## 1 Biorefinery of cellulosic primary sludge towards targeted Short Chain Fatty Acids,

2 phosphorus and methane recovery

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# 11 Abstract

12 Cellulose from used toilet paper is a major untapped resource embedded in municipal 13 wastewater which recovery and valorisation to valuable products can be optimized. Cellulosic 14 primary sludge (CPS) can be separated by upstream dynamic sieving and anaerobically digested to recover methane as much as  $4.02 \text{ m}^3$ /capita·year. On the other hand, optimal acidogenic 15 16 fermenting conditions of CPS allows the production of targeted short-chain fatty acids (SCFAs) 17 as much as 2.92 kg COD/capita·year. Here propionate content can be more than 30% and can 18 optimize the enhanced biological phosphorus removal (EBPR) processes or the higher valuable 19 co-polymer of polyhydroxyalkanoates (PHAs). In this work, first a full set of batch assays were 20 used at three different temperatures (37, 55 and 70 °C) and three different initial pH (8, 9 and 21 10) to identify the best conditions for optimizing both the total SCFAs and propionate content 22 from CPS fermentation. Then, the optimal conditions were applied in long term to a Sequencing

23	Batch Fermentation Reactor where the highest propionate production (100-120 mg COD/g
24	TVS <sub>fed</sub> ·d) was obtained at 37°C and adjusting the feeding pH at 8. This was attributed to the
25	higher hydrolysis efficiency of the cellulosic materials (up to 44%), which increased the
26	selective growth of Propionibacterium acidopropionici in the fermentation broth up to 34%.
27	At the same time, around 88% of the phosphorus released during the acidogenic fermentation
28	was recovered as much as 0.15 kg of struvite per capita year. Finally, the potential market value
29	was preliminary estimated for the recovered materials that can triple over the conventional
30	scenario of biogas recovery in existing municipal wastewater treatment plants.
31	
32	Keywords: cellulosic primary sludge; acidogenic fermentation; propionate; resource recovery;
33	struvite
34	Highlights
35	• Separation and fermentation of cellulosic primary sludge enable wastewater-based
36	biorefinery
37	• Initial pH (8) and T (37 °C) selected Propionibacterium acidopropionici which
38	optimized the propionate recovery
39	• Struvite recovery from CPS fermentation liquid best integrates the biorefinery concept
40	• The CPS-based biorefinery could triple the economic value of wastewater
41	1. Introduction
42	Primary sludge (PS) usually contains a large quantity of biodegradable organic compounds such
43	proteins, carbohydrates, cellulose and other organic materials. Among them, cellulose
44	represents approximately 30-50% of the influent suspended solids in wastewater treatment
45	plants (WWTPs) of Western European countries (STOWA report, 2012) where toilet paper is

46 flushed into the sewers system. In these countries, the average per capita consumption of toilet 47 paper was estimated around 15 kg per year which is 3 times more than the global average 48 consumption (4.4 kg/capita·year) and 10 times more if compared with the consumptions of 49 developing countries (Pulp and Paper Industry Intelligence, 2011). Due to the flushing of toilet paper in public sewers, in Western European countries cellulose usually enters municipal 50 51 wastewater treatment plants (WWTPs) and is only partially degraded and valorised. However, 52 flushing toilet paper may probably be considered more environmentally friendly practice 53 compared to disposal in toilet trash and following transportation to landfills or incinerators, that 54 is implemented in countries where sewers infrastructure can have clogging problems (Genty et 55 al. 2013).

Usually, the rate-limiting step of cellulose degradation is the hydrolysis process (Noike et al.,
1985), which makes difficult its degradation during the conventional biological treatments in
WWTPs.

59 Verachtert et al., (1982) reports that 60% of the cellulosic material is degraded during 4-5 weeks 60 of aerobic conditions, while 40% persists undegraded in the excess sludge. However, if 61 anaerobic digestion of excess sludge is accomplished, additional 50% of the present cellulose 62 could be degraded.

Ruiken et al, 2014 carried out batch experiments to investigate the mechanism of toilet paper under anaerobic conditions. The authors found 100% of removal after 8 days at 30°C of temperature, confirming that the cellulose degradation is indeed a slow process. On the other hand, the presence of cellulose in activated sludge and digested sludge calls for more in-depth studies on the conversion processes of these fibres (Ruiken et al., 2014; Rusten et al., 2006). When properly separated and refined, the cellulose can be used as raw material to make paper products or adhesion binders for asphalts (STOWA report 2012; Godow et al., 2013). In

