

# Association of *Juniperus deppeana* (Cupressaceae: Pinales) seeds with Mexican cottontail rabbit (*Sylvilagus cunicularius*; Leporidae: Lagomorpha) latrines

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#### ABSTRACT

Seed aggregation in latrines of rabbits is a little-studied process that may contribute substantially to seed dispersal and survival. We located Juniperus deppeana trees and the latrines used by the endemic Mexican cottontail rabbit Sylvilagus cunicularius within a 1 ha fragment of J. deppeana-dominated forest and evaluated their patterns of aggregation as microhabitats used by cottontail rabbits to consume the J. deppeana galbulus (fleshy cones) or to deposit their pellets with defecated seeds. Based on mean latrine area, we marked a  $2 \text{ m} \times 2 \text{ m}$ plot centred on the base of every tree or latrine within the study site. We counted all pellets within each plot and analysed them for the presence of seeds of Juniperus or any other species. We found seeds in 13% of all pellets, mainly those deposited in the latrines. If seeds are scarified by passing through the intestinal tract, seed germination in latrines may increase the probability of survival and establishment. Thus, the community structure and density could change in time as rabbits are changing the places where they place their latrines.

#### **ARTICLE HISTORY**

Received 28 May 2014 Accepted 7 June 2016

#### **KEYWORDS**

Endozoochory; rabbits; lagomorphs; seed aggregation; Juniper galbulus

# Introduction

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In plant-disperser interactions, the time and place of seed deposition play critical roles in seedling establishment and regeneration of plant populations (Schupp et al. 2010). Seed dispersal is linked to the disperser's movement patterns and behaviour, which may determine the site and conditions dispersed seeds face, the pattern of seed aggregation, the probability of seedling establishment and plant survival, and, ultimately, species diversity and spatial distribution patterns (Williams et al. 2000; Houseman 2014). For example, birds such as many species in the Turdidae, Sylviidae and Muscicapiade are well-documented seed dispersers (Herrera 1995). Additionally, mammals such as primates (LoGiudice and Ostfeld 2002; Wehncke et al. 2003) and bats (Muscarella and

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Fleming 2007; Bolívar-Cimé et al. 2015) are known dispersers, but less is known on the role of lagomorphs.

Lagomorphs make a significant contribution to ecological systems by consuming fruits and dispersing seeds in arid and semi-arid environments (Muñoz-Reinoso 1993; Nogales et al. 1995; Izhaki and Ne'eman 1997; Castro et al. 2008), dune systems (Calviño-Cancela 2002, 2004; Pakeman et al. 2002; Dellafiore et al. 2006, 2010; Larrinaga 2010), pasturelands (Malo and Suárez 1995; Malo and Yanes 1999), thickets (Staniforth and Cavers 1977; Cosyns et al. 2005), wetlands (Zedler and Black 1992; Chang et al. 2005), and woodlands and abandoned lands (Schupp et al. 1997a; Escribano-Avila et al. 2012). These studies have highlighted the role of lagomorphs as important seed-dispersal agents in diverse environments in conserved and disturbed landscapes. However, none of these studies related the number of seeds dispersed to different microhabitats and to temporal variations to describe the natural history and behavioural ecology of rabbits.

Lagomorphs are ideal subjects to study the effects of seed aggregation on landscape dynamics because they deposit most of their faecal pellets in fixed sites, called 'latrines' or 'dunghills' (Sneddon 1991; Zollner et al. 1996). The importance of latrines for seed dispersal and establishment needs to be evaluated due to their potential effects on community structure and composition (Pakeman et al. 1999; Verdú et al. 2009).

The Mexican cottontail Sylvilagus cunicularius Waterhouse (Leporidae: Lagomorpha) is endemic to Mexico (Vázquez et al. 2007) and occurs over a wide altitudinal range, from arid lowlands at sea level to temperate highlands at 3500 m above sea level (asl) (Cervantes et al. 1992). This rabbit inhabits pine and oak-pine forests which includes the fragmented juniper forests along the Mexican Transvolcanic Belt, all with an understory of clumped grasses (Fa et al. 1992). Sylvilagus cunicularius feeds on fruits, young leaves of forbs, shrubs, grasses and cultivated plants, and deposits its pellets mainly in latrines (Cervantes et al. 1992) which are typically placed in open spaces, perhaps as a means to avoid predation. The Mexican cottontail rabbit is sympatric with Sylvilagus floridanus Allen and Romerolagus diazi Ferrari-Pérez, but the pellets from these species can be distinguished with high accuracy based on size, shape and colour (Fa et al. 1992). Junipers are among the few deciduous gymnosperms that develop fleshy cones that function similarly to the fleshy fruits of angiosperms (Herrera 1992). Female cones of Juniperus deppeana Steudel may contain one to three seeds, and the peak fruit production is between December and June (Martínez et al. 2007). When eaten by birds or lagomorphs, the seeds pass through the gut without suffering apparent damage and can thus be dispersed (Adams 1993; Schupp et al. 1997b; Chambers et al. 1999).

