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Force majeure: will climate change affect our ability to attain Good Environmental Status for marine biodiversity?

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Abstract

The EU Marine Strategy Framework Directive (MSFD) requires that Good Environmental Status (GEnS), is achieved for European seas by 2020. These may deviate from GEnS, its 11 Descriptors, targets and baselines, due to *endogenic managed pressures* (from activities within an area) and externally due to *exogenic unmanaged pressures* (e.g. climate change). Conceptual models detail the likely or perceived changes expected on marine biodiversity and GEnS Descriptors in the light of climate change. We emphasise that marine management has to accommodate '*shifting baselines*' caused by climate change particularly during GEnS monitoring, assessment and management and '*unbounded boundaries*' given the migration and dispersal of highly-mobile species. We suggest climate change may prevent GEnS being met, but Member States may rebut legal challenges by claiming that this is outside its control, *force majeure* or due to 'natural causes' (Article 14 of the MSFD). The analysis is relevant to management of other global seas.

Keywords: Marine Strategy Framework Directive, European, legislation, habitat management, climate change, Oceans Acts

Introduction

Integrated marine management, conservation and protection must maintain and protect the natural structure and functioning while at the same time ensure that the seas deliver the benefits required by society (Elliott, 2011). This should accommodate many local activities and pressures, those emanating from inside the sea area being managed, and wider pressures, such as global climate change, emanating from outside the area. The European Marine Strategy Framework Directive (MSFD, 2008/56/EC; European Commission 2008) aims to ensure that through the measures and monitoring performed by the EU Member States, that Good Environmental Status (GEnS) will be achieved for European seas by 2020 (e.g. Borja et al., 2013a) (NB, the acronym GEnS is used here following Mee et al., 2008 and Borja et al, 2010a, to be differentiated from Good Ecological Status (GEcS) à la the EU Water Framework Directive, WFD 2000/60/EC; European Commission 2000). GEnS is defined according to a set of 11 Descriptors (as different components of environmental status, Tables S1 and S2 (Supplementary Material)) and their component 29 Criteria (Table S1). The Descriptors and Criteria will then be deemed to have been met according to a set of 56 indicators and whether these achieve a set of targets (i.e. quality objectives, or reference conditions), selected by the European Commission (2010) (see also Borja et al., 2013b).

Borja et al. (2013a) propose the operational definition that: 'GEnS is achieved when physicochemical (including contaminants, litter and noise) and hydrographical conditions are maintained at a level where the structuring components of the ecosystem are present and functioning, enabling the system to be resistant (ability to withstand stress) and resilient (ability to recover after a stressor) to harmful effects of human pressures/activities/impacts, where they maintain and provide the ecosystem services that deliver societal benefits in a sustainable way (i.e. that pressures associated with uses cumulatively do not hinder the ecosystem components in order to retain their natural diversity, productivity and dynamic ecological processes, and where recovery is rapid and sustained if a use ceases)'.

All regional seas, their catchments and the adjacent areas will be affected by climate change (Cheung *et al.*, 2013a, b; Poloczanska *et al.*, 2013; Stocker *et al.*, 2013, Frost, *et al.*, 2015), one of many stressors in a wider typology of marine hazards and risks (Elliott et al, 2014). Conceptual models derived here show the dominant pathways, causes and consequences of climate change on the marine system and their link to the MSFD elements used to assess environmental status and functioning. These indicate the ways in which climate change could compromise achieving GEnS for the Descriptors which are referred to throughout by their numbers: 1, Biological Diversity; 2, Non-indigenous species; 3, Exploited fish and shellfish; 4, Food webs; 5, Human-induced eutrophication; 6, Seafloor integrity; 7, Hydrographical integrity; 8, Contaminants; 9, Contaminants in seafood; 10, Litter; 11, Energy and noise. Given their interlinked nature, this review considers all Descriptors but focuses on the biodiversity

Descriptors (1, 2, 4 and 6), although it is argued that if the Descriptor 1 is satisfactory then, by definition, so will be the other Descriptors and vice versa (Borja, et al (2010a). By considering all Descriptors, the aim here is to show the predominant effect of climate change on the biodiversity ones.

The MSFD process includes a set of steps (Figure 1, inner circle, created here using the MSFD, Claussen *et al.*, 2011, and CEC 2014): defining the main vision, giving a current assessment of their seas, develop a set of indicators against the Descriptors and Criteria needed to define GEnS (by 2012) (see Borja *et al.*, 2010a, 2013a,), indicate the monitoring needed (by 2014) and a programme of measures (by 2015) and implement strategies (by 2016) to reach GEnS by 2020 (with a six-years review). Successful management then requires targets, trends (qualitative or quantitative), baseline values and thresholds, i.e. precise values of metrics or indices against which the monitoring is carried out and which then should act as triggers for the measures to be implemented (e.g. Teixeira *et al.*, 2014). The indices or metrics must be SMART, i.e. Specific, Measurable, Achievable, Realistic and Time-bounded so that monitoring can determine compliance and management measures if GEnS is not achieved (Elliott, 2011).

This review shows how climate change influences or inhibits the MSFD implementation and the ability to detect GEnS; this follows the DAPSI(W)R Risk Assessment and Risk Management framework where the causes of change (the **D**rivers, **A**ctivities and **P**ressures) lead to **S**tate changes on the natural system and then to **I**mpacts on societal **W**elfare (Atkins *et al.*, 2011; Cormier *et al.*, 2013; Elliott, 2014). Finally, adverse S and I(W), the non-achievement of GEnS, then require a societal **R**esponse (including monitoring and measures) aimed at ensuring GEnS is reached and thus that Member States fulfil the legal obligation of this and all EU Directives (Boyes and Elliott, 2014). Thus the legal repercussions of not meeting GEnS due to climate change are addressed especially when the spatial overlaps of the associated EU Directives require some marine areas to reach GEnS under the MSFD, Good Ecological and Chemical Status under the Water Framework Directive, and Favourable Conservation Status (FCS) for the biodiversity conservation objectives under the Wild Birds and Habitats & Species Directives (2009/147/EC; 92/43/EEC) (Frost et al, 2015).

Climate change was not explicitly included in the MSFD and indeed is mentioned only twice. Elsewhere it is mentioned as a major pressure (CEC, 2014) but unusually was given greater prominence in the proposed Directive (CEC, 2005) compared to the final MSFD. Climate change is implicit in the list of characteristics, pressures and impacts (Annex III) but, for example, ocean acidification is listed as a characteristic rather than a pressure. It is of note that in the proposed MSFD (CEC 2005), the highly variable nature of marine ecosystems and the changes over time in human activities and pressures, were cited as the reasons for having an adaptive, flexible and dynamic definition of GEnS. The wording had then changed in the final

Directive to: 'In view of the dynamic nature of marine ecosystems and their natural variability, and given that the pressures and impacts on them may vary with the evolvement of different patterns of human activity and the impact of climate change, it is essential to recognise that the determination of good environmental status may have to be adapted over time.'

Biodiversity is threatened by the spatial extent, temporal duration and severity of pressures and the repercussions differ with area (CEC 2013). Lethal and sub-lethal changes occur at cellular, individual and population levels due to hazardous inputs from physical (e.g. noise), chemical (contaminants) and biological (non-indigenous species) stressors. Overfishing affects the communities and ecosystem as does physical disturbance and loss of habitat. These are included in the MSFD as pressure-related Descriptors (No. 2, 5, 7, 8, 9, 10 and 11) and so global climate change needs to be judged against a background of these locally and regionally managed pressures (Elliott 2011, 2014).

Integrated conceptual models and the evidence-base for the ecosystem effects of climate change

The repercussions of climate change on the ability to meet GEnS and to determine whether marine ecosystems are experiencing variability or direct climate change effects are grouped here as linked conceptual models and main topics (Figures 2 to 10; sections I to VIII below). Figure 2 cross-refers to Figures 3 to 10 onto which are superimposed the numbered main MSFD Descriptors likely to be affected by climate change and its consequences. Table 1 gives examples of the main literature base for the conclusions. The diagnosis of the state of the oceans and future prognoses for the trajectories of change are detailed in the recent Inter-Governmental Panel on Climate Change reports (Pörtner and Karl, 2014; Wong and Losada, 2014).

I Altered temperature regime – species re-distribution and community response

Species distributions are changing, as temperature regime changes, in relation to their thermal tolerances, ability to adapt, extend their range or become extinct (Table 1, Figure 3). The degree to which temperature increases and its seasonal timing may vary between regions. For example, annual mean temperatures in Europe are likely to increase more than the global mean with winter temperatures in northern Europe being notably higher and summer temperatures increasing significantly in the Mediterranean region (Christensen *et al.* 2007). As such, poleward extension, southerly contraction and depth changes cannot be assumed to be uniform among species (Table 3).

The increase of southern species into northern areas, loss of northern species with migrations, increased probability of Non-indigenous Species (NIS) influx and successful colonisation if the vector of transfer is not controlled, and a colonisation pathway is available will all prevent achieving GEnS for D2 (Occhipinti-Ambroggi, 2007; Hellmann *et al.*, 2008). This depends on

the physiological tolerances and competitive abilities of the individual species and the suitability of receiving conditions.

A biogeographical shift has occurred since the mid-20th C in the NE Atlantic, e.g. calanoid copepods experienced a 1000 km northward shift (Beaugrand *et al.*, 2002; Beaugrand, 2009, Brown *et al.*, 2011) and a switch in congeneric *Calanus* species (Beaugrand, 2003; Reid *et al.*, 2003). As these species reflect climate patterns and have a key role in the North Sea food-web (Kirby and Beaugrand, 2009), achieving GEnS for Descriptor 4 will be more difficult. Similarly, a climate-driven regime shift, decline in North Sea total copepod abundance (O'Brien *et al.*, 2013), increase in phytoplankton biomass (Reid *et al.*, 1998; McQuatters-Gollop *et al.*, 2011), but decrease in diatoms and dinoflagellates (O'Brien *et al.*, 2012) have resulted in trophic mismatches (Beaugrand, 2004, Edwards *et al.*, 2002; Edwards and Richardson, 2004), again reducing achieving GEnS for Descriptor 4. However, as an example of equivocal evidence, at regional and European levels, a projected sea surface warming of 2.29 ± 0.05 °C may reduce zooplankton and phytoplankton biomasses by 11% and 6%, respectively (Chust *et al.*, 2014a, b); the resultant influence in nutrient use and bloom-forming species will affect GEnS for Descriptor 5.

Several MSFD Descriptors are influenced by two thirds of North Sea fish shifting or retracting northwards or deeper, in line with increasing sea surface temperature (SST), and replacement by southerly species (e.g. red mullet, anchovy, sardine, and John Dory) (Dulvy *et al.*, 2008). Given the warming and increasing availability of shallow water winter habitats, any indicators reliant on community structure and population dynamics and their effects on fisheries and predator-prey relationships will affect GEnS indicators for D3 and 4. For example, plaice and sole have migrated northwards 142 and 93 km since 1913 (Engelhard *et al.*, 2011). Plaice have generally moved northeast and into deeper waters (a depth change of approx. 20 m) whilst sole are more prevalent south-eastwards in shallower waters of <10 m. This may be due to physiological differences between plaice which prefers cooler waters compared to sole which prefers warmer waters (Engelhard *et al.*, 2011). However, the extent to which this will affect GEnS for the fisheries Descriptors cannot yet be predicted (Pörtner and Peck, 2010).

In influencing several GEnS Descriptors, Nicolas *et al.* (2014) directly linked the recent increase in SST to the decrease in *Calanus* biomass and a significant decline in adult and juvenile cod density (despite decreased fishing pressure) in the southern North Sea. Higher temperatures in early spring compromised cod recruitment due to reduced spawning success, earlier egg hatching, faster rapid larval development, a mismatch between prey availability and requirement and changes in primary production. This directly depleted cod rather than producing a marked northward migration and suggests that management measures to achieve GEnS for Descriptor 3 (fisheries) are unlikely to be effective against further decline. This also

suggests that the mechanisms behind changing species distributions are more complex than simple latitudinal migration.