70 addition, cellulose can be used to produce valuable chemicals or biofuels, such as short-chain 71 fatty acids (SCFAs), poly lactic acid, bioethanol (Van der Hoek et al., 2015; Honda et al., 2002). 72 The recovery of cellulosic primary sludge (CPS) in a water resource recovery facitity (WRRF) 73 can be performed by fine-mesh sieves (<500 µm) and the resulting primary sludge where the 74 fraction cellulose achieves 79% of the total mass and 84% of the organic mass (STOWA report 75 2012; Ruiken et al., 2014; Rusten et al., 2006). Currently, only few studies have investigated 76 the best pathways to valorise the CPS (Ruiken et al., 2014; Honda et al., 2002; Ghasimi et al., 77 2016), while recovery of propionate-rich SCFAs have never been studied within a wider 78 biorefinery concept. In this regard, carbon upgrading to SCFAs, mainly acetate, propionate and 79 butyrate, etc, is a cost-effective strategy to produce intermediates which can be processed to (bio)products with higher potential market value than methane (CH<sub>4</sub>) from biogas 80 81 (Kleerebezem et al., 2015; Holtzapple et al., 2009). Moreover, recent studies suggest that 82 propionate can best enhance the biological phosphorus removal (BPR) processes in biological 83 nutrients removal systems (Chen et al., 2004; Oehmen et al., 2006). On the other hand, higher 84 propionate/acetate ratio promotes the selective growth of polyphosphate accumulating organisms compared to the glycogen accumulating organisms in enhanced biological 85 phosphorus removal systems (Oehmen et al., 2006). In addition, SCFAs with higher 86 87 propionate/acetate ratio promote the production of co-polymers characterised by less stiffness 88 and brittleness, higher flexibility (higher elongation to break), and higher tensile strength and 89 toughness (Laycock et al., 2014; Frison et al., 2015). Consequently, the selective production of 90 SCFA from sewage sludge under optimized acidogenic fermenting conditions have become an 91 emerging research field that enables wastewater-based biorefineries (Lee et al., 2014; Basset et 92 al., 2016).

Other authors (Zurzolo et al., 2016) studied the SCFAs production from the fermentation of 93 94 conventional primary and secondary sludge, while the potential of SCFAs production and 95 nutrients recovery (e.g. struvite) from CPS is still unknown. In this regard, the rates and the 96 yields of CPS fermentation could be influenced by key operating parameters such as pH, sludge 97 retention time (SRT) and temperature. pH affects the hydrolysis and the subsequent 98 acidification step during the fermentation. Alkaline conditions (pH >9) promotes the 99 fermentation of primary sludge and inhibits the methanogenic activity, to achieve higher 100 conversion to SCFAs (Wu et al., 2010). On the other hand, it is reported that the optimum pH 101 range for the hydrolysis and acidogenesis of cellulose is between 5.6 and 7.3 (Hu et al., 2004). 102 The fermentation of sewage sludge is usually performed at mesophilic (30-40 °C) or moderate 103 thermophilic (50-55 °C) conditions. Thermophilic condition may increase the substrate 104 degradation rate, but this is unfavourable for the both energy balance and for the process 105 stability. Therefore, mesophilic conditions are still recommended to achieve a robust and stable 106 sludge fermentation (Yu et al., 2002). However, the combined effect of operating temperature, 107 pH and SRT on the production of SCFAs from CPS fermentation is still unknow. In addition, 108 the fermentation of sewage sludge involves relevant orthophosphate release in the liquid phase, 109 which can be effectively recovered through struvite crystallization (Tong et al., 2009; Zhang et 110 al., 2009).

In this paper, the optimization of the SCFAs production was explored through the fermentation of CPS at different temperature (37, 55 and 70 °C) and initial sludge pH (uncontrolled pH, 8, 9 and 10) to maximize: (1) the production of propionate (Pr) in SCFAs, (2) the recovery of phosphorus (PO<sub>4</sub>-P) as struvite from the fermentation liquid; (3) the final biogas production from the fermentation solid. The resulting optimized parameters were then used to set-up and study the long-term operation of a Sequencing Batch Fermentation Reactor (SBFR), that provided the results for the forthcoming scale-up for WRRFs. Finally, the market added value of the recovered materials from CPS fermentation was estimated based on the experimental results. Based on the rates and mass flows obtained in the SBFR, the scale-up of this scheme will be integrated in the real wastewater treatment plant of Carbonera (Treviso) within the European Horizon2020 Innovation Action "SMART-Plant".

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### 124 **2. Material and Methods**

### 125 2.1 Source and physicochemical characteristics of the CPS

126 Real and raw thickened PS was collected once per week and for 3 months during spring season 127 and dry weather from the municipal WWTP of Verona (Veneto Region, Italy). Primary sludge 128 was settled in primary clarifiers after the removal of grit, sand particles and oil from the influent 129 wastewater. Then, the PS was thickened up to around 4% total solids (TS) using gravity belt 130 thickening (Klein Technical Solutions, Germany). After sampling the PS was mixed and 131 homogenized with toilet paper obtaining a total concentration of cellulose around 70-75% in 132 agreement with other literature studies (Ruiken et al., 2014). Before the preparation of CPS, the 133 toilet paper was soaked in wastewater for 4 h to achieve a CPS with similar characteristics to 134 those found in WWTPs. The main characteristics of CPS were as follows: total solids (TS) 56.0 135  $\pm$  17.2 g TS/L, volatile solids (TVS) 48.2  $\pm$  14.0 g TVS/L, pH of 6.3  $\pm$  0.1, total chemical 136 oxygen demand (tCOD) and soluble chemical oxygen demand (sCOD) concentrations of 949 137  $\pm$  156 mg COD/g TVS and 48  $\pm$  26 mg COD/g TVS, respectively, ammonium concentration (NH<sub>4</sub>-N)  $1.7 \pm 0.4$  mg N/g TVS and ortophosphate concentration  $0.5 \pm 0.1$  mg PO<sub>4</sub>-P /g TVS. 138 139 The concentration of total SCFAs detected in the CPS was  $26.2 \pm 9.8$  mg COD/g TVS.