In this respect, the goal of this study was to assess the temporal and spatial patterns of *Juniperus deppeana* seed dispersal by the endemic Mexican rabbit *Sylvilagus cunicularius*. We hypothesised that more *Juniperus deppeana* seeds will be found in rabbit latrines when compared to other microhabitats, opening the possibility of latrines contributing to the recruitment of new plants.

# **Materials and methods**

#### Study site

The study site (19°20'4.81"N, 98°21'51.60"W; 2300 m asl) was located 14 km west of the city of Tlaxcala, in east-central Mexico. It is a 10 ha fragment of an original *J. deppeana* 

forest within a grassland dominated by *Muhlenbergia implicata* (Kunth) Trinius, *Stipa ichu* (Ruiz and Pavón) Kunth, *Aristida schiedeana* Trinius and Ruprecht (Poaceae), and *Rhus standleyi* F.A. Barkley (Anacardiaceae) (Hudson et al. 2005). This fragment is surrounded by corn fields and forested fragments where *Eucalyptus globulus* Labillardière was introduced in 1980. The climate is temperate subhumid, with a mean annual rainfall of 800–1000 mm, mainly during the summer months, from June through September (only 5% of the precipitation falls during winter). Furthermore, the site has a topographic slope of 10°, causing a slight runoff during rains.

## Sampling design

A 100 m × 100 m plot was randomly selected at the study site and all *J. deppeana* trees (female, male and juvenile trees) and rabbit latrines were located. The total crown cover of *Juniperus* within the plot was about 4000 m<sup>2</sup>. The area of each latrine and of the crown of each juniper was estimated as an ellipse by measuring the longest axis and the axis perpendicular to that. The sampling area of the latrine and under the crown of each juniper tree was standardised with respect to the mean latrine size ( $\bar{X} = 3.3 \pm 0.08 \text{ m}^2$ , standard error (SE)), rounded up to 4 m<sup>2</sup> to have uniform areas to compare. Pellets were found on bare ground and below juniper crowns but were absent from other habitats, such as pasturelands and shrublands. In addition, the numbers of pellets in other habitats are low and the highest concentrations of pellets are located in latrines.

## Pellet and seed collection

A 4 m<sup>2</sup> plot was centred on each latrine and a 4 m<sup>2</sup> plot was also set below each *Juniperus* tree by tracing a 1.13 m radius and the distance at which the sample area was 30 cm from the trunk of the plant. At the beginning of the sampling period (first week of July 2004), all fresh (recently defecated) pellets within each plot were collected, but the older pellets were left *in situ* to keep the odor mark of each latrine (Sneddon 1991; Zollner et al. 1996). All fresh pellets (i.e. not older than the sampling period) were collected roughly every 2 months from 15 August 2004 through August 2005. The pellets were counted, collected and stored in paper bags until each one was examined under a stereoscopic microscope to extract the seeds, which were separated by appearance and weighed on a digital scale (ADAM AAA100L, Danbury CT, USA) to the nearest 0.01 mg.

#### Statistical analyses

We analysed the latrines and trees (female, male and juvenile trees) on a three-dimensional (x, y, z) coordinate system to identify their spatial distribution (where x and yindicate the Cartesian location in metres inside the 100 m × 100 m study area, and z is the area of latrines or the canopy of each juniper). Data were tested using RookCase spatial autocorrelation analysis with the equations for global Moran's I join counts with Queen's adjacency coefficient of contiguity between latrines and trees (four cardinal plus four diagonal neighbours). This is a test to explore the spatial association between latrines and plant canopies. The significance was assessed using Moran's Index using a Monte Carlo test with 500 permutations (Sawada 1999). No autocorrelation would indicate that latrine location distribution is independent of tree gender and size (Legendre 1993).

The density of pellets and number seeds per pellet were analysed with repeated measures analysis of variance (ANOVA) with microhabitat (latrine or tree) as a fixed factor and time as the repeated measure. The data were previously tested for homogeneity of variances and normality, and rank-transformed when necessary (Conover and Iman 1981). All analyses were made with Statistica 7 (Stat Soft, Inc. Tulsa, USA).