Given the key role of the benthos in the functioning of hard and soft substrata, climate change affects the likelihood to achieve GEnS for several Descriptors, especially the biodiversity ones (Figure 3). The intertidal community has responded more quickly to climate driven warming for species close to their physiological tolerance limits. For example, the abundances of the two cooccurring intertidal Lusitanian barnacles Chthamalus montagui and C. stellatus have markedly increased since the mid-1900s in the British Isles whereas the Boreal Semibalanus balanoides proliferated during cooler periods but declined significantly due to temperature-driven competition (Birchenough et al., 2013; Mieszkowska et al. 2006, 2014). In soft sediments, the bivalve Macoma balthica has moved several 100s of km north in the Wadden Sea (Beukema et al., 2009). In contrast, there is less evidence for shifts in subtidal benthic communities which appear more buffered to increasing temperature than intertidal communities, plankton or fish (Hinz et al., 2011). Distributions of several warmer water species recorded in the English Channel in the late 1950s now remain largely unchanged. Despite this, Rombouts et al. (2012) predicted that, with continued warming, several key (ecologically or commercially important) species may be displaced northwards from the English Channel by 2100, due to increasingly unsuitable habitat. This ultimately reduces the possibility of achieving GEnS for the biodiversity and fisheries elements but shows the importance of the indicators chosen. As an indication of the importance of the offshore limits for GEnS, and what may be regarded as unbounded boundaries, Hinz et al. (2011) also suggested that depth changes may be more apparent than latitudinal ones since shallow coastal waters are more susceptible to temperature change.

Species, such as the Horse mussel *Modiolus modiolus*, which reach their southerly limit around the British Isles and are vulnerable but have a key role in benthic productivity and a high associated biodiversity (Gormley *et al.*, 2013). Its loss or northward migration would have a significant, negative impact on marine habitat functioning and thus achieving GEnS for the biodiversity and fisheries Descriptors. A progressive loss of suitable habitat and progressive spawning and recruitment failure at sub-optimum temperatures, will cause overall long-term decline.

As a dominant indicator for D4 (food-webs, Tasker, et al., 2010), recent warming-induced declines or migrations in overwintering distributions of many coastal wading birds will again reduce GEnS being attained for D1 and D4. These changes may have resulted from redistributing individuals rather than changes in survival, either due to cold-weather movements or changing juvenile recruitment patterns, for example, by seaducks taking advantage of ice-free conditions in the Baltic, and in coastal waterfowl changing estuarine overwintering use again indicating changing baselines in GEnS determination. Hence, any reliance on GEnS indicators

relating to dominant piscivorous seabirds (as proposed by the HELCOM and OSPAR Regional Seas Conventions) will be affected.

Recent models predict both warming-induced increasing and decreasing abundance of many wintering wader and waterbird populations in the UK. This may reduce the possibility not only of achieving GEnS but also Favourable Conservation Status under the EU Birds and Habitats Directives. While most current Special Protected Areas (Frost *et al.*, 2015) are likely to continue to support internationally important numbers of wintering waterbirds, even under a high-emissions 2080 scenario, there could be large overall changes to biodiversity.

Distributional shifts in response to climate change are not uniform between species and are not necessarily a simple, linear, change in depth distribution, poleward migration and/or contraction at the distribution edges (Richardson *et al.*, 2012). Furthermore, species ranges may fluctuate with harsh and mild periods (Parmesan, 2006), again increasing variability in the system and making GEnS for many Descriptors difficult to judge or baselines difficult to define. The availability of new, suitable habitat (in terms of temperature regime) does not necessarily mean that species will expand into it especially as hydrodynamic conditions, the biology of the newly colonising species compared to that of the established species and barriers to dispersal all influence the ability of a species to migrate (Hiscock *et al.*, 2004). Thermally-induced changes in parasite distribution may occur; for example, the protist *Perkinsus marinus* (which infects oysters) extended its range by over 500 km per year in the US (Parmesan, 2006). However, this may be exacerbated by relaying of shellfish in aquaculture. In addition to the increase in NIS, proliferation of such parasites may weaken populations at the edges of their ranges thus preventing achieving GEnS for, amongst others, D2 (Non-indigenous species) and the resulting effects on fisheries (D3).

Species distribution changes can bring together species that have not previously interacted (Staudinger *et al.*, 2013; Albouy *et al.*, 2014). Species with long generation times, low fecundity, low mobility, poor dispersal ability and populations living near the extremes of their physiological tolerances, and species with little phenotypic plasticity will be most vulnerable whilst those with high dispersal potential and phenotypic plasticity, which can adjust to and thrive in changing environments, will be favoured by climate change. However, the ability of a migrating, warm-adapted species to become established will depend on its competitive ability and the many interlinked responses (Figure 3) means that the repercussions for GEnS are almost impossible to predict. For example, contrary to a general assumption, species with broad ranges can be susceptible to extinction through climate change when species typically adapted to cooler environments persist under warming, preventing poleward expansion (Atkins and Travis, 2010; Staudinger *et al.*, 2013). In contrast, the amphipod *Echinogammarus marinus* is highly temperature sensitive and has retracted by 5°N at its European southerly edge (Guerra *et al.*,

2014). Thus a challenge for policy implementation is that the response of species to climate change may contradict expectation, making it difficult to disentangle localised human activities from those caused by climate change and, hence, manage changes in GEnS. Furthermore, species entering an area due to changes in the environmental conditions, may be at the edges of their distributions and hence could be rare and fragile, and thus of wider marine conservation interest.

II Altered temperature regime – individual physiological/phenological response

Temperature-induced physiological changes ultimately determine ecosystem composition, spatial structure and functioning (e.g. Pörtner and Karl, 2014) and thus many indicators of GEnS (Figure 4). Demographic changes resulting from alterations to recruitment, growth and survival together with phenological changes (e.g., timing of spawning, migration and spring blooms) lead to potential predator-prey mismatches (Philippart *et al.*, 2003; Dulvy *et al.*, 2008), thus reducing attaining GEnS for D4. Elevated temperature may increase reproductive success and larval survival of species at their northern limits, facilitating their poleward extension, assuming suitable habitat and a lack of physical and hydrological barriers (Rombouts *et al.*, 2012). Hence temperature changes may influence species interactions such as predation and competition, inherent to any indicators used for GEnS for D1, D2, D4 and D6 (Beukema *et al.*, 2009; Somero, 2012).

Temperature thresholds for survival and reproduction (e.g. see Rasmussen 1973) indicate that climate change will induce summer, warm-adapted spawners to have earlier and/or longer spawning, a longer growing season and greater productivity whereas winter spawners will have later and shorter spawning periods (Guerra *et al.*, 2014) (Figure 4). Temperature and salinity influence on the metabolic rate of organisms ultimately affects reproductive output and moulting. The advance of spring events such as spawning and migration has been recorded worldwide (Parmesan, 2006) thus again indicating moving baselines and reducing the ability to attain GEnS for several functional indicators and Descriptors.

As an example, thermal-induced early migration to spawning grounds occurs in flounder (*Platichthys flesus*) and veined squid (*Loligo forbesi*) amongst other species (Philippart *et al.*, 2003; Southward *et al.*, 2005; Teal *et al.*, 2008, 2012; Jansen and Gislason, 2011; Fincham *et al.*, 2013). Flounder spawning has advanced by 1.5 weeks yr⁻¹ since 1970 due to increasing temperature in the eastern English Channel and in the central and southern North Sea but not in areas of more stable temperatures (e.g. Bristol Channel) (Fincham *et al.*, 2013). Earlier spawning in North Sea mackerel (Jansen and Gislason, 2011) and early maturation of herring and sole (Fincham *et al.*, 2013) has been due to increased temperature although accelerated maturation in sole may be an evolutionary response to commercial fishing (Mollet *et al.*, 2007).

Such interactions conflate the responses due to different pressures, such as over-fishing and climate change, in turn making it difficult to determine the influence on achieving GEnS across D1 and D3 (Perry *et al.*, 2010; Pörtner and Peck, 2010; Griffith *et al.*, 2012). Teal *et al.* (2012) found an increased growth rate with higher summer temperatures in the warm-adapted sole and a positive correlation between size at the end of the first year and temperature whereas this was less in the cooler-adapted plaice, thus further advantaging sole and disadvantaging plaice (Dulvy *et al.* 2008; Engelhard *et al.* 2011; Teal *et al.*, 2012), again influencing stock production (Fincham *et al.*, 2013) and thus GEnS for D3.

Temperature-related early spawning, growth and maturation may benefit some but not all species, e.g. the northern bivalve *Macoma balthica*, if this results in a mismatch with the spring phytoplankton bloom and so reduced recruitment (Philippart *et al.*, 2003). Furthermore, a negative relationship between *M. balthica* recruitment and temperature produces a low density associated with warm winters (Beukema *et al.*, 2009). Similarly, reduced growth and higher mortality occurs with warm summers and mild winters, to prevent sufficient body condition gained during the summer to counteract autumn/winter weight loss. Given the central role of prey species such as *Macoma*, there is a large influence in certain marginal sea areas on achieving GEnS for D1, D4 and D6 (Figure 4). Beukema *et al.* (2009) also found that predation on *M. balthica* by the shrimp *Crangon crangon* explained half of interannual variability in recruitment again influencing GEnS for D4. Somero (2012) similarly found an increase in predation of *Mytilus edulis* by the seastar *Pisaster ochraceus*, correlated with increased temperature and had the potential to alter ecosystem structure and function. Given that *Crangon* and *Mytilus* are important commercial shellfish then any loss will prevent GEnS being achieved especially for D3.

Survival under temperature changes in marine ectotherms is partly due to physiological tolerance and adaptation and/or behavioural response (Hofmann and Todgham, 2010). Unregulated physiological processes may reduce growth and reproductive output and increase mortality, each affecting indicators of GEnS for several Descriptors. As yet, there are no indicators of GEnS related to individual functioning but rather to community and population effects, hence the concern regarding integrated effects across levels of biological organisation. Somero (2012) noted an increase in heart rate with increasing body temperature up to a critical point (CT_{max}) in *M. edulis* after which it decreases rapidly, and in the decapod crustacean *Petrolisthes*, lethality occurs at temperatures above CT_{max} with the most warm-adapted species acclimating to higher temperatures, increasing risk from climate change. In fish, limited cardiac performance may compromise migration ability and, thus, reproductive ability. Somero (2012) therefore hypothesised that the latitudinal range was, to some extent, dictated by cardiac

function. Given that such changes have a knock-on effect to fisheries populations then there is a resultant adverse effect on GEnS, again for D1 and D3.

In covering species of marine conservation importance, the above changes and their influence on GEnS Descriptors have also been shown in marine angiosperms. There are distribution changes in the intertidal seagrass *Zostera noltii* and decreased photosynthesis and growth with increased temperature even causing mortality (Massa *et al.*, 2009, 2011; Valle *et al.*, 2014). Whilst it would be unusual, but not impossible, particularly as the frequency of extreme weather events increases, for intertidal seagrasses to experience these temperatures in NW Europe, sublethal effects on growth occur in the subtidal species *Zostera marina* at 25°C, 2°C above the expected summer temperature for SW Europe. However, these experiments involving heat-shock protein expression do not indicate the implications of prolonged, regular exposure to elevated temperatures where there is potential for sub-lethal effects that may have population level implications. This makes it difficult to determine the long-term effect of such changes on the ability to meet GEnS especially with such moving baselines on decadal scales and where adaptation may occur over tens of generations.

In addition, *Zostera* species are particularly sensitive to habitat conditions and to erosion, increased turbidity, sedimentation and nutrient concentrations, showing the influence on GEnS for D1, D4, D5, D6 and D7 via physico-chemical changes (Figures 4 to 8). Through these processes, the increased frequency of storms may cause mortality and loss of seagrasses. Hence, given that recolonisation predominantly occurs through rhizome growth from adjacent vegetated areas (Boese *et al.*, 2009), seagrasses may disappear rather than change distribution. This will affect the ability to attain thresholds for indicators of GEnS that reflect such distributional and associated foodweb changes. Habitat change such as a loss of vegetation cover may also limit behavioural thermoregulation in some species. In contrast, habitats which retain their structural complexity under climate change may continue to provide shelter for the resident organisms which may respond more slowly to increased temperature than might be expected (Staudinger *et al.*, 2013). This reinforces the need to study habitat resilience in relation to changing environmental conditions and thus the ability to meet GEnS.

III Increased relative sea-level rise - physiographic changes

Physical changes ultimately affects indicators of biodiversity and ecological functioning and so changes due to climate change (Figure 5) reduce or change habitat such as a loss of productive intertidal area and a gain in less-productive subtidal systems, and a potential reduction in productivity and carrying capacity (Gray and Elliott, 2009). Loss of habitat requires either compensation and/or mitigation measures as any such loss of carrying capacity impinges on achieving GEnS for several of the Descriptors.