### 141 2.2 Operating temperatures and pH of the CPS fermentation

142 In order to acquire full understanding of inputs and outputs being investigated, the complete 143 matrix of batch fermentation experiments of CPS was performed at: (a) different temperatures, 144  $37 \pm 1$  °C (mesophilic),  $55 \pm 1$  °C (thermophilic) and  $70 \pm 1$  °C (hyperthermophilic); (b) different 145 initial value of pH (8, 9 and 10) to investigate the effect of these operating parameters on the 146 SCFAs production and composition. The batch fermentation experiments were performed in 147 duplicates by using 1 L glass bottles, with a working volume of 0.6 L. The initial pH was 148 adjusted using sodium hydroxide (NaOH). In addition, a batch experiment with uncontrolled 149 pH was used as the reference fermenting conditions. The batch assays were kept at controlled 150 temperature for 16 days, while pH was not controlled during the fermentation tests. The reactors 151 were sealed with rubber stopper and opened for only approximately 1 min for sampling and to 152 measure the pH using a pH sensor (Eutech pH 700). The samples were centrifuged, filtered 153 through membrane filters (Whatman, 0.45 µm), then analysed for PO<sub>4</sub>-P, NH<sub>4</sub>-N, sCOD 154 concentrations and SCFAs concentration and composition. Total solids (TS) and volatile solids 155 (TVS) were determined at the beginning and end of the fermentation experiments.

The actual production of SCFAs was always calculated subtracting the initial SCFAs concentration of the raw sludge. The yield of SCFAs production was expressed as mg COD/L of SCFA per g TVS/L in the feed sludge (mg COD/g TVS<sub>fed</sub>). Similarly, the released PO<sub>4</sub>-P was determined as mg PO<sub>4</sub>-P/gTVS<sub>fed</sub>.

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## 161 2.3 Optimization of the CPS fermentation to enhance propionate production

Based on the full set of batch experiments the response surface methodology (RSM) wasapplied to further advance the propionate production based on the fermenting temperature and

initial pH value. The regression model used is shown in Equation 1 and the target responses were the production of SCFAs ( $Y_{SCFAs}$ ) and the content of propionate over those SCFAs (%*Pr*).

166 
$$Y(z) = \beta_0 + \beta_1 x_1 + \beta_2 x_2 + \beta_{1,2} x_1 x_2 + \beta_{1,1} x_1^2 + \beta_{2,2} x_2^2$$
(Eq. 1)

167 where Y(z) is the response variable (i.e.  $Y_{SCFAs}$  (mg COD/g TVS<sub>fed</sub>) or %*Pr* (gCOD/gCOD x 168 100),  $x_1$  is the initial pH and  $x_2$  is the temperature (°C),  $\beta_0$  is the model constant,  $\beta_1$  and  $\beta_2$  are 169 linear coefficients,  $\beta_{1,2}$  is the cross-product coefficient, and  $\beta_{1,1}$  and  $\beta_{2,2}$  are the quadratic 170 coefficients.

Finally, a statistical analysis was carried out by means of the analysis of variance (ANOVA) to
test the significance of predicted and experimental results, under a significance level of 0.05
(p). The regression model and the statistical analyses were performed using the software R 3.2.3
(The R Foundation for Statistical Computing).

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## 176 2.4 Sequencing Batch Fermentation Reactor

177 A sequencing batch fermentation reactor (SBFR) with a reaction volume of 4 L was operated 178 at 37±1 °C by a thermostatic bath, while the HRT was kept constant at 4 days by the daily 179 exchange of 25% of the reactor volume between fermented and fresh CPS. The SBFR was 180 equipped with a blade stirrer installed in the bottom. In this work, two representative 181 experimental periods were carried out (period 1 and 2) where the steady-state conditions were 182 considered when the calculated Relative Standard Deviation (RSD) of the propionate 183 production was below 10% for at least 3 times the HRT of the SBFR (Ghasem et al., 2008). 184 The Equation 2 reports how the RSD was calculated:

186 where  $\sigma$  (Pr production) is the standard deviation of the propionate production found during the 187 period, while  $\mu$  (Pr production) is the average of the propionate production found during the 188 period.

The period 1 (0-18 days) was operated to determine the yield of Pr production without any adjustment of pH in the fed CPS. In the period 2 (19-96 days), every day the pH of the fresh CPS was adjusted to 8 before the feeding of the reactor. Steady-state conditions were achieved between days 4 and 16 for period 1, while during days 36-96 for period 2.

193 Samples were periodically taken from the effluent of the SBFR and analysed for PO<sub>4</sub>-P, NH<sub>4</sub>-194 N, SCFAs, chemical oxygen demand (total COD, soluble COD), total solids (TS), volatile 195 solids (TVS) and pH. The composition of the SCFAs (i.e. acetate, propionate) were also investigated to determine the propionate/acetate ratio (g CODpropionate/g CODacetate) as 196 197 monitoring parameter during the experiment. Propionate/acetate between 0.25-0.75 g 198 COD<sub>propionate</sub>/g COD<sub>acetate</sub> is considered the optimal biological phosphorus removal processes 199 (Broughton et al., 2008; Yuan et al., 2012). On the other hand, higher COD<sub>propionate</sub>/g COD<sub>acetate</sub> 200 ratios promote the production of polyhydroxyvalerate (PHV) instead of polyhydroxybutyrate 201 (PHB) improving the mechanical and physical properties of the biologically recovered PHAs 202 (Jiang and Chen, 2009).

