscidalis***	Villefranche	400	>10 dilute 2X	
1150	Vfr16.08	***Cunina***	***octonaria***	Villefranche	200	<0.5	
1151	Vfr16.09	***Laodicea***	***undulata***	Villefranche	200	5	
1152	Vfr16.11	***Zanclea***		Villefranche	300	5	
1153	Vfr16.14	***Helgicirrha***	***schulzii***	Villefranche	300	5	
1154	Vfr16.15	***Zanclea***		Villefranche	300	10	

1. add CTAB 1x buffer ~~200~~ 300 ul (=1 volume)
2. add 10 µl Proteinase K 10 mg/ml
3. 55°C from 1230 to 0830
4. add 1volume of chloroform+isoamylalcohol (=3-Methyl-1-butanol) 24+1, vortex vigorously
5. centrifuge 5 min. max. rpm
6. upper phase in new eppentube, avoid any contamination with the interphase or lower phase
7. add 1 ul linear acrylamide 25µg/µl (Sigma GenElute)
8. add 2 volumes of ethanol 100%, mix
9. 15 min centrifugation at max. rpm
10. pipet out liquid, gently wash with 0.2 ml 70 % ethanol
11. dry DNA at 55°C 5min
12. dissolve in TE buffer as given above (normally 100 µl)
13. ~~8~~ 6 µl on gel