A global sea-level rise (SLR) of 0.2 to 0.6 m is predicted by 2100 and possibly 1 to 2 m if glacial meltwater is included (Katselidis *et al.*, 2014). Intertidal areas with a fixed high-water mark will experience coastal squeeze where the wetland, estuarine and high-shore habitats are prevented from migrating landward (Elliott *et al.*, 2014) although habitat loss will be variable due to shore type, sediment composition, habitat type, topography, exposure to erosive forces and inundation patterns (Pontee, 2013). Coastal squeeze may also create shore steepening due to the low water mark retreating landward more rapidly than the high water mark, which may be fixed or where it has advanced towards a retreating low water mark (Taylor *et al.*, 2004).

The MSFD D7 (hydrographical conditions) and D6 (seafloor integrity) address physical effects of climate change. Coastal erosion, linked to global warming and sea level rise, may impact most sandy beaches globally (Feagin *et al.*, 2005) and coastal infrastructure and/or sea defences or areas backed by natural barriers (Katselidis *et al.*, 2014) confine beaches to unnaturally narrow strips devoid of typical plant and animal communities. Increasing erosion due to SLR (Feagin *et al.*, 2005; Poulter *et al.*, 2009; Katselidis *et al.*, 2014) is largely assumed to be associated with soft sediment habitats, although rocky areas are also potentially vulnerable (Jackson and McIlvenny, 2011). Many beaches are erosional and a small increase in sea level will cause a large increase in erosion rates in the 19th and 20thC, especially in areas with hard engineering structures (Zhang *et al.*, 2004; Hanley *et al.*, 2014).

Dune plant communities, important in conservation, will be altered through confinement, fragmentation of communities, breakdown of succession, lack of species for stabilisation and the presence of remnant populations superimposed on erosion and physical barriers (Feagin et al., 2005). Agricultural development, urbanisation, tourism and recreation combined with climate change now threatens remaining dunes (Hanley *et al.*, 2014).

Habitat and biodiversity loss through coastal squeeze and coastal erosion/deposition may occur progressively whilst storm surges or periods of extreme weather (e.g. high winds and high rainfall) may cause rapid and significant geomorphological changes (Elliott et al., 2014). Storm surges are linked to erosion, are increasing in frequency and severity, and cause significant damage (Zhang *et al.*, 2004; Hanley *et al.*, 2014; Pörtner and Karl, 2014) sufficient to prevent achieving GEnS, particularly in terms of D1, D3, D4 and D6.

Finally, erosional changes in topography or shore profile induce sediment and habitat structure changes and hence biological community structure and ultimately D1 and D4 (Snelgrove and Butman, 1994). SLR is of concern as some MSFD indicators relate to the large and charismatic, conservation important species, e.g. to reduce nesting habitat for loggerhead turtles, especially where there are physical barriers to landward migration (Katselidis *et al.*, 2014). Similarly, Galbraith *et al.* (2002) and Erwin *et al.* (2006) highlighted that although increased inundation

may benefit waterfowl, it removes nesting and feeding habitat for many waterbirds. Similarly, North Sea plaice nursery grounds may be adversely affected by climate change with temperature-induced changes in currents, leading to reduced connectivity between spawning and nursery grounds (Hufnagl *et al.*, 2013). Hence the likely difficulty in attaining indicators of the Biodiversity Descriptors for GEnS which especially include those higher level and charismatic species such as fish, birds and sea turtles.

IV Increased climate variability effects on coastal hydrodynamics

Climate change will increase the variability and determine the trajectory of the over-riding influence of physical forcing factors on the structure and functioning of the marine system (e.g. Gray and Elliott, 2009) (Figure 6). This will remove coastal habitats and their prey populations, and require new refuge areas; hydrodynamic-induced erosion will remove some habitats but may supply sediment and change bathymetric patterns to secure new habitats especially in sheltered areas. For example, long-term variation in shallow coastal soft sediment communities follows substratum changes, hence increased storminess associated with climate change may reduce structure and function (Davoult et al., 1998; Smits et al., 2005; Weisse et al., 2005). While such communities may recover from severe physical disturbance, little is known about the impact of recurring events acting on communities that are partially recovering (Allan, 2006). This predominant effect on D6 and D7 then ultimately affects the seabed and nektonic components. However, the change is storminess changes with geographical area if at all (Muschinski and Katz, 2013) giving equivocal evidence despite the high control by coastal hydrodynamic features on ecological structure and processes (Scavia et al., 2002). Furthermore, even with such changes, the ability of coastal populations, which are adapted and hence resilient to high wave conditions, to be adversely affected by such hydromorphological changes is unknown. For example, benthic communities especially in highly-mobile sediments already are adapted to substratum changes such that they may be resilient to climate-induced variability (Gray and Elliott, 2009; Duarte et al, 2013b). Similarly, it is not possible to predict the result of ocean current pattern changes on larval transport with population and community level consequences (Harley et al., 2006), which affect many Descriptors and indicators. Therefore, the ability to achieve GEnS, especially for the Biodiversity Descriptors and D6 and D7, cannot be reliably predicted given that any knock-on effects from the coupled hydrophysical-ecological response are buffered by inherent variability and resilience.

V Changes to large scale climatic patterns due to land run-off

Large scale climatic patterns influence catchment run-off, including nutrients and contaminants, into semi-enclosed seas, hence showing interlinked responses (Figure 7). In Europe, the most influential patterns are the Atlantic Multidecadal Oscillation (AMO) (Kerr, 2000), the NAO

(Hurrell, 1995), and the East Atlantic (EA) Oscillation (Barnston and Livezey, 1987). Their impacts on biodiversity and ecosystems are well-known e.g. for the AMO (Drinkwater *et al.*, 2014; Gnanadesikan *et al.*, 2014; Harris *et al.*, 2014; Mieszkowska *et al.*, 2014; Nye *et al.*, 2014) and NAO (Ottersen *et al.*, 2001; Ji *et al.*, 2010; Henderson *et al.*, 2011; Kröncke *et al.*, 2011; David *et al.*, 2012; Henson *et al.*, 2012; Beaugrand *et al.*, 2014), but less for EA (Borja *et al.*, 2008; Chaalali *et al.*, 2013).

Arguably the greatest challenge in predicting the effects of climate change on the hydrodynamics of a catchment and hence the nutrient inputs to, and response in, enclosed coastal seas is the ability to understand these interlinked relationships (Meier *et al.*, 2011). In particular, nutrient run-off will create the adverse consequences of eutrophication (i.e. D5) but this is difficult to predict against a background of inherent variability due to changes in land-use patterns. Similarly, the influence of nutrients entering from the Northern Atlantic into the northern European seas due to NAO conditions (Frigstad *et al.*, 2013) gives effects over and above the influence of agricultural nutrients and industrial contaminants. Hence, there is the possibility of not meeting GEnS due to NAO irrespective of anthropogenic influences on land and again Member States may consider that such changes are outside their control.

As an indication of the influence of land-based climate patterns on the adjacent coastal and marine areas, modelling has shown the trajectory of recent (decadal) changes and indicated the overall future patterns in salinity and temperature (Andersen, 2012). For example, for the Baltic sea, climate change may create a warmer and less saline sea than seen in records since 1850, and climate-induced effects will occur earlier than previously thought (Meier *et al.*, 2012a, b). This results in adaptations by the food webs (Niiranen *et al.*, 2013) thus reducing the ability to meet GEnS for D4 or at least requiring the revision of the baseline and target values for this Descriptor.

VI Increased relative sea-level rise changing estuarine hydrodynamics

Tidal wetlands may be included within the MSFD if it is applied up to the high-water mark. As estuaries and other transitional waters are tidal then it is possible, but as yet undecided in all Member States, that they will be included in the MSFD. However, some countries, including the UK, have decided that estuaries will be excluded and that the MSFD will be applied from Mean High Water on the coast and seawards from the 'Bay closing lines' across the mouths of estuaries. Because of this, estuarine characteristics are included here briefly and only for comparison and completeness (Figure 8).

Climate change repercussions on the hydrogeomorphology of estuaries, such as SLR, increased salinity incursion and current pattern changes ultimately alter the fundamental characteristics of estuaries and their biodiversity (see Elliott and Whitfield, 2011), especially as most species

distributions reflect salinity tolerances (Whitfield *et al.*, 2012). Estuarine water budget changes, such as those caused by changed tidal and NAO patterns, also alter the salinity balance in estuaries and hence the distribution of brackish-tolerant species (e.g. Scavia *et al.*, 2002). However, the net effect of increased seawater influx as the result of SLR and the changes to catchment water balance and freshwater delivery into estuaries are unknown although the upper estuarine fauna may change with increased seawater incursion (Little, 2012).

As most sediment inputs in estuaries are from marine sources, estuarine bathymetry is influenced if marine incursion is increased and sea-level rises. This may impact wintering waterbird communities, especially where coastal defences are maintained. Similarly, any significant warming will reduce the Arctic and subarctic breeding ranges of wintering waterbirds and so despite improving winter conditions in the British Isles, wintering populations of many species here may decline due to the habitats necessary outside the area. Therefore, again, any GEnS high-level indicators focusing on waders and seabirds will be influenced by climate change and conditions well outside areas controlled by a Member State.

Given these overall changes, climate changes will adversely influence the GEnS Descriptors for biodiversity (D1), foodwebs (D4) and hydrophysical (D6) characteristics of the water column and substratum. Despite this, the ability of estuarine communities to withstand a larger variability while already being adapted to a high inherent variability (Elliott and Quintino, 2007) will make climate change responses difficult to detect.

VII Increased ocean acidification and seawater physico-chemical changes

The central conceptual model proposed here (Figure 2) centres on the direct and indirect effects of elevated atmospheric CO₂ inducing a 0.3 to 0.4 unit decrease in pH by 2100, i.e. ocean acidification (OA) although daily and annual variations compound difficulties in detecting change (Blackford and Gilbert, 2007; Dupont *et al.*, 2013; Williamson *et al.*, 2013; Artioli *et al.*, 2014). Although the vulnerability of marine biodiversity to OA may be minor (Hendriks *et al.*, 2010), detailed meta-analyses, especially on the benthos (macrofauna and macroalgae) and plankton, have highlighted important negative effects (Kroeker *et al.*, 2010) (Figure 9). This includes changes to physiology, growth and reproduction, loss or reduction of calcareous microplankton and macroalgae, and resultant changes to the planktonic and shore food-webs (Kroeker *et al.*, 2010; Durrieu de Madron *et al.*, 2011; Asnaghi *et al.*, 2013; Wittman and Pörtner, 2013).

Ocean acidification could increase the toxicity of contaminated sediments due to diagenesis, the mobility of metals in sediment pore water, with increases in the overlying water column and bioaccumulation of metals, in turn producing biological responses such as in clams (Carere *et al.*, 2011). Experimental exposure of metal-rich sediments to different predicted pH

concentrations show an increased metal toxicity and resultant effect on crustaceans (Roberts *et al.*, 2013). However, although this may affect many components comprising the GEnS, especially relating to ecological structure and functioning and even contaminant exposure (D8, 9), the timescale of effects, the rate at which the system over many generations can adapt to reduced pH and hence the final ecological consequences are unknown.

The ecological effects of OA based on laboratory and field studies is equivocal, even between different strains of a single species of phytoplankton (Langer *et al.*, 2009) such that it is difficult to predict long-term changes. Declining calcification rates with decreasing pH make coccolithophores particularly vulnerable (Riebesell *et al.*, 2000; Zondervan *et al.*, 2001; Fabry *et al.*, 2008) although their abundance has increased during the past two decades, a likely response to warming sea-surface temperature (Beare *et al.*, 2013; Beaugrand *et al.*, 2013). As a further concern, bivalve shellfisheries and aquaculture may also be affected by pH effects on the young stages especially at a time when aquaculture may be increasing to accommodate reductions in wild fisheries.

The scientific uncertainty shows the need for further study on ecosystem responses to OA as some species will be more susceptible whereas others may tolerate or adapt to the changing conditions. As biotic responses to OA will occur over decadal timescales together with other environmental pressures, genetic variability/selection, phenotypic plasticity and a wider range of ecological interactions (Williamson *et al.* 2013), then experiments or modelling are needed to interrogate such changes. The cumulative effects of these changes on achieving GEnS cannot as yet be predicted especially on the complex behaviour of Descriptors D1, D5, and D8 and its repercussions.