Twice per week the "cellulosic materials" in the influent and effluent from the SBRF were quantified as the volatile fraction at 550°C of the solids recovered and washed after the sieving at mesh 54  $\mu$ m. During the periods 1 and 2, samples of biomass from the SBFR were taken and characterized by the FISH quantification of the *Propionibacterium acidopropionici* using the Apr820 and DAPI probes, following the methodology described Nielsen et al. (2009). Forty images of each sample were taken using a fluorescence microscope (Leica DM2500) and then analysed with the Image J software. 210 During the steady-state conditions of the SBFR, struvite recovery tests were performed from 211 the CPS fermentation liquid. The fermentation liquid from the supernatant was obtained after 212 the centrifugation at 4000 rpm for 10 minutes of the effluent from the SBFR. The experiments 213 started with the addition of 5 g/L of seed struvite crystals and magnesium hydroxide (Mg(OH)<sub>2</sub>) according to a  $PO_4^{3-}$ : Mg<sup>2+</sup> molar ratio of 1:1.5, and adjusting the initial pH at 8.5 with NaOH 214 215 (0.1 M). Samples were collected at 5 min, 10 min, 15 min, 30 min and 60 min, filtered through 216 cellulose membrane filters (Munketll Ahlstrom) and analysed to determine their PO<sub>4</sub>-P 217 concentration. After the experiments, the precipitated solids were washed with distilled water 218 to remove impurities and soluble salts. The recovered solids were dried at 45 °C for 24 h to 219 avoid thermal decomposition (Bhuiyan et al., 2008). The crystals produced were analysed 220 according to Fattah et al. (2012) and the molar ratio between nitrogen and phosphorus was used 221 to confirm the struvite formation.

222

# 223 2.5 Biochemical methane potential tests

The BMP test were also investigated. The BMP test were carried out following the procedure defined by Angelidaki et al., (2009) at 37 °C, while parallel tests were carried out using raw primary sludge to compare biogas production and composition from CPS and PS BMP tests. More details of this method are reported in Supporting Information.

228

229 2.6 Analytical methods

230 Soluble COD, TSS and TVS were measured according to Standards Methods (APHA-AWWA-

231 WPCF, 2012). NH<sub>4</sub>-N concentration was measured by an ion selective electrode (Orion 9512).

232 The concentration of SCFAs was determined by gas chromatography (Dionex ICS-1100 with

AS23 column). PO<sub>4</sub>-P concentration was measured by ion chromatography (Dionex ICS-900
with AS14 column) and calibrated using a combined five anion standard (Thermo Scientific<sup>™</sup>
Dionex<sup>™</sup> Ion Standards).

236

### 237 **3. Results and Discussion**

238 3.1 SCFAs production and composition from the batch fermentation experiments

No relevant lag-phase was observed before the production of SCFAs started. The concentration
increased gradually until the peak and plateau values were reached (between the 9th and 13th

241 day) at 37 and 55 °C (Figures S1, Supplementary Material). On the other hand, the SCFAs

242 production at 70 °C increased up to day 2 and then decreased (Figure S1 in the Supplementary

243 Material). The highest production yield of SCFA of 340.4 mg COD/g TVS<sub>fed</sub> was observed at

244 37 °C (Figure 1), while the lowest were 155.4 and 46.1 mg COD/g TVS<sub>fed</sub>

observed at 55°C and 70°C respectively. Therefore, the temperature had a major effect on the production of SCFAs. In particular, the effect the fermentation temperature had on the hydrolyses and the acidogenic process can clearly be detected from the pH profile over the batch experiments (SI Figure S2), since it was adjusted only at the beginning of the experiments. Moreover, higher variation of pH resulted in higher SCFAs production, thus pH seemed to act as a surrogate parameter for monitoring the fermentation process.

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- 252

# (Figure 1)

253

Although the increase of pH can positevely influence the fermentation efficiencies (Wu et al.,

255 2010), in this study the highest SCFAs production were observed at pH 8, while at pH 9 and 10

the production of SCFAs were much lower at 37 °C (Figure 1). On the other hand, under thermophilic and hyper-thermophilic conditions, fermentation pH was almost stable and the production of SCFAs was comparable notwithstanding the initial pH condition.

259 Acetate and propionate were the most relevant SCFAs in the fermentation liquid for all the 260 fermentation experiments at 37 °C (acetate 55-80%, propionate 12-33%), with minor 261 concentration of mainly butyrate and n-valerate (around 6-9% and 3-4% respectively) (Figures 262 S3(a), S3(b), S3(c) in the Supporting Information). Propionate was mostly produced under mesophilic condition in a range of percentage between 25-33%, while under thermophilic and 263 264 hyper thermophilic propionate contents below 20% were observed. The fermentation 265 temperature increase led to propionate/acetate ratio decrease: the higher ratio of 0.6 266 gCOD/gCOD was observed at 37 °C and initial pH of 8.