#### Results

#### Spatial association

There was no spatial association between the 105 latrines, which occupied 347 m<sup>2</sup>, and the 328 *J. deppeana* trees, which occupied 4000 m<sup>2</sup> (I = 0.008, P = 0.35; Table 1), indicating a random distribution: therefore, rabbit latrines are not under the canopies of the plants. The mean distance between latrines and junipers was  $30 \pm 1.8$  m (standard deviation (SD)), and the mean crown size of junipers was  $12 \pm 1.6$  m<sup>2</sup> (SD).

#### Pellet deposition

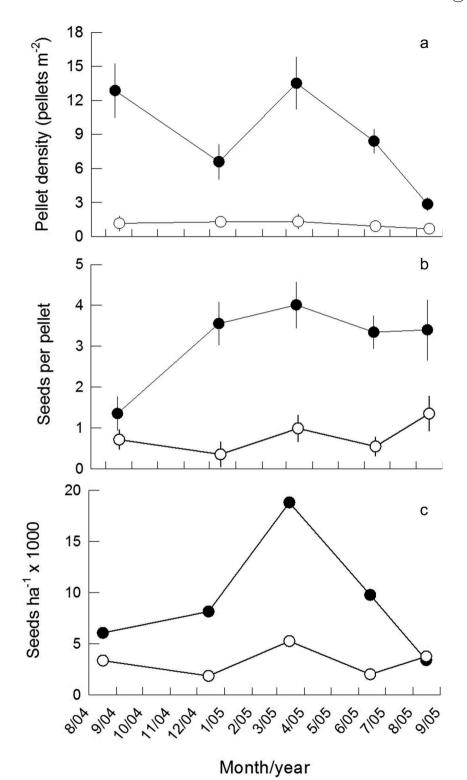
Only one rabbit species (*Sylvilagus cunicularius*) was registered at the study site, as indicated by the homogeneous size and appearance of the pellets (Fa et al. 1992). There were no significant differences between types of trees in the density of deposited pellets and seeds ( $F_{2,431} = 1.2$ , P > 0.05, and  $F_{2,431} = 1.6$ , P > 0.05). There was a significantly higher density of pellets in latrines than under the juniper trees ( $F_{1, 431} = 362$ , P < 0.0001). In addition, there was a significant effect of time ( $F_{4, 1724} = 26$ , P < 0.0001) and a significant microhabitat × time interaction ( $F_{4, 1724} = 16$ , P < 0.0001), with the most pellets counted in September 2004 and April 2005 in latrines, while seed counts beneath trees were relatively uniform throughout the study (Figure 1(a)).

#### Seed deposition

There were seeds in 14% of the 25,649 *S. cunicularius* pellets we examined; 13% had *J. deppeana seeds*, 1% *Opuntia tomentosa* var. *tomentosa* seeds and 0.1% had unidentified Solanaceae seeds. There were pellets with seeds in all latrines and in all plots under the junipers.

**Table 1.** Moran's *I* and normal *Z*-values for the autocorrelation by join counts (Queen's adjacency) between latrines (*I*) and female, male and juvenile *Juniperus deppeana* trees (*t*). Values in parentheses indicate the expected value of join counts for each category. P > 0.05 in all cases.

Association	tl joins	Z	tt joins	Z	<i>II</i> joins	Ζ
Latrine – female	5 (63)	-1.4	119 (162)	-0.87	107 (6)	9
Latrine – male	6 (67)	-1.35	171 (211)	-0.79	107 (5)	10
Latrine – juvenile	2 (57)	-1.68	48 (92)	-1	107 (9)	7



**Figure 1.** (a) Density of *Sylvilagus cunicularius* pellets; (b) number of *Juniperus deppeana* seeds per pellet; and (c) number of seeds in the hectare under study deposited under trees (empty symbols) or in latrines (filled symbols) from August 2004 through August 2005. In (a) and (b), vertical bars indicate one SE.

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The repeated measures ANOVA indicated a significant effect of microhabitat  $(F_{1, 431} = 188.1, P < 0.0001)$ , time of collection  $(F_{4, 1724} = 10.1, P < 0.0001)$ , and microhabitat × time interaction  $(F_{4, 1724} = 11.3, P < 0.0001)$ . The Fisher's least significant difference multiple comparison procedure indicates there were more seeds in pellets from latrines than in pellets below the trees, except in September of 2004 (Figure 1(b)). Multiplying the density of pellets by the number of seeds per pellet and by the area covered by latrines or tree canopies, we found that seeds from latrines accounted for 47.5 to 83% of the total number of seeds per hectare; numbers of seeds were similar in the two microhabitats in September, but in March many more seeds were found in latrines (Figure 1(c)).