VIII Loss of polar ice cover and global transport repercussions

The 2014 IPCC report (Pörtner and Karl, 2014) highlights the increasing loss of polar ice cover which, together with opening Arctic shipping routes (Verny and Grigentin, 2009), is likely prevent GEnS being reached (Figure 10). The exchange of NIS via the Arctic occurred previously in warm periods of the Pleistocene (Dodson *et al.*, 2007), but now will be exacerbated by ballast water transport and other vessel vectors (Lewis *et al.*, 2004). These increasing vectors produce hazards and risks associated with NIS (Elliott *et al.*, 2014) although as yet it is unknown whether these become introduced, invasive or nuisance species, and whether they can be prevented or controlled (Olenin *et al.*, 2011). Consequently any D2 indicators of GEnS will be influenced by species either drifting on re-established current systems or via increased vessel transport. Most notably, this is similar to species entering the Mediterranean via the Suez Canal (cf. Galil *et al.*, 2014) but again raises the question of, firstly,

whether such vectors can be controlled and, secondly, whether a Member State may be liable for failing to meet GEnS for something out of its control.

There may be repercussions for the other biodiversity Descriptors and eutrophication (D5) given any resulting colonisation. For example, in 1999, a Pacific Ocean diatom species, *Neodenticula seminae*, occurred in the North Atlantic due to the summer of 1997/1998 experiencing the lowest extent of Arctic sea ice, leaving an ice-free passage (Reid *et al.*, 2007). It has become established in the North Atlantic phytoplankton community although as yet there appears to be no adverse effects other than a change to community composition. In addition, the input of fresh water, from Greenland ice melting, may increase nutrient inputs, give significant earlier blooms in the Arctic (Kahru *et al.*, 2011) and allow Atlantic phytoplankton species into the Arctic (Hegseth and Sundfjord, 2008).

Opening these routes also increases regional emissions of greenhouse gases and other hazardous materials (Macdonald *et al.*, 2005) (hence affecting D8, 9, 10) although the new routes could reduce net shipping emissions globally. Furthermore, although not yet quantified, the northern increase of shipping will increase the noise field in the NE Atlantic, thus potentially causing GEnS for D11 to fail, although as yet these changes cannot be quantified in scale, extent or duration.

Discussion

Trajectories of change and meeting baselines for GEnS

The MSFD follows the sequence of descriptor-criteria-indicator-target-monitoring-measures-management (European Commission, 2010) (Figure 1, inner circle) which aims to address anthropogenic stressors in a region but, as emphasised here, these cannot be separated form changes due to climate change. By definition, detecting GEnS (and also GEcS and FCS) is against the perceived and required status, i.e. a baseline, threshold or reference condition (e.g. Borja *et al.*, 2012) irrespective of whether that status changes due to climate change. In the highly variable marine environment this is made even more challenging due to changing and ill-defined boundaries, what may be termed *moving baselines* and *unbounded-boundaries*, and the status of any component (e.g. mobile species, hydrographic patterns) is influenced not only by activities and pressures in an area but also as the consequences of events at large distances. Hence, there is the challenge of detecting a signal (such as failure to meet GEnS) against a background of inherent variability (the so-called signal-to-noise ratio).

Summarising the MSFD Descriptor entries in Figures 2-10 in Table 2 shows the dominance of the repercussions for the biodiversity Descriptors (D1, 2, 4 and 6) especially the Biodiversity Descriptor 1 and its Criteria, indicators, metrics, targets and baselines. In particular, marine biodiversity change has to be judged against a defined baseline/reference/threshold value or

situation, the essence of the MSFD in determining whether GEnS is or is not met. This requires indicators or indices/metrics to be agreed in relation to monitoring, measures and management (Teixeira *et al.*, 2014). As shown here, wide-ranging climate change effects may prevent many indicators and targets set for achieving GEnS being met. The large empirical and modelling evidence (described above and in Table 1) shows the relative confidence that such changes will occur. However, as yet there is not the ability to extrapolate to quantitative predictions, especially in relation to the GEnS Descriptors.

Managing the marine ecosystem centres on separating the manageable endogenic signal from the effects of exogenic pressures, such as climate change, and from natural variability (Elliott, 2011). Although management resources are limited, the ability to apportion change to endogenous anthropogenic pressures is especially important to setting GEnS targets and identifying management measures (Greenstreet *et al.*, 2012) as is setting baselines against which ecosystem change will be interpreted. Hence the inherent difficulties in the ability to meet GEnS for biodiversity in areas where targets are impaired relies on understanding degradation and recovery of marine systems following the occurrence and removal of stressors (Borja *et al.*, 2010b; Duarte *et al.*, 2013b; Tett *et al.*, 2013)

As indicated above, North Atlantic marine and coastal species are responding to climate change through distribution and regime shifts (Table 3) and these have repercussions for achieving GEnS for most of the Descriptors. As a species distribution changes due to climate change, its value as an indicator of anthropogenic change is compromised (Beaugrand, 2003). If such a species is designated as an MSFD indicator, its shifting abundance due to climate must be incorporated into any target set. It may not be practical, however ecologically or economically important or well-studied a taxon, to set a target for it at the limits of its distribution if that taxon disappears due to climate-driven biogeographical shifts (McQuatters-Gollop, 2012). Hence, any species whose abundance is governed by exogenous drivers is unlikely to be a good indicator for endogenic anthropogenic pressures. Despite this, species are often designated of conservation importance because of their fragility or rarity which may be due to their occurrence at their geographical limit or a particular set of conditions.

Historical data are valuable in setting baseline conditions but this may not be possible due to climate change, especially as marine areas have changed considerably in the past six decades (Beaugrand *et al.*, 2002; Edwards and Richardson, 2004; McQuatters-Gollop *et al.*, 2011), thus altering regional food-webs and fish stocks (Kirby and Beaugrand, 2009). Compounding this are the effects of combined stressors, for example climate change and overfishing (Damanaki, 2011), hence with repercussions for GEnS for D3 on fisheries exploitation (Perry *et al.*, 2010; Pörtner and Peck, 2010; Greenstreet *et al.*, 2012; Griffith *et al.*, 2012;). The interaction of climate change with multiple pressures and their cumulative, synergistic and antagonistic effects

is particularly unknown (Brown et al., 2013). Interactions between climate, plankton and fish stocks may indicate that recovery of the latter to their previous levels is not possible, even assuming sustainable management. Thus the challenge then lies in deciding, for example amongst the MSFD indicators, which fish stocks can be regarded as a realistic 'baseline' but also in understanding that these changes may mean that GEnS cannot be either agreed or achieved.

The setting of targets and adequate monitoring is of utmost importance for an accurate assessment and consequently implementing adequate management measures to achieve GEnS. This requires that the science behind the pressure-impact-response sequence is adequate, i.e. that the amount of pressure required to produce an effect and then effect of a management response is well known. This is not the case - for example, as indicated for D5 (eutrophication) the science was inadequate where four ecosystems had different trajectories, ecological tipping points and hysteresis in response to nutrient abatement measures (Duarte *et al.*, 2009, 2013b; Andersen, 2012).

Similar failures under different pressures (e.g. fishing, aggregate extraction, etc.) could be due to broad changes in environmental and climate conditions, all affecting ecosystem dynamics, especially during conservation management in the expected long time of recovery after taking management measures (Borja *et al.*, 2010b). Hence setting reliable management targets in response to multiple shifting baselines under climate change is essential (Duarte *et al.*, 2009; 2013a). Ecosystem response thresholds have been also related to marine regime shifts that are characterised by various drivers, scales and potential for management action (Meiner and Reker, 2013). Thus reliable thresholds and targets will need revising with moving baselines, being dynamic instead of static. Thus the increased 'noise' in the system, due to climate change, will require the yet to be defined thresholds (as class limits) which may need to be fuzzy to reflect the moving baselines. Hence given the usual short-term societal response (Swaney *et al.*, 2012), the unpredictability of the changes require adaptive management which is made more challenging if the background involves moving baselines. This is particularly important in the case of the MSFD and its six-year reporting cycles but it remains to be seen whether such cycles can accommodate those moving baselines.

The repercussions for monitoring and management measures

Across Europe, there are many well-established regular monitoring programmes (Smith *et al.*, 2010; Patrício *et al.*, 2014) for all biodiversity components, which require to be continued but the extensive reduction in monitoring effort is an increasing cause for concern especially given current economic constraints (Borja and Elliott, 2013). These monitoring programmes are designed to assess the main changes resulting from single or multiple pressures but will be

required to be spatially and temporally extended to detect further changes arising from climate change in relation to other pressures, such as organic enrichment and fishing (e.g. for benthos: Kröncke and Reiss, 2010). Although as shown here there are good spatial data for some components, there is limited information on the effects of climate change on elements of high conservation importance such as cetaceans and seals (Evans and Bjørge, 2013). Spatial data sets may need combining to give time-series and observations on particular surrogate species (e.g. intertidal organisms) could indicate trends in climate change effects (e.g. Mieszkovska *et al.*, 2006; Nicolas *et al.*, 2011, 2014).

Given the above difficulty of setting baselines and determining whether GEnS has been achieved, a major consideration is the adequacy of monitoring proposed, whether the monitoring cycle will be sufficient and how many monitoring cycles will be needed to accurately detect change. There is still uncertainty regarding the monitoring and measures required and their robustness particularly to enable accurate assessments for complying with the MSFD and other directives (Boyes and Elliott, 2014), to improve the state of the marine environment and to ensure its sustainable development.

The separation of the effects of the local and wider pressures described above and detection of shifting baselines will require high-resolution spatio-temporal data (McQuatters-Gollop et al 2007; Couvet *et al.*, 2011. However, Patrício *et al.* (2014) showed that although European monitoring programmes address most biodiversity components and Descriptors (although less so for D6, Seafloor Integrity), the ability to detect variation due to climate change depends on the sampling intensity, frequency, geographical scale of the monitoring, standardisation (over time) of sampling and analysis techniques and data quality rather than on the number of programmes. In addition, Patrício *et al.* (2014) questioned whether the monitoring programmes are scientifically sound and fit-for-purpose.

The empirical evidence presented of the responses of marine species to climate change will require to be supplemented by spatial modelling (e.g. Rombouts *et al.*, 2012; Gormley *et al.*, 2013) which predict species distributions from habitat information (Monk *et al.*, 2012). However, the often mismatch between observed and expected distributions (see Reiss *et al.*, 2014) emphasises the inadequacy of current monitoring. Therefore, whilst modelling is of value, it has limitations, hence not only are changes to GEnS difficult to predict, they also cannot be detected or have a cause attributed to them especially of climate-driven timescales. Hence the background of climate change will increase the scientific resources needed in the MSFD implementation.

Given the above, climate change will have repercussions at each stage of the MSFD implementation (Figure 1, outer boxes). It requires the initial assessment to be revised given that

ecological and hydrophysical characteristics will change and the pressures list for an area has to be expanded to include external pressures such as climate change; some of the Descriptors, Criteria and indicators, especially those which rely on the distribution of particular species, will have to be revised or even omitted as being unsuitable. Most importantly, climate-affected baselines will have to be constantly revised during the six-year iterative cycle hence requiring extensive spatial and temporal monitoring to detect the signal-noise ratio obscured by climate change. The management measures proposed should address the causes and consequences of the endogenic managed pressures as well as the consequences of exogenic unmanaged pressures emanating from climate change (Elliott, 2011; Field et al., 2014). For example, while a management measure to control the inflow of non-indigenous species in ballast water can be proposed, the separation of these species from those entering via increased polar connectivity will be difficult to detect and to control.

Climate change, MSFD and the legal repercussions - 'Force majeure or natural causes'

This review emphasises the difficulty of implementing MSFD and achieving GEnS because of climate change. Any member State not fulfilling a Directive faces infraction proceedings for which there are considerable fines from the European Court of Justice. Article 14 of the MSFD indicates the following special cases for not meeting environmental targets or attaining GEnS: a) action or inaction for which the Member State concerned is not responsible, b) natural causes, c) force majeure, d) modifications or alterations to the physical characteristics of marine waters brought about by actions taken for reasons of overriding public interest which outweigh the negative impact on the environment, including any transboundary impact, e) natural conditions which do not allow timely improvement in the status of the marine waters concerned. Hence in any legal challenge, Member States may claim that climate change is preventing GEnS or its targets and indicators being met or met within the time stated due to clause (a), b), c) and/or e), because of shifting baselines, compromising the use of static reference conditions or targets, or without a return to a previous state of the system after restoration, because of changes in ecosystems due to climate change. The available scientific information, either empirical or modelling, will thus play a predominant role in addressing such a challenge and this will also centre on whether climate change is 'natural' or human-induced.