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# 268 3.2 Optimal key operating parameters for CPS fermentation

269 The 3D rensponse surface methodology (RSM) was applied to optimize both the total SCFAs 270 production and the propionate percentage based on the initial pH and the fermentation 271 temperature. The results of the regression model indicated that the increase of the fermentating 272 temperature has a detrimental effect on total SCFAs and Pr productions than initial pH fixed 273 value (SI Figure S2a and 2b). Figure 2a and 2b shows that SCFAs and Pr percentage 274 productions decrease with the increase of fermentation temperature, obtaining higher SCFAs 275 productions from CPS fermentation at 37 °C. So, the response surface plot indicates that the 276 most favourable operating conditions to maximize SCFAs production were 37 °C of 277 temperature and initial pH of 8 (Figure 2a), while the highest percentage of propionate can be 278 obtained at lower initial pH (7.5 < pH < 8.0) and a temperature of 37 °C. Under these operating

279	conditions the production of SCFAs and propionate percentage were 281.5 mg COD/g TVS
280	and 30.3%, respectively (Figure 2a and 2b).
281	
282	(Figure 2a and 2b)
283	
284	The statistical significance of total SCFAs production and propionate percentage models were
285	evaluated by ANOVA (Table 1).
286	
287	(Table 1)
288	
289	The combination of low p value and high $R^2$ indicated that the model explains a lot of variation
290	within the data and is significant. The models of F-value showed a low p-value (<0.016), which
291	implied that both models were significantly affected by temperature and initial pH, and able to
292	estimate total SCFAs production and %Pr. The predicted high $R^2$ (between 0.7 and 0.9)
293	indicated both models were sufficiently high to show the significance of the fit of the models
294	(Table 1). Table 2 reported the coefficients of the quadratic models for the responses of the
295	$Y_{SCFA}$ and %Pr. For %Pr model, pH, temp and the interaction effect $\beta_{1,2}$ and $\beta_{2,2}$ were not
296	significant (p>0.05), while only $\beta_{1,1}$ was found significant (p<0.05). For SCFAs production
297	model, all the terms were found not significant (p>0.05) meaning that the effect on response is
298	considerable.

Figure 3 shows the comparison between the predicted and experimental values for the total SCFAs and the percent of propionate production. The experimental results are consistent with the regression model ( $R^2 > 0.86$ ) for both key parameters.

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- 304

#### (Figure 3)

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306 3.3 Long-term SCFAs production and phosphorus recovery in the SBFR

During the period 1 (0-18 days), the sCOD varied in the range 10-12 gCOD/L and lower yields
of Pr production were observed (around 60 mgCOD<sub>propionate</sub>/gTVS<sub>fed</sub>·d). In period 2, the sCOD
higher and more stable in the range 14-18 gCOD/L (see Figure S4, Supporting Information).
The increase of the sCOD was a result of the higher degradation of the cellulosic materials
observed in period 2 (Table 3).

- 312
- 313

### (Table 3)

314 The effect of the influent pH was clearly observed by the productivity of the SCFAs, which 315 increased from 162.4±12.8 mg COD/g TVS<sub>fed</sub> d (period 1) to 253.8±26.1 mg COD/g TVS<sub>fed</sub> d 316 (period 2). Moreover, the increase of the pH influent had effect on the propionate production, 317 which gradually increased and reached a stable production of 100-120 mg COD/g TVS<sub>fed</sub> d during days 36-96 of period 2 (Figure 4). The latter is higher than the batch experiments, 318 319 probably due to the speciation of the microbial community accomplished in the SBFR. In fact, 320 the percentage of propionate to total SCFAs was up to 46%. This corresponded to 321 propionate/acetate ratio of 0.9 gCOD/gCOD, which was higher than the period 1 (0.6 322 gCOD/gCOD).

324	Many authors reported that the fermentation of cellulosic compounds at relatively low pH, lactic
325	acid could be produced (Abdel-Rahman et al., 2013). Indeed, despite the initial pH of the CPS
326	in period 2 (19-96) was adjusted to 8, the average pH in the SBFR dropped to $5.1\pm0.1$ due to
327	simultaneous production of the SCFAs and alkalinity consumption (Figure 4). As a
328	consequence, bacteria of the genus Propionibacterium may produce propionate from lactate as
329	the end-product of their anaerobic metabolism (Liu et al., 2012). In period 2, FISH analyses
330	(see Table S1 and Figure S6 in Supporting Information) confirmed selective growth of
331	Propionibacterium acidopropionici that were at 33.8%, more abundant than period 1 (24.5%).
332	So, the beneficial speciation of the microbial community in the SBFR seems to be related with
333	the higher solubilization of the cellulosic materials achieved in period 2.
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334

335

(Figure 4)

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337 Furthermore, NH<sub>4</sub>-N and PO<sub>4</sub>-P were released in the CPS fermentation liquid of the SBFR as 338 much as  $1.6 \pm 0.5$  mg P/g TVS<sub>fed</sub>·d and  $6.1 \pm 1.4$  mg N/g TVS<sub>fed</sub>. d. As a consequence, the 339 average concentrations of PO<sub>4</sub>-P and NH<sub>4</sub>-N in sludge fermentation liquid during steady conditions were 130  $\pm$  23 mg P/L and 430  $\pm$  29 mg N/L, respectively. Due to the high P and N 340 341 content, the recovery of the released PO<sub>4</sub>-P from CPS fermentation by struvite crystallization 342 was examined and the average efficiency of phosphorus recovery was 88%.