# Discussion

We found that *S. cunicularius* deposited a greater density of seeds in latrines than below the canopy of *J. deppeana*, and that there are spatiotemporal differences in the number of seeds deposited even considering the relative cover of both habitats, suggesting that rabbits play a very clear role as dispersers of juniper seeds. The patchy deposition of seed-containing pellets in latrines suggests that recruitment of *J. deppeana* individuals could benefit from latrine-use behaviour. However, we only address a small part of the recruitment process and do not evaluate seedling recruitment and survival. Further work is necessary. The proportion of pellets of the Mexican cottontail rabbit *Sylvilagus cunicularius* with seeds (14%) was higher than the 3% of pellets containing *Retama sphaerocarpa* Boissier seeds reported for *Oryctolagus cuniculus* Linnaeus (Malo and Yanes 1999), or the 7% of *O. cuniculus* pellets having seeds belonging to several shrub species (Malo et al. 2000), but lower than the proportions found in other species of arid zones, which range from 43 to 64% (Nogales et al. 1995; Izhaki and Ne'eman 1997; Santos et al. 1999). Such differences could be related to the variability of fruit production from year to year.

Our results further suggest that the high density of seed dispersal by rabbits to latrines may have influence on the population dynamics, structure and colonisation of plant communities, as proposed for grassland (Malo et al. 1995; Malo and Suárez 1995; Pakeman et al. 1999) and dune (Dellafiore et al. 2010) ecosystems. There is also a possibility of extensive secondary dispersal if rainfall washes the pellets away from the latrines (Nogales et al. 1995), especially in highly sloped sites such as in our own study site.

There is a clear temporal pattern in seed deposition in latrines, which seems linked to the period when most *Juniperus* female cones are available, which is from June to December (Martínez et al. 2007). These months are characterised by heavy rainfall and water runoff, likely resulting in the secondary movement of many pellets and seeds away from the latrines. This probably influences *J. deppeana* recruitment of seedlings in or downslope of the latrines, as we have indeed observed in the field. Also, seed germination and sapling survival and growth may be enhanced, because latrines may provide nutrients, organic matter and humidity that ameliorate the conditions of formerly bare soils (Willott et al. 2000; Verdú et al. 2009). The concentration of seeds could increase their vulnerability to predation (LoGiudice and Ostfeld 2002)

or, contrariwise, increase the probability that a few of them may germinate to become seedlings.

In rabbits, the intestinal retention period may be between 20 and 30 hours (Illius and Gordon 1992; Cosyns et al. 2005), allowing sufficient time for seeds to be dispersed long distances (up to 150 m) from the parent plant (Cosyns et al. 2005). While the home range of *S. cunicularius* is unknown, estimates for other *Sylvilagus* species of similar body size suggest it is likely between 0.2 and 7.7 ha (Bond et al. 2001). Due to their pellet-depositing behaviour and subsequent dispersal, rabbits are in fact moving the seeds away from the parent plants (Calviño-Cancela 2002).

We did not find seeds or cones in bird droppings or any other animal faeces, suggesting that rabbits are the major dispersers. In this sense it has been reported that O. cuniculus disperses 14% of the seeds of Corema album (Linnaeus) D.Don ex Steudel, which is relatively low compared with the 40% dispersed by the blackbird Turdus merula Linnaeus (Passeriformes: Turdidae) (Calviño-Cancela 2002). Hence, rabbits may be important dispersal agents, especially in areas with sparse plant cover (Santos et al. 1999), and abandoned agricultural fields (Escribano-Avila et al. 2012), or when there is a lack of other dispersers. In our study system, rabbits are feeding on the fruits of J. deppeana, which are hard, fibrous and brown, characteristics suggested to be associated with mammal rather than bird dispersal of juniper seeds (Chambers et al. 1999). In addition, the fruits fall to the ground beneath the trees. Therefore, it is expected that dispersal will be predominantly by mammals such as lagomorphs and the ring-tailed cat Bassariscus astutus Lichtenstein (Carnivora: Procyonidae). Latrine formation and distribution in S. cunicularius can thus modulate spatiotemporal differences in the density of deposited seeds, but field experiments and observations should be done to test whether this increases the potential of J. deppeana to colonise new microhabitats.

## Acknowledgements

We thank F. Aureli and M. Calviño-Cancela for comments on an earlier draft, and E.W. Schupp and another, anonymous reviewer for many suggestions and constructive criticism. This work was supported by the projects UVER-PTC-223 and PFA C-703/2013 to AJM, as well as doctoral scholar-ships 218107 (ELD) and 260164 (PSP).

#### **Disclosure statement**

No potential conflict of interest was reported by the authors.

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