Article 14 requires Member States to prevent a 'deterioration in environmental status' due to the above causes but it is contended here that this rests on (i) proving that climate change does represent a deterioration rather than merely a change to another ecological state, and (ii) being able to address (mitigate) such a change. Of immediate relevance is that measures to deal with the consequences of Article 14 should be identified to the Commission at the time the overall programme of measures is proposed, i.e. 2015. The Article further advocates Member States take a regional approach to the causes of change but there is a further allowance that any actions

requiring disproportionate costs will not be required as long as there is no further deterioration to the environmental status. The final section of the Article implies that Member States have to fully justify their decision to avoid taking steps to counter environmental change but as long as not achieving GEnS is not permanent.

As an example of the impending argument and role of science, global temperature changes will cause local physiological changes in organisms thus impacting bioenergetic rates such as growth and feeding and potentially leading to changes in spawning thresholds (see Rasmussen 1973, Rijnsdorp et al. 2009). These organism changes will influence various metrics/indicators used to determine GEnS. This then has the potential to prevent GEnS being reached for several Descriptors and thus Member States will be threatened by infraction proceedings and being subject to heavy fines. Hence, a Member State could be penalised as the result of consequences of climate change which are not the result of its own action but rather global patterns. As in previous cases, the Member State would then engage scientific opinion and evidence to counter the claim and demonstrate either that this change is outside its control (force majeure), is a natural event and/or the system has not deteriorated but just changed. Hence, Member States should take this into account when designing monitoring networks, in order to quantify natural variability due to climate change. This information should then be used when assessing the status, to determine if the ecosystems experience either an unusual change or they do not return to the previous state when management measures are taken. This information should be used to justify, with appropriate and extensive scientific evidence, why GEnS is not achieved.

Concluding Remarks and Recommendations

As shown here, climate change produces impediments to implementing the MSFD and achieving GEnS and there are repercussions of those impediments: (1) the science-base is good on conceptual aspects but is required to give precise links between changes in biota and climate features; the 'so-what?' and what-if?' questions cannot yet be answered. New scientific developments may overcome this during several iterations of the MSFD process. At the organismal level, knowledge is needed on how abiotic factors interact to control the vital processes (survival, growth, feeding) of different life stages; this will allow parameterizing models to project the cumulative impacts caused by, for example, warming, reduced dissolved oxygen concentrations and decreased pH values. A mechanistic, cause-and-effect understanding is needed of how key abiotic factors interact to affect vital rates (including optimal and suboptimal limits defining the species fundamental niche) (Pörtner and Peck, 2010). At the population level, additional process knowledge is needed including how extrinsic and intrinsic properties of populations may be linked (e.g. see Planque *et al.* (2011) for marine fish populations). Species-level responses to habitat change caused by multiple, interacting stressors will probably differ among populations and so ecosystem-level projections must accommodate

changes in the strength of species interactions via bottom-up, top-down and intra-guild processes.

- (2) Climate change produces 'shifting baselines' which need to be accommodated in monitoring, particularly during the assessment of GEnS and marine management; actions will have to account for 'unbounded boundaries' given the ecology and climate change-induced migrations and dispersal of highly-mobile, nekton and plankton species. Hence, long-term and spatially large datasets are essential for signal-noise separation, to identify changes in ecological indicators, detect sudden and gradual ecosystem shifts and regime changes, and provide a baseline against which to interpret future changes. However, given that such datasets do not exist for most components then this may not be achieved. As the MSFD takes the current conditions as the baseline, predictions are required against current values.
- (3) More cost-effective spatial and temporal monitoring is required using current (e.g. Continuous Plankton Recorder, FerryBox), semi-autonomous or autonomous (sea gliders and wave gliders, moorings) or remote systems at the ecohydrodynamic rather than geographic scale. However, as monitoring budgets are being reduced, joint monitoring programmes are required across a suite of Descriptors. The absence of empirical data will increase the use of modelling but the error limits on the models may be large, and increase because of climate change, or even be unknown, thus giving poor predictability. Furthermore, existing models are adequate for scenario and semi-quantitative testing but not for detailed quantitative and accurate predictions.
- (4) Member States at present are only considering the means of determining GEnS on a Descriptor-by-Descriptor basis but at some stage before 2020 they need to consider aggregating these to give GEnS for a regional or sub-regional area (Borja *et al.*, 2014). Hence while assessing climate change on single Descriptors is the first priority, interactions amongst Descriptors and their changes due to climate change need addressing. Unless GEnS is defined across the Descriptors then ecosystem health (*sensu* Tett *et al.*, 2013), will not be determined. However, it is questioned whether the science is adequate to judge changes in health due to climate change and whether any resulting system is regarded as 'unhealthy' ('deteriorated' à *la* MSFD) or just different.
- (5) The challenges for marine monitoring and management result from having climate change superimposed on the effects of local activities and where climate change may either exacerbate or mask anthropogenic changes in the Descriptors. For example, whilst anthropogenic nutrient inputs from its catchment will be controlled by Member States to achieve GEnS for eutrophication, bloom-forming species not otherwise in an area may arrive and cause failure

- (e.g. Andersen, 2012). Detecting change against a greater inherent variability will increase monitoring costs, a challenge in economically difficult times (Borja and Elliott, 2013).
- (6) Climate-driven spatial and temporal variation should be interrogated including a potential geographic disparity to achieving GEnS across the marine environment in general and across the regional seas in NE Atlantic in particular. Raised temperature may have greater effects in northern than southern Europe but these are equivocal. Hence, baselines will have to be revised on a site-specific basis although the evidence needs to be extrapolated to show the short, medium and long-term effects and the speed of environmental response. Modelling is required to indicate how quickly communities can reach a new equilibrium but there is now an urgent need to show adaptation (or the lack of it) overs 10s to 100s of generation times for marine organisms.
- (7) Although not discussed further here, society will place emphasis on the repercussions of non-achieving GEnS for the Ecosystem Services and Societal Benefits obtained from the regional seas (e.g. Atkins *et al.*, 2011). The loss of these due both to managed pressures but also climate change has to be determined and emphasised to environmental managers and policymakers (Luisetti *et al.*, 2014; Turner et al, 2014).
- (8) The failure to meet GEnS because of climate change has wide-ranging legal repercussions and could lead to a Member State being placed in infraction proceedings. A legal challenge will arise not because of the pressures inside the waters of a Member State under which they might have some control (Endogenic managed pressures), but because of the external and non-controlled pressures (Exogenic unmanaged pressures). For example, a NW European Member State may be threatened with legal (infraction) proceedings for failing GEnS for non-indigenous species entering via new Arctic-related vectors over which the Member State has no control (*cf.* the corresponding case for the Suez Canal, Galil *et al.*, 2014). The legal defence, that the failure was the result of third-party actions, natural causes or *force majeure*, would require to be supported by robust science.
- (9) These lessons are relevant and applicable not only to European seas and the implementation of the MSFD but also to other global areas, for example during the implementation of the Canada Oceans Act and the US Oceans Act 2000 (US Congress 2002). While the latter does not give the same degree of detail as the MSFD in achieving healthy and productive seas and it does not mention climate change in its few pages, determining and managing change due to separating this from other anthropogenic pressures have to be considered.

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Supporting Information

Table S1. Qualitative Descriptors, Criteria and indicators, selected by the European Commission (2010), and to be used in the assessment of the environmental status of marine waters, in the context of the Marine Strategy Framework Directive.

Table S2. Normative definition of Good Environmental Status as suggested by Task Groups for each Descriptor and modified from Cardoso *et al.* (2010). Key: GEnS: good environmental status; IAS: Invasive alien species; NIS: non-indigenous species; EnQS: Environmental Quality Standards.

References

Albouy C, Velez L, Coll M, Colloca F, Le Loc'h F, Mouillot D, Gravel. 2014. From projected species distribution to food-web structure under climate change. *Global Change Biology*, **20**: 730-741.

Alcock R. 2003. The effects of climate change on rocky shore communities in the Bay of Biscay, 1895–2050. *PhD. Thesis, University of Southampton*: 296 p.

Allan R. 2006. Impacts of Climate Change on Storminess in Marine Climate Change Impacts Annual Report Card 2006 (Eds. Buckley PJ, Dye SR and Baxter JM), Online Summary Reports, MCCIP, Lowestoft, www.mccip.org.uk.

Andersen JH. 2012. Ecosystem-based management of coastal eutrophication. Connecting science, policy and society. PhD thesis. University of Copenhagen. 56 pp + annexes.

Artioli Y, Blackford JC, Nondal G, Bellerby RGJ, Wakelin SL, Holt JT, Butenschön M, Allen JI. 2014. Heterogeneity of impacts of high CO2 on the North Western European Shelf. *Biogeosciences*, 11: 601-612.

Asnaghi V, Chiantore M, Mangialajo L, Gazeau F, Francour P, Alliouane S, Gattuso J-P. 2013. Cascading Effects of Ocean Acidification in a Rocky Subtidal Community. *PLoS ONE*, 8: e61978.

Atkins JP, Burdon D, Elliott M, Gregory AJ. 2011. Management of the Marine Environment: Integrating Ecosystem Services and Societal Benefits with the DPSIR Framework in a Systems Approach. *Marine Pollution Bulletin*, 62(2): 215-226.

Atkins KE, Travis JMJ. 2010. Local adaptation and the evolution of species under climate change. *Journal of Theoretical Biology* 266: 449-457

Barnston AG, Livezey RE. 1987. Classification, seasonality, and persistence of low-frequency atmospheric circulation pattern. *Monthly Weather Review*, 115: 1083-1126.

Beare D, McQuatters-Gollop A, van der Hammen T, Machiels M, Teoh SJ, Hall-Spencer J. 2013. Long-term trends in calcifying plankton and pH in the North Sea. *PLOS ONE* 8, e61175.

Beaugrand G. 2003. Long-term changes in copepod abundance and diversity in the north-east Atlantic in relation to fluctuations in the hydroclimatic environment. *Fisheries Oceanography* 12, 270-283.

Beaugrand G. 2004. The North Sea regime shift: evidence, causes, mechanisms and consequences. *Progress in Oceanography* 60, 245-262.

Beaugrand G. 2009. Decadal changes in climate and ecosystems in the North Atlantic Ocean and adjacent seas. *Deep Sea Research Part II: Topical Studies in Oceanography*, 56: 656-673.

Beaugrand G. 2012. Unanticipated biological changes and global warming. *Marine Ecology Progress Series* 446, 293-301.

Beaugrand G, Goberville E, Luczak C, Kirby RR. 2014. Marine biological shifts and climate. *Proceedings of the Royal Society B: Biological Sciences*, 281.

Beaugrand G, McQuatters-Gollop A, Edwards M, Goberville E. 2013. Long-term responses of North Atlantic calcifying plankton to climate change. *Nature Climate Change* 3, 263-267.

Beaugrand G, Reid PC, Ibanez F, Lindley JA, Edwards M. 2002. Reorganization of North Atlantic marine copepod biodiversity and climate. *Science* 296, 1692-1694.

Beukema JJ. 2002. Expected changes in the benthic fauna of Wadden Sea tidal flats as a result of sea-level rise or bottom subsidence. *Netherlands Journal of Sea Research* 47, 25–39.

Beukema JJ, Dekker R, Jansen JM. 2009. Some like it cold: populations of the tellinid bivalve *Macoma balthica* (L.) suffer in various ways from a warming climate. *Marine Ecology Progress Series*. 384: 135-145.

Birchenough SNR, Bremner J, Henderson P, Hinz H, Jenkins S, Mieszkowska N. 2013. Shallow and shelf subtidal habitats and ecology. 2013. Marine Climate Change Impacts Partnership MCCIP Annual Report Card 2012-2013. Published online 28 November 2013 doi:10.14465/2013.arc20.193-203.

Birchenough SNR, Degraer S, Reiss H, Borja A, Braeckman U, Craeymeersch J, de Mesel I, Kerckhof F, Kröncke I, Mieszkowska N, et al., 2011. Responses of marine benthos to climate change. In: Reid PC, Valdés L (Eds.) ICES Status report on climate change in the North Atlantic. *ICES Cooperative Research Report*, **310**: 123-146.

Blackford, J. and Gilbert, F. 2007. pH variability and CO2 induced acidification in the North Sea. *Journal of Marine Systems*, **64**, 229–241.

Boese B, Kaldy JE, Clinton PJ, Eldridge PM, Folger CL. 2009. Recolonization of intertidal *Zostera marina* L. (eelgrass) following experimental shoot removal. *Journal of Experimental Marine Biology and Ecology*. 347: 69-77.