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344 3.4 Specific per capita recovery of valuable resources from CPS and preliminary economics

345 The selective production of mixture of SCFAs through acidogenic fermentation was considered 346 best available carbon source to enhance the nutrients removal in the mainstream or in the 347 sidestream (Frison et al., 2013, 2016). Recently, Longo et., 2017 considered this practice an 348 economic and environmentally friendly solution to reduce energy and chemical consumption 349 for the removal of nitrogen and phosphorus. Moreover, a number of Horizon2020 Innovation Actions (such as SMART-Plant (www.smart-plant.eu) or INCOVER (www.incover.eu) were 350 351 evaluated and funded to demonstrate the technical, economic and environmental long-term 352 viability of further (bio)conversion of the SCFAs to biopolymers, such as PHAs, that could 353 enable the recovery of high added value products by minor integration of existing WWTPs. 354 Although the best technical and economical evaluation should always be referred to single 355 WWTP, the specific economic advantages of alternative for CPS processing in comparison with 356 the only biogas production is estimated below.

357 Every year, around 36-43 kg of COD are discharged in municipal wastewater by individuals 358 (capita) (adapted Metcalf and Eddy, 2014). The observed average removal efficiency of COD 359 by the sieving municipal wastewater is between 10-60% (Ruiken et, 2014) and 12-13 kg 360 COD/capita·year could be recovered as suspended solids from municipal wastewater. In this 361 study the BMP test (SI Figure S5) showed that CPS may produce up to 0.30-0.34 m<sup>3</sup> CH<sub>4</sub>/kg COD<sub>fed</sub> equivalent to 3.7-4.5 m<sup>3</sup> CH<sub>4</sub>/capita·year (Figure 5a), which is in agreement with other 362 363 studies (Ghasimi et al., 2016). Therefore, the convertible COD to CH<sub>4</sub> by anaerobic digestion 364 in a current WWTP (Table 4) would be around 11 kg COD/capita·year, which represents around 365 25% of the total COD influent in a WWTP. In a CPS-based biorefinery scenario (Table 4), 366 considering the observed SCFAs production rate, around 3.0 kg COD<sub>SCFA</sub>/capita·year could be 367 produced by the fermentation of CPS, where acetate and propionate represent 1.30 and 1.17 kg 368 COD<sub>SCFA</sub>/capita·year, respectively. However, around 2.7-3.3 m<sup>3</sup>CH<sub>4</sub>/capita·year of residual 369 CH<sub>4</sub> could be further produced by the anaerobic digestion of CPS after fermentation (Figure 370 5b) Moreover, during the fermentation of CPS, nutrients are released and 88% of the

371 phosphorus could be recovered in the form of struvite, that amounts to 0.07-0.15 kg
372 struvite/capita·year.

373 Assuming CH<sub>4</sub> a market price of 0.11 €/m<sup>3</sup> (Energy Information Administration, 2017), the best 374 valorisation of CH<sub>4</sub> from CPS can be as high as 0.46 €/capita year. On the other hand, better 375 value can derive from valorising CPS first to the suitable mix of SCFAs (mainly acetate and 376 propionate) and struvite from the fermentation liquid, while CH<sub>4</sub> can be recovered after 377 digestion of fermentation solids. Although the market price of the recovered materials is very 378 volatile and often unknown because of the variable purity and quality, according to a recent 379 review, acetate and propionate price can be as high as 0.45 and 1.01 €/kg respectively (Global 380 Chemical Price, 2017; ICIS, 2017), while struvite can be sold up to 0.76 €/kg (Molinos-Senante 381 et al., 2011; P-REX report, 2014). Therefore, the SCFAs and struvite route before the bio-382 methanization can increase the market value potential of CPS up to 1.55-1.95 €/capita·year 383 (Table 4).

- 384 (Table 4)
- 385

(Figure 5)

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387 4. Conclusions

This paper studied the maximum potential recovery of SCFAs, particularly propionate, struvite and CH<sub>4</sub> from the CPS. Based on the results of the RSM, the optimal production of propionate is obtained by the fermentation of CPS at mesophilic conditions (37 °C) and at initial pH between 7.5-8. By the long-term operation of a SBFR the observed production of propionate in the fermentation liquid was 100-120 mg COD/g TVS<sub>fed</sub>·d, with a propionate/acetate ratio of 0.9 g COD/g COD. Best performances in the SBFR may be attributed to the observed enhanced growth of *Propionibacterium acidopropionici*. At the same time, 88% of the phosphate released in the fermentation liquid can be recovered as struvite. From a techno-economic point of view,
the integration of the wastewater dynamic sieving to recover CPS in a WWTP may make
existing units (e.g., gravity sludge thickener) redundant and available to be revamped to
controlled fermenter to recover optimal mixture of SCFAs. In addition, the recovery of SCFAs
and struvite before the bio-methanization can increase the market value potential of CPS up to
1.55-1.95 €/capita·year.

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#### 409 **6. References**

Abdel-Rahman, M. A., Tashiro, Y., Sonomoto, K. (2013). Recent advances in lactic acid
production by microbial fermentation processes. Biotechnology advances, 31(6), 877-902.

Angelidaki, I., Alves, M., Bolzonella, D., Borzacconi, L., Campos, J.L., Guwy, A.J.,
Kalyuzhnyi, S., Jenicek, van Lier, J.B., 2009. Defining the biomethane potential (BMP) of solid
organic wastes and energy crops: a proposed protocol for batch assays. Water Science and
Technology 59(5), 927-934.

416 APHA-AWWA-WPCF, 2012. Standard Methods for the Examination of Water and
417 Wastewater. American Public Health Association/American Water WorksAssociation/Water
418 Environment Federation, 22nd ed. Washington D.C, USA.

- Basset, N., Katsou, E., Frison, N., Malamis, S., Dosta, J., Fatone, F. (2016). Integrating the
  selection of PHA storing biomass and nitrogen removal via nitrite in the main wastewater
  treatment line. Bioresource technology, 200, 820-829.
- 422 Bhuiyan, M. I. H., Mavinic, D. S., Koch, F. A. (2008). Phosphorus recovery from wastewater
- 423 through struvite formation in fluidized bed reactors: a sustainable approach. Water Science and
- 424 Technology, 57(2), 175-181.
- 425 Broughton, A., Pratt, S., Shilton, A. 2008. Enhanced biological phosphorus removal for
- 426 highstrength wastewater with a low rbCOD:P ratio. Bioresource Technology, 99(5), 1236-41.
- 427 Chen, Y., Randall, A.A., McCue, T., 2004. The efficiency of enhanced biological phosphorus
  428 removal from real wastewater affected by different ratios of acetic to propionic acid. Water
  429 Research 38 (1), 27-36.
- Fattah, K.P., Mavinic, D.S., Koch, F. a., 2012. Influence of Process Parameters on the
  Characteristics of Struvite Pellets. J. Environ. Eng. 138, 466. doi:10.1061/(ASCE)EE.19437870.0000576
- Genty A., Kowalska M., Wolf O., 2013. Development of EU Ecolabel and GPP Criteria for
  Flushing Toilets and Urinals Technical Report. European Commission
- 435 Frison, N., Di Fabio, S., Cavinato, C., Pavan, P., Fatone, F. 2013. Best available carbon sources
- 436 to enhance the via-nitrite biological nutrients removal from supernatants of anaerobic co-
- 437 digestion. Chemical Engineering Journal 215-216, 15-22
- Frison, N., Katsou, E., Malamis, S., Oehmen, A., Fatone, F., 2015. Development of a novel
  process integrating the treatment of sludge reject water and the production of 407
  polyhydroxyalkanoates (PHAs). Environmental Science and Technology 49(18), 10877-10885.

- 441 Frison, N., Katsou, E., Malamis, S., Fatone, F. (2016). A novel scheme for denitrifying
- 442 biological phosphorus removal via nitrite from nutrient-rich anaerobic effluents in a short-cut
- sequencing batch reactor. Journal of chemical technology and biotechnology, 91(1), 190-197.
- 444 Gadow, S.I., Jiang, H., Watanabe, R., Li, Y-Y., 2013. Effect of temperature and temperature
- shock on the stability of continuous cellulosic-hydrogen fermentation. Bioresource Technology
- 446 Ghasem, N., Henda, R., 2008 Principles of Chemical Engineering Processes: Material and
- 447 Energy Balances, Second Edition. CRC Press Book.
- 448 Ghasimi, D.S.M., de Kreuk, M., Maeng, S.K., Zandvoort, M.H., van Lier, J.B., 2016. High-rate
- thermophilic bio-methanation of the fine sieved fraction from Dutch municipal raw sewage:Cost-effective potentials for on-site energy recovery. Applied Energy 569-582.
- 451 Global Chemical Price (IGP). http://www.globalchemicalprice.com/chemical-market-452 reports/acetic-acid-weekly-report-17-june-2017
- Holtzapple, M. T., Granda, C. B., 2009. Carboxylate platform: the MixAlco process part 1:
  Comparison of three biomass conversion platforms. Applied Biochemistry and Biotechnology
  156(1), 95-106.
- 456 Hart, N. R. de, E. D. Bluemink, A.J. Geilvoet and J.F. Kramer, 2014. Bioplastic uit slib.
  457 Verkenning naar
- 458 PHA-productie uit zuiveringsslib (Grondstoffenfabriek). STOWA report 2014-10. 88 pp.
- 459 Honda, S., Miyata, N., Iwahori, K., 2002. Recovery of biomass cellulose from waste sewage
- 460 sludge. Journal of Material Cycles and Waste Management 4, 46-50.
- 461 Hu, Z-H., Wang, G., Yu, H-Q., 2004. Anaerobic degradation of cellulose by rumen
  462 microorganisms at various pH values. Biochemical Engineering Journal 21, 59-62.