Borja A, Prins T, Simboura N, Andersen JH, Berg T, Marques J-C, Neto JM, Papadopoulou N, Reker J, Teixeira H, Uusitalo L. (2014) Tales from a thousand and one ways to integrate marine ecosystem components when assessing the environmental status. *Frontiers in Marine Science*. 1:22. doi: 10.3389/fmars.2014.00022

Borja, Á, Dauer DM, Elliott, Simenstad, CA. 2010b. Medium- and long-term recovery of estuarine and coastal ecosystems: patterns, rates and restoration effectiveness. *Estuaries and Coasts* 33:1249–1260.

Borja Á, Elliott M. 2013. Marine monitoring during an economic crisis: the cure is worse than the disease. *Marine Pollution Bulletin* 68: 1-3.

Borja Á, Fontán A, Muxika I. 2013b. Interactions between climatic variables and human pressures upon a macroalgae population: Implications for management. *Ocean & Coastal Management*, **76**: 85-95.

Borja A, Fontán A, Sáenz J, Valencia V. 2008. Climate, oceanography, and recruitment: the case of the Bay of Biscay anchovy (*Engraulis encrasicolus*). *Fisheries Oceanography*, 17: 477-493.

Borja Á, Dauer DM, Grémare A. 2012. The importance of setting targets and reference conditions in assessing marine ecosystem quality. *Ecological Indicators*, 12: 1-7.

Borja A, Elliott M, Andersen JH, Cardoso AC, Carstensen J, Ferreira JG, Heiskanen A-S, Marques JC, Neto J, Teixeira H, et al., 2013a. Good Environmental Status of marine ecosystems: What is it and how do we know when we have attained it? *Marine Pollution Bulletin*, 76: 16-27.

Borja Á, Elliott M, Carstensen J, Heiskanen A-S, van de Bund W. 2010a. Marine management - Towards an integrated implementation of the European Marine Strategy Framework and the Water Framework Directives. *Marine Pollution Bulletin*, 60: 2175-2186.

Boyes SJ, Elliott M. 2014. Marine Legislation – the ultimate 'horrendogram': International Law, European Directives & National Implementation. *Marine Pollution Bulletin*, in press. doi: 10.1016/j.marpolbul.2014.06.055

Bremner J. 2008. Species' traits and ecological functioning in marine conservation and management. *Journal of Experimental Marine Biology and Ecology*, **366**: 37-47.

Brown CJ, Schoeman DS, Sydeman WJ, Brander K, Buckley LB, Burrows M, Duarte CM, Moore PJ, Pandolfi JM, Poloczanska E, Venables W, Richardson AJ. 2011. Quantitative approaches in climate change ecology. *Global Change Biology* vol 17: 3697 – 3713.

Brown, C. J., M. I. Saunders, H. P. Possingham, A. J. Richardson, 2013. Managing for Interactions between Local and Global Stressors of Ecosystems. PLoS ONE, 8: e65765.

Cardoso AC, Cochrane S, Doerner D, Ferreira JG, Galgani F, Hagebro C, Hanke G, Hoepffner N, Keizer PD, Law R, et al., 2010. *Scientific support to the European Commission on the Marine Strategy Framework Directive*. Management Group Report. EUR 24336 EN – Joint Research Centre, Luxembourg: Office for Official Publications of the European Communities: 57 pp.

Carere M, Miniero R, Cicero MR. 2011. Potential effects of climate change on the chemical quality of aquatic biota. *Trends in Analytical Chemistry*, 30: 1214-1221.

Caswell BA, Frid CLJ. 2013. Learning from the past: functional ecology of marine benthos during eight million years of aperiodic hypoxia, lessons from the Late Jurassic. *Oikos*, 122: 1687-1699.

CEC. 2005. Proposal for a Directive of the European Parliament and of the Council establishing a Framework for Community Action in the field of Marine Environmental Policy (Marine Strategy Directive) [SEC(2005) 1290]. COM (2005) 505 final 2005/0211 (COD). Commission of the European Communities, Brussels, 24.10.2005.

CEC. 2013. COMMISSION STAFF WORKING DOCUMENT: Climate change adaptation, coastal and marine issues: *Accompanying the document* Communication from the Commission to the European Parliament, the Council, The European Economic and Social Committee and the Committee of the Regions, An EU Strategy on adaptation to climate change, SWD (2013) 133 final, Brussels.

CEC. 2014. COMMISSION STAFF WORKING DOCUMENT Annex Accompanying the document Commission Report to the Council and the European Parliament The first phase of

implementation of the Marine Strategy Framework Directive (2008/56/EC) - The European Commission's assessment and guidance {COM(2014) 97 final}, SWD(2014) 49 final, Brussels.

Chaalali A, Beaugrand G, Boët P, Sautour B. 2013. Climate-Caused Abrupt Shifts in a European Macrotidal Estuary. *Estuaries and Coasts*, 36: 1193-1205.

Cheung WWL, Sarmiento JL, Dunne J, Frolicher TL, Lam VWY, Deng Palomares ML, Watson R, Pauly D. 2013a. Shrinking of fishes exacerbates impacts of global ocean changes on marine ecosystems. *Nature Climate Change*, 3: 254-258.

Cheung WWL, Watson R, Pauly D. 2013b. Signature of ocean warming in global fisheries catch. *Nature*, **497**: 365-368.

Christensen JH, Hewitson B, Busuioc A, Chen A, Gao X, Held I, Jones R, Kolli RK, Kwon W-T, Laprise R, et al. 2007: Regional Climate Projections. In: *Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*, In: Solomon S, Qin D, Manning M, Chen Z, Marquis M, Averyt KB, Tignor M, Miller HL. (eds.). Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.

Chust G. Albaina A, Aranburu A, Borja Á, Diekmann OE, Estonba A, Franco J, Garmendia JM, Iriondo M, Muxika I, et al., 2013a. Connectivity, neutral theories and the assessment of species vulnerability to global change in temperate estuaries. *Estuarine, Coastal and Shelf Science*, **131**: 52-63.

Chust G, Castellani C, Licandro P, Ibaibarriaga L, Sagarminaga Y, Irigoien X. 2014a. Are *Calanus* spp. shifting poleward in the North Atlantic? A habitat modelling approach. *ICES Journal of Marine Science*, 71: 241-253.

Chust G, Allen JI, Bopp L, Schrum C, Holt J, Tsiaras K, Zavatarelli M, Chifflet M, Cannaby H, Dadou I, et al., 2014b. Biomass changes and trophic amplification of plankton in a warmer ocean. *Global Change Biology*, 20: 2124-2139.

Chust G, Irigoien X, Chave J, Harris RP. 2013b. Latitudinal phytoplankton distribution and the neutral theory of biodiversity. *Global Ecology and Biogeography*, 22: 531-543.

Claussen U, Connor D, de Vrees L, Leppänen J, Percelay J, Kapari M, Mihail O, Ejdung G, Rendell J. 2011. Common Understanding of (Initial) Assessment, Determination of Good Environmental Status (GES) and Establishment of Environmental Targets (Art. 8, 9 & 10 MSFD). WG GES EU MSFD (https://circabc.europa.eu/sd/d/ce7e2776-6ac6-4a41-846f-a04832c32da7/05_Info_Common_understanding_final.pdf)

Cochrane SKJ, Connor DW, Nilsson P, Mitchell I, Reker J, Franco J, Valavanis V, S. Moncheva S, Ekebom J, Nygaard K, et al., 2010. *Marine Strategy Framework Directive – Task*

Group 1 Report Biological Diversity. EUR 24337 EN – Joint Research Centre, Luxembourg: Office for Official Publications of the European Communities: 110 pp.

Cormier R, Kannen A, Elliott M, Hall P, Davies IM. (Eds) 2013. Marine and Coastal Ecosystem-based Risk Management Handbook. *ICES Cooperative Research Report*, No. 317, March 2013, International Council for the Exploration of the Sea, Copenhagen, 60pp, ISBN 978-87-7472-115-1.

Couvet D, Devictor V, Jiguet F, Julliard R. 2011. Scientific contributions of extensive biodiversity monitoring. *Comptes Rendus Biologies*, 334: 370-377.

Cury PM, Shin Y-J, Planque B, Durant JM. 2008. Ecosystem oceanography for global change in fisheries. *Trends in Ecology & Evolution*, **23**: 338-346.

Damanaki M, 2011. European fisheries reform speech. Brussels, 13 July 2011. http://www.guardian.co.uk/environment/2011/jul/13/europe-fisheries-reform-speech-maria-damanaki.

David V, Ryckaert M, Karpytchev M, Bacher C, Arnaudeau V, Vidal N, Maurer D, Niquil N. 2012. Spatial and long-term changes in the functional and structural phytoplankton communities along the French Atlantic coast. *Estuarine, Coastal and Shelf Science*, 108: 37-51.

Davoult D, Dewarumez J-M, Migne A. 1998, Long-term changes (1979-1994) in two coastal benthic communities (English Channel): analysis of structural developments. *Oceanologica* Acta. 21(4): 609-617.

Defeo O, Castilla JC. 2012. Governance and governability of coastal shellfisheries in Latin America and the Caribbean: multi-scale emerging models and effects of globalization and climate change. *Current Opinion in Environmental Sustainability*, **4**: 344-350.

Dodson JJ, Tremblay S, Colombani F, Carscadden JE, Lecomte F. 2007. Trans-Arctic dispersals and the evolution of a circumpolar marine fish species complex, the capelin (*Mallotus villosus*). *Molecular Ecology* 16, 5030–5043 doi: 10.1111/j.1365-294X.2007.03559.x

Drinkwater KF, Beaugrand G, Kaeriyama M, Kim S, Ottersen G, Perry RI, Pörtner H-O, Polovina JJ, Takasuka A. 2010. On the processes linking climate to ecosystem changes. *Journal of Marine Systems*, **79**: 374-388.

Drinkwater KF, Miles M, Medhaug I, Otterå OH, Kristiansen T, Sundby S, Gao Y. 2014. The Atlantic Multidecadal Oscillation: Its manifestations and impacts with special emphasis on the Atlantic region north of 60°N. *Journal of Marine Systems*, 133: 117-130.

Duarte CM, Conley DJ, Carstensen J, Sánchez-Camacho M, 2009. Return to Neverland: Shifting baselines affect restoration targets. *Estuaries and Coasts*, 32: 29-36.

Duarte CM, Borja A, Carstensen J, Elliott M, Krause-Jensen D, Marbà N. 2013b. Paradigms in the Recovery of Estuarine and Coastal Ecosystems. *Estuaries and Coasts*, in press. DOI 10.1007/s12237-013-9750-9

Duarte L, Viejo RM, Martínez B, deCastro M, Gómez-Gesteira M, Gallardo T. 2013a. Recent and historical range shifts of two canopy-forming seaweeds in North Spain and the link with trends in sea surface temperature. *Acta Oecologica*, **51**: 1-10.

Dulvy NK, Rogers SI, Jennings S, Stelzenmüller V, Dye SR, Skoldal HR. 2008. Climate change and deepening of the North Sea fish assemblage: a biotic indicator of warming seas. *Journal of Applied Ecology*. 45: 1029-1039

Dupont S, Dorey N, Stumpp M, Melzner F, Thorndyke M, 2013. Long-term and trans-life-cycle effects of exposure to ocean acidification in the green sea urchin Strongylocentrotus droebachiensis. *Marine Biology*, 160(8), 1835-1843, doi:10.1007/s00227-012-1921-x

Durrieu de Madron X, Guieu C, Sempéré R, Conan P, Cossa D, D'Ortenzio F, Estournel C, Gazeau F, Rabouille C, Stemmann L, et al. 2011. Marine ecosystems' responses to climatic and anthropogenic forcings in the Mediterranean. *Progress in Oceanography*, 91: 97-166.

Dutertre M, Beninger PG, Barillé L, Papin M, Haure J. 2010. Rising water temperatures, reproduction and recruitment of an invasive oyster, Crassostrea gigas, on the French Atlantic coast. *Marine Environmental Research*, **69**: 1-9.

Edwards M, Beaugrand G, Reid PC, Rowden AA, Jones MB. 2002. Ocean climate anomalies and the ecology of the North Sea. *Marine Ecology Progress Series* 239, 1-10.

Edwards M, Richardson AJ. 2004. Impact of climate change on marine pelagic phenology and trophic mismatch. *Nature* 430, 881-884.

Elliott M, Whitfield A. 2011. Challenging paradigms in estuarine ecology and management. *Estuarine, Coastal & Shelf Science*, 94: 306-314.

Elliott M, Quintino V. 2007. The Estuarine Quality Paradox, Environmental Homeostasis and the difficulty of detecting anthropogenic stress in naturally stressed areas. *Marine Pollution Bulletin*, 54, 640-645.