463 ICIS: <u>https://www.icis.com/resources/news/2007/10/01/9065938/chemical-profile-propionic-</u>
464 <u>acid/</u>

- Jiang, Y. and Y. Chen, 2009. "The effects of the ratio of propionate to acetate on the
  transformation and composition of polyhydroxyalkanoates with enriched cultures of glycogen-
- 467 accumulating organisms." Environmental Technology 30(3): 241-249.
- Kleerebezem, R., Joosse, B., Rozendaal, R., Van Loosdrecht, M. C., 2015. Anaerobic digestion
  without biogas? Reviews in Environmental Science and Bio/Technology, 2015.
- 470 Laycock, B., Halley, P., Pratt, S., Werker, A., Lant, P., 2014. The chemomechanical properties
- 471 of microbial polyhydroxyalkanoates. Progress in Polymer Science 39(2), 397-442.
- 472 Lee, W.S., Chua, A.S.M., Yeoh, H.K., Ngoh, G.C., 2014. A review of the production and
  473 applications of waste-derived volatile fatty acids. Chemical Engineering Journal 235, 83-89.
- 474 Liu, L., Zhu, Y., Li, J., Wang, M., Lee, P., Du, G., Chen, J. (2012). Microbial production of
- propionic acid from propionibacteria: current state, challenges and perspectives. Critical
  reviews in biotechnology, 32(4), 374-381.
- 477 Longo, S., Frison, N., Renzi, D., Fatone, F., Hospido, A. (2017). Is SCENA a good approach
- 478 for side-stream integrated treatment from an environmental and economic point of view?. Water
- 479 Research, 125, 478-489.
- 480 Metcalf and Eddy (2014) Wastewater Engineering Treatment and Resource Recovery, Fifth
- 481 Edition. McGraw-Hill Education, New York.
- 482 Molinos-Senante, M., Hernández-Sancho, F., Sala-Garrido, R., Garrido-Baserba, M. (2011).
- 483 Economic feasibility study for phosphorus recovery processes. Ambio, 40(4), 408-416.

484 Nielsen, P. H., Daims, H., Lemmer, H., Arslan-Alaton, I., Olmez-Hanci, T. (Eds.). (2009). FISH
485 handbook for biological wastewater treatment. Iwa publishing.

Noike, T., Endo, G., Chang, J.E., Yaguchi, J.I., Matsumoto J.I., 1985. Characteristics of
carbohydrate degradation and the rate-limiting step in anaerobic-digestion. Biotechnology
Bioengineering 27, 1482-1489.

489 Oehmen, A., Saunders, A.M., Vives, M.T., Yuan, Z., Keller, J., 2006. Competition between
490 polyphosphate and glycogen accumulating organisms in enhanced biological phosphorus
491 removal systems with acetate and propionate as carbon sources. Journal of Biotechnology 123
492 (1), 22–32.

493 P-REX-Sustainable sewage sludge management fostering phosphorus recovery and energy 494 efficiency. Report market for phosphorus recycling products on (http://p-495 rex.eu/uploads/media/D11 1 Market Overview and Flows.pdf) Project supported by the 496 European Commission within the Seventh Framework Programme Grant agreement No. 308645 497

Ruiken, C.J., Breuer., G., Klaversma, E., Santiago, T., van Loosdrecht, M.C.M., 2014. Sieving
wastewater - Cellulose recovery, economic and energy evaluation. Water Research 47, 43-48.

Rusten, B., Odegaard, H., 2006. Evaluation and testing of fine mesh sieve technologies for
primary treatment of municipal wastewater. Water Science and Technology 54(10), 31-38.

502 STOWA report, 2012. Verkenning naar mogelijkheden Voor Verwaarding Van zeefgoed.

503 Tong, J., Chen, Y., 2009. Recovery of nitrogen and phosphorus from alkaline fermentation

504 liquid of waste activated sludge and application of the fermentation liquid to promote biological

505 municipal wastewater treatment. Water Research 43(12), 2969-2976.

506U.S.EnergyInformationAdministration(EIA):507https://www.eia.gov/dnav/ng/hist/n3035us3m.htm

Van der Hoek, J.P., Struker, A., de Danschutter, J.E.M., 2015. Amsterdam as a sustainable
European metropolis: integration of water, energy and material flows. Urban Water Journal
14(1), 61-68.

- 511 Verachtert, H., Ramasamy, K., Meyers, M., Bever, J., 1982. Investigation on cellulose
  512 degradation in activated sludge plants. Journal of Applied Bacteriology 52, 185-190.
- 513 Wu, H., Gao, J., Yang, D., Zhou, Q., Liu, W., 2010. Alkaline fermentation of primary sludge
  514 for short-chain fatty acids accumulation and mechanism. Chemical Engineering Journal 160,

515 1-7.

- Yu, H-Q., Fang, H.H.P., Gu, G-W., 2002. Comparative performance of mesophilic and
  Thermophilic acidogenic upflow reactors. Process Biochemistry 38, 447-454.
- Yuan, Z., Pratt, S., Batstone, D.J. 2012. Phosphorus recovery from wastewater through
  microbial processes. Current Opinion Biotechnology, 23(6), 878-83.
- Zhang, C., Chen, Y., 2009. Simultaneous Nitrogen and Phosphorus Recovery from SludgeFermentation Liquid Mixture and Application of the Fermentation Liquid To Enhance
  Municipal Wastewater Biological Nutrient Removal. Environmental Science and Technology
  43(16), 6164-6170.
- 524 Zurzolo F., Yuan Q., Oleszkiewicz J. A., 2016. Increase of Soluble Phosphorus and Volatile
  525 Fatty Acids During Co-fermentation of Wastewater Sludge. Waste and Biomass Valorization
  526 7, 317-324.
- 527