Elliott M. 2011. Marine science and management means tackling exogenic unmanaged pressures and endogenic managed pressures – a numbered guide. *Marine Pollution Bulletin*, 62: 651-655.

Elliott M. 2014. Integrated marine science and management: wading through the morass. *Marine Pollution Bulletin*, in press, 86(1/2). P1-3. doi: 10.1016/j.marpolbul.2014.07.026

Elliott M, Cutts ND, Trono A. 2014. A typology of marine and estuarine hazards and risks as vectors of change: a review for vulnerable coasts and their management. *Ocean & Coastal Management* 93: 88-99.

Engelhard GH, Pinnegar JK, Kell LT, Rijnsdorp AD. 2011. Nine decades of North Sea sole and plaice distribution. *ICES Journal of Marine Science*. 68(6): 1090-1104

Eriksson Wiklund AK, Dahlgren K, Sundelin B, Andersson A. 2009. Effects of warming and shifts of pelagic food web structure on benthic productivity in a coastal marine system. *Marine Ecology Progress Series*, **396**: 13-25.

Erwin RM, Sanders GM, Prosser DJ, Cahoon DR. 2006. High tides and rising seas: potential effects on estuarine waterbirds. *Studies in Avian Biology*. 32: 214-228.

European Commission, 2000. Directive 2000/60/EC of the European Parliament and of the Council of 23 October 2000 establishing a framework for community action in the field of water policy. *Official Journal of the European Union*, L327: 1-72.

European Commission, 2008. Directive 2008/56/EC of the European Parliament and of the Council establishing a framework for community action in the field of marine environmental policy (Marine Strategy Framework Directive). *Official Journal of the European Union*, L164: 19-40.

European Commission, 2010. Commission Decision of 1 September 2010 on criteria and methodological standards on good environmental status of marine waters (notified under document C (2010) 5956)(2010/477/EU). Official Journal of the European Union, L232: 12-24.

Evans PGH, Bjørge A. 2013 Impacts of climate change on marine mammals, *MCCIP Science Review* 2013, 134-148, doi:10.14465/2013.arc15.134-148.

Fabry VJ, Seibel BA, Feely RA, Orr JC. 2008. Impacts of ocean acidification on marine fauna and ecosystem processes. *ICES Journal of Marine Science* 65, 414-432.

Feagin RA, Sherman DJ, Grant WE. 2005. Global sea-level rise and the loss of sand dune plant habitats. *Frontiers in Ecology and the Environment*. 3(7): 359-364

Ferreira JG, Andersen JH, Borja A, Bricker SB, Camp J, Cardoso da Silva M, Garcés E, Heiskanen A-S, Humborg C, Ignatiades L, et al., 2010. Marine Strategy Framework Directive – Task Group 5 Report Eutrophication. *EUR 24338 EN – Joint Research Centre, Luxembourg: Office for Official Publications of the European Communities*: 49 pp.

Field, C, Barros V, Mach K, Mastrandrea M. (Coordinating lead authors). 2014. Climate Change 2014: Impacts, Adaptation, and Vulnerability. IPCC WGII AR5 Technical Summary: 76 pp.

Fincham J, Rijnsdorp AJ, Engelhard GH. 2013. Shifts in the timing of spawning of sole linked to warming sea temperatures. *Journal of Sea Research*. 75: 69-76.

Finney BP, Alheit J, Emeis K-C, Field DB, Gutiérrez D, Struck U. 2010. Paleoecological studies on variability in marine fish populations: A long-term perspective on the impacts of climatic change on marine ecosystems. *Journal of Marine Systems*, **79**: 316-326.

Frigstad H, Andersen T, Hessen DO, Jeansson E, Skogen M, Naustvoll L-J, Miles MW, Johannessen T, Bellerby RGJ. 2013. Long-term trends in carbon, nutrients and stoichiometry in Norwegian coastal waters: Evidence of a regime shift. *Progress in Oceanography*, 111: 113-124.

Frost, M, Baxter J, Bayliss-Brown G, Buckley P, Cox M, Dye S, Stoker B, Withers HN (in press). Climate change and the implementation of key marine biodiversity legislation in the UK. *Aquatic Conservation: Marine and Freshwater Ecosystems*, in press.

Galbraith H, Jones R, Park R, Clought J, Herrod-Julius S, Harrington B, Page G. 2002. Global climate change and sea level rise: potential losses of intertidal habitat for shorebirds. *Waterbirds: The International Journal of Waterbird Biology*. 25(2): 173-183.

Galgani F, Fleet D, van Franeker J, Katsanevakis S, Maes T, Mouat J, Oosterbaan L, Poitou I, Hanke G, Thompson R, et al. 2010. Marine Strategy Framework Directive – Task Group 10 Report Marine litter. *EUR 24340 EN – Joint Research Centre, Luxembourg: Office for Official Publications of the European Communities*: 48 pp.

Galil BS, Marchini A, Occhipinti-Ambrogi A, Minchin D, Narščius A, Ojaveer H, Olenin S. 2014. International arrivals: widespread bioinvasions in European Seas. *Ethology Ecology & Evolution*, DOI: 10.1080/03949370.2014.897651

García R, Holmer M, Duarte CM, Marbà N. 2013. Global warming enhances sulphide stress in a key seagrass species (NW Mediterranean). *Global Change Biology*: 19: 3629-3639.

Gnanadesikan A, Dunne JP, Msadek R. 2014. Connecting Atlantic temperature variability and biological cycling in two earth system models. *Journal of Marine Systems*, 133: 39-54.

Gormley KSG, Porter JS, Bell MC, Hull AD, Sanderson WG. 2013. Predictive Habitat Modelling as a Tool to Assess the Change in Distribution and Extent of an OSPAR Priority Habitat under an Increased Ocean Temperature Scenario: Consequences for Marine Protected Area Networks and Management. *PLOS ONE*. 8(7):1-16.

Gray JS, Elliott M. 2009. *Ecology of Marine Sediments: science to management*. OUP, Oxford, 260pp.

Greenstreet SPR, Rossberg AG, Fox CJ, Le Quesne WJF, Blasdale T, Boulcott P, Mitchell I, Millar C, Moffat CF. 2012. Demersal fish biodiversity: species-level indicators and trends-based targets for the Marine Strategy Framework Directive. *ICES Journal of Marine Science*, 69: 1789-1801.

Griffith GP, Fulton EA, Gorton R, Richardson A. 2012. Predicting Interactions among fishing, ocean warming, and ocean acidification in a marine system with whole-ecosystem models. *Conservation Biology*, 26(6): 1145–1152.

Guerra A, Leite N, Marques JC, Ford AT, Martins I. 2014. Predicting the variation in *Echinogammarus marinus* populations at its southernmost limits under global warming scenarios: Can the sex ratio make a difference? *Science of the Total Environment*. 466-467: 1022-1029.

Hale R, Calosi P, McNeill L, Mieszkowska N, Widdicombe S. 2011. Predicted levels of future ocean acidification and temperature rise could alter community structure and biodiversity in marine benthic communities. *Oikos*, 120: 661-674.

Hanley ME, Hoggart SPG, Simmonds DJ, Bichot A, Colangelo MA, Bozzeda F, Heurtefeux H, Ondiviela B, Ostrowski R, Recio M, et al. 2014. Shifting sands? Coastal protection by sandbanks, beaches and dunes. *Coastal Engineering*. 87: 136-146

Harley CDG, Hughes AR, Hultgren KM, Miner BG, Sorte CJB, Thornber CS, Rodriguez LF, Tomanek L, Williams SL. 2006. The impacts of climate change in coastal marine systems. *Ecology Letters*. 9: 228-241.

Harris V, Edwards M, Olhede SC. 2014. Multidecadal Atlantic climate variability and its impact on marine pelagic communities. *Journal of Marine Systems*, 133: 55-69.

Hegseth EN, Sundfjord A. 2008. Intrusion and blooming of Atlantic phytoplankton species in the high Arctic. *Journal of Marine Systems*, 74: 108-119.

Hellmann JJ, Byers JE, Bierwagen BG, Dukes JS. 2008. Five Potential Consequences of Climate Change for Invasive Species. *Conservation Biology*, **22**: 534-543.

Helmuth B, Mieszkowska N, Moore P, Hawkins SJ. 2006. Living on the edge of two changing worlds: forecasting the responses of rocky intertidal ecosystems to climate change. *Annual Review of Ecological Evolutionary Systems*. 37: 373-404.

Hemery G, D'Amico F, Castege I, Dupont B, D'Elbee J, Lalanne Y, Mouches C. 2008. Detecting the impact of oceano-climatic changes on marine ecosystems using a multivariate index: The case of the Bay of Biscay (North Atlantic-European Ocean). *Global Change Biology*, **14**: 27-38.

Henderson PA, Seaby RMH, Somes JR. 2011. Community level response to climate change: The long-term study of the fish and crustacean community of the Bristol Channel. *Journal of Experimental Marine Biology and Ecology*, 400: 78-89.

Hendriks IE, Duarte CM, Álvarez M. 2010. Vulnerability of marine biodiversity to ocean acidification: A meta-analysis. *Estuarine*, *Coastal and Shelf Science*, 86: 157-164.

Henson S, Lampitt R, Johns D. 2012. Variability in phytoplankton community structure in response to the North Atlantic Oscillation and implications for organic carbon flux. *Limnology and Oceanography*, 57: 1591–1601.

Hewitt JE, Thrush SF. 2010. Empirical evidence of an approaching alternate state produced by intrinsic community dynamics, climatic variability and management actions. *Marine Ecology Progress Series*, **413**: 267-276.

Hinz H, Capasso E, Lilley M, Frost M, Jenkins S. 2011. Temporal differences across a biogeographical boundary reveal slow response of sub-littoral benthos to climate change, *Marine Ecology Progress Series*, 423, 69 - 82.

Hiscock K, Southward A, Tittley I, Hawkins S. (2004) Effects of changing temperature on benthic marine life in Britain and Ireland, *Aquatic Conservation: Marine and Freshwater Ecosystems*, 14, 333-362.

Hoegh-Guldberg O, Bruno JF. 2010. The Impact of Climate Change on the World's Marine Ecosystems. *Science*, **328**: 1523-1528.

Hofmann GE, Todgham AE. 2010. Living in the now: physiological mechanisms to tolerate a rapidly changing environment. *Annual Review of Physiology*. 72: 127-145

Hufnagl M, Peck MA, Nash RDM, Pohlmann T, Rijnsdorp AD. 2013. Changes in potential North Sea spawning grounds of plaice (*Pleuronectes platessa* L.) based on early life stage connectivity to nursery habitats. *Journal of Sea Research*. 84: 26-39.

Hurrell JW, 1995. Decadal trends in the North Atlantic Oscillation: regional temperatures and precipitation. *Science* 269, 676–679.

Jackson AC, McIlvenny J. 2011. Coastal squeeze on rocky shores in northern Scotland and some possible ecological impacts. Journal of Experimental Marine Biology and Ecology. 400: 314-321.

Jansen T, Gislason H. 2011. Temperature affects the timing of spawning and migration of North Sea mackerel. *Continental Shelf Research*. 31: 64-72.

Jennings S, Brander K. 2010. Predicting the effects of climate change on marine communities and the consequences for fisheries. *Journal of Marine Systems*, **79**: 418-426.

Ji R, Edwards M, Mackas DL, Runge JA, Thomas AC. 2010. Marine plankton phenology and life history in a changing climate: current research and future directions. *Journal of Plankton Research*, 32: 1355-1368.

Junker K, Sovilj D, Kröncke I, Dippner J. W. 2012. Climate induced changes in benthic macrofauna - A non-linear model approach. *Journal of Marine Systems*, **96-97**: 90-94.

Kahru M, Brotas V, Manzano-Sarabia M, Mitchell BG. 2011. Are phytoplankton blooms occurring earlier in the Arctic? *Global Change Biology*, 17: 1733-1739.

Katselidis KA, Schofield G, Stamou G, Dimopoulos P, Pantis JD. 2014. Employing sea-level rise scenarios to strategically select sea turtle nesting habitat important for long-term management at a temperate breeding area. *Journal of Experimental Marine Biology and Ecology*. 450:47-54.

Kerr RA, 2000.A North Atlantic climate pacemaker for the centuries. *Science* 288 (5473): 1984–1986.

Kirby RR, Beaugrand G. 2009. Trophic amplification of climate warming. *Proceedings of the Royal Society B-Biological Sciences* 276, 4095-4103.

Kroeker KJ, Kordas RL, Crim RN, Singh GG. 2010. Meta-analysis reveals negative yet variable effects of ocean acidification on marine organisms. *Ecology Letters*, 13: 1419-1434.

Kröncke I, Reiss H. 2010. Influence of macrofauna long-term natural variability on benthic indices used in ecological quality assessment. *Marine Pollution Bulletin* 60, 58-68.

Kröncke I, Reiss H, Eggleton JD, Aldridge J, Bergman MJN, Cochrane S, Craeymeersch JA, Degraer S, Desroy N, Dewarumez, J-M., et al. 2011. Changes in North Sea macrofauna communities and species distribution between 1986 and 2000. *Estuarine, Coastal and Shelf Science*, 94: 1-15.

Langer G, Nehrke G, Probert I, Ly J, Ziveri P. 2009. Strain-specific responses of *Emiliania huxleyi* to changing seawater carbonate chemistry. *Biogeosciences*, 6, 2637–2646.

Law R, Hanke G, Angelidis M, Batty J, Bignert A, Dachs J, Davies I, Denga Y, Duffek A, Herut B, et al. 2010. Marine Strategy Framework Directive – Task Group 8 Report Contaminants and pollution effects. *EUR 24335 EN – Joint Research Centre, Luxembourg: Office for Official Publications of the European Communities*: 161 pp.

Lejeusne C, Chevaldonné P, Pergent-Martini C, Boudouresque CF, Pérez, T. 2010. Climate change effects on a miniature ocean: the highly diverse, highly impacted Mediterranean Sea. *Trends in Ecology & Evolution*, **25**: 250-260.

Levin LA, Ekau W, Gooday AJ, Jorissen F, Middelburg JJ, Naqvi SWA, Neira C, Rabalais NN, Zhang J. 2009. Effects of natural and human-induced hypoxia on coastal benthos. *Biogeosciences J1 - BG*, **6**: 2063-2098.

Lewis PN, Riddle MJ, Hewitt CL. 2004. Management of exogenous threats to Antarctica and the sub-Antarctic islands: balancing risks from TBT and non-indigenous marine organisms. *Marine Pollution Bulletin*, 49(11-12): 999-1005.

Lima FP, Ribeiro PA, Hawkins SJ, Santos AM. 2007. Do distributional shifts of northern and southern species of algae match the warming pattern? *Global Change Biology* **13**: 2592-2604.

Little S. 2012. The impact of increasing saline penetration upon estuarine and riverine benthic macroinvertebrates. PhD Thesis. Loughborough University, UK. Available from: https://dspace.lboro.ac.uk/2134/9737

Luisetti T, Turner RK, Jickells T, Andrews J, Elliott M, Schaafsma M, Beaumont N, Malcolm S, Burdon D, Adams C, Watts W. 2014. Coastal Zone Ecosystem Services: From science to values and decision making; a case study. *Science of the Total Environment* 493: 682–693.

Macdonald RW, Harner T, Fyfe J. 2005. Recent climate change in the Arctic and its impact on contaminant pathways and interpretation of temporal trend data. *Science of the Total Environment*, 342: 5-86.

Martinez B, Arenas F, Rubal M, Burgues S, Esteban R, Garcia-Plazaola I, Figueroa FL, Pereira R, Saldana L, Sousa-Pinto I, et al. 2012. Physical factors driving intertidal macroalgae distribution: physiological stress of a dominant fucoid at its southern limit *Oecologia* 170(2): 341-353.

Massa SI, Pearson GA, Aires T, Kub M, Olsen JL, Reinhard R, Serrão EA, Arnaud-Haond S. 2011. Expressed sequence tags from heat-shocked seagrass Zostera noltii (Hornemann) from its southern distribution range. *Marine Genomics*. 4(3): 181-188

Massa S, Arnaud-Haond S, Pearson GA, Serrão EA. 2009. Temperature tolerance and survival of intertidal populations of the seagrass Zostera noltii (Hornemann) in Southern Europe (Ria Formosa, Portugal). *Hydrobiologia*. 619:195-201 DOI 10.1007/s10750-008-9609-4

McQuatters-Gollop A. 2012. Challenges for implementing the Marine Strategy Framework Directive in a climate of macroecological change, *Phil. Trans. R. Soc. A*, vol 370, no. 1980, p5636-5655 http://rsta.royalsocietypublishing.org/content/370/1980/5636.abstract

McQuatters-Gollop A, Raitsos DE, Edwards M, Pradhan Y, Mee LD, Lavender SJ, Attrill MJ. 2007. A long-term chlorophyll dataset reveals regime shift in North Sea phytoplankton biomass unconnected to nutrient levels. *Limnology and Oceanography* 52, 635-648.

McQuatters-Gollop A, Reid PC, Edwards M, Burkill P, Castellani C, Batten S, Gieskes W, Beare D, Bidigare R, Head E, et al., 2011. Is there a decline in marine phytoplankton? *Nature* 472, E6-E7.

Mee LD, Jefferson RL, Laffoley DdA, Elliott M. 2008. How good is good? Human values and Europe's proposed Marine Strategy Directive. *Marine Pollution Bulletin*, 56: 187-204.

Meier HEM, Andersson H, Arheimer B, *et al.* 2012b. Comparing reconstructed past variations and future projections of the Baltic Sea ecosystem—first results from multi-model ensemble simulations. *Environmental Research Letters*, **7**, 034005.

Meier HEM, Eilola K, Almroth E. 2011. Climate-related changes in marine ecosystems simulated with a 3-dimensional coupled physical-biogeochemical model of the Baltic Sea. *Climatic Research* 48, 31-55

Meier HEM, Hordoir R, Andersson HC, Dieterich C, Eilola K, Gustafsson BG, Höglund A, Schimanke S. 2012a. Modeling the combined impact of changing climate and changing nutrient loads on the Baltic Sea environment in an ensemble of transient simulations for 1961–2099. *Climate Dynamics* DOI 10.1007/s00382-012-1339-7

Meiner A, Reker J. 2013. Balancing the future of Europe's coasts — knowledge base for integrated management. European Environment Agency, Report, 12: 64 pp.

Mendoza-González G, Martínez ML, Rojas-Soto OR, Vázquez G, Gallego-Fernández J. B. 2013. Ecological niche modeling of coastal dune plants and future potential distribution in response to climate change and sea level rise. *Global Change Biology*, **19**: 2524-2535.

Mieszkowska N, Burrows MT, Pannacciulli FG, Hawkins SJ. 2014. Multidecadal signals within co-occurring intertidal barnacles Semibalanus balanoides and Chthamalus spp. linked to the Atlantic Multidecadal Oscillation. *Journal of Marine Systems*, 133: 70-76.

Mieszkowska N, Kendall MA, Hawkins SJ, Leaper R, Williamson P, Hardman-Mountford NJ, Southward AJ. 2006. Changes in the range of some common rocky shore species in Britain - a response to climate change? *Hydrobiologia* 555: 241-251.

Mollet FM, Kraak SBM, Rijnsdorp, AD. 2007. Fisheries-induced evolutionary changes in maturation norms in the North Sea sole *Solea solea. Marine Ecology Progress Series*. 351: 189-199.

Monk J, Ierodiaconou D, Harvey E, Rattray A, Versace VL. 2012. Are we predicting the actual or apparent distribution of temperate marine fishes? *PLOS One*: 7(4): 1-11

Munday PL, Warner RR, Monro K, Pandolfi JM, Marshall DJ. 2013. Predicting evolutionary responses to climate change in the sea. *Ecology Letters*, **16**: 1488-1500.

Muschinski T, Katz JI. 2013. Trends in hourly rainfall statistics in the United States under a warming climate. *Nature Climate Change* 3, 577-580.

Neiva J, Assis J, Fernandes F, Pearson GA, Serrão EA. 2014. Species distribution models and mitochondrial DNA phylogeography suggest an extensive biogeographical shift in the high-intertidal seaweed Pelvetia canaliculata. *Journal of Biogeography*: 41(6): 1137-1148.

Nicolas D, Chaalali A, Drouineau H, Lobry J, Uriarte A, Borja A, Boët P. 2011. Impact of global warming on European tidal estuaries: some evidence of northward migration of estuarine fish species. *Regional Environmental Change*, **11**: 639-649.

Nicolas D, Rochette S, Llope M, Licandro P. 2014. Spatio-temporal variability of the North Sea cod recruitment in relation to temperature and zooplankton. *PLOS ONE*. 9(2): 1-13.

Niiranen S, Yletyinen J, Tomczak MT, Blenckner T, Hjerne O, MacKenzie BR, Müller-Karulis B, Neumann T, Meier HEM. 2013. Combined effects of global climate change and regional ecosystem drivers on an exploited marine food web. *Global Change Biology*, **19**: 3327-3342.

Nye JA, Link JS, Hare JA, Overholtz WJ. 2009. Changing spatial distribution of fish stocks in relation to climate and population size on the Northeast United States continental shelf. *Marine Ecology Progress Series*, **393**: 111-129.

Nye JA, Baker MR, Bell R, Kenny A, Kilbourne KH, Friedland KD, Martino E, Stachura MM, Van Houtan KS, Wood R. 2014. Ecosystem effects of the Atlantic Multidecadal Oscillation. *Journal of Marine Systems*, 133: 103-116.

O'Brien TD, Li WKW, Moran XAG. 2012. ICES Phytoplankton and Microbial Plankton Status Report 2009/2010, ICES Cooperative Research Report. International Council for the Exploration of the Sea, Copenhagen, p. 197.

O'Brien TD, Wiebe PH, Falkenhaug T. 2013. ICES Zooplankton Status Report 2010/2011, ICES Cooperative Research Report. International Council for the Exploration of the Sea, Copenhagen, p. 210.

Occhipinti-Ambrogi A. 2007. Global change and marine communities: Alien species and climate change. *Marine Pollution Bulletin*, **55**: 342-352.

Olenin S, Elliott M, Bysveen I, Culverhouse P, Daunys D, Dubelaar GBJ, Gollasch S, Goulletquer P, Jelmert A, Kantor Y, et al. 2011. Recommendations on methods for the detection and control of biological pollution in marine coastal waters. *Marine Pollution Bulletin*, 62(12): 2598-2604.

Olenin S, Alemany F, Cardoso AC, Gollasch S, Goulletquer P, Lehtiniemi M, McCollin T, Minchin D, Miossec L, Occhipinti-Ambrogi A, et al. 2010. Marine Strategy Framework

Directive – Task Group 2 Report Non-indigenous species. *EUR 24342 EN – Joint Research Centre, Luxembourg: Office for Official Publications of the European Communities*: 44 pp.

Otero M, Garrabou J, Vargas M. 2013. Mediterranean Marine Protected Areas and climate change: A guide to regional monitoring and adaptation opportunities. *Malaga, Spain: IUCN*: 52 p.

Ottersen G, Planque B, Belgrano A, Post E, Reid P, Stenseth N. 2001. Ecological effects of the North Atlantic Oscillation. *Oecologia*, 128: 1-14.

Ottersen G, Kim S, Huse G, Polovina JJ, Stenseth NC. 2010. Major pathways by which climate may force marine fish populations. *Journal of Marine Systems*, **79**: 343-360.

Parmesan C. 2006. Ecological and evolutionary responses too recent climate change. *Annual Review of Ecology and Evolutionary Systems* 37: 637-669.

Patrício J, Little S, Mazik K, Thomson S, Zampoukas Z, Teixeira H, Solaun O, Uyarra MC, Papadopoulou N, Kaboglu G, et al. 2014. Report on SWOT analysis of monitoring. DEVOTES Deliverable 1.4. http://www.devotes-project.eu/wp-content/uploads/2014/02/DEVOTES_D1-4 Report-on-SWOT-analysis-of-monitoring.pdf

Perry AL, Low PJ, Ellis JR, Reynolds JD. 2005. Climate change and distribution shifts in marine fishes. *Sciencexpress*, 12 May 2005: 1-8.

Perry RI, Cury P, Brander K, Jennings S, Möllmann C, Planque B. 2010. Sensitivity of marine systems to climate and fishing: Concepts, issues and management responses. *Journal of Marine Systems*, 79: 427–435.

Philippart CJM, van Aken HM, Beukema JJ, Bos OG, Cadee GC, Dekker R. 2003. Climate-related changes in recruitment of the bivalve *Macoma balthica*. *Limnology and Oceanography*. 48(6): 2171-2185

Piet GJ, Albella AJ, Aro E, Farrugio H, Lleonart J, Lordan C, Mesnil B, Petrakis G, Pusch C, Radu G, Ratz HJ. 2010. Marine Strategy Framework Directive – Task Group 3 Report Commercially exploited fish and shellfish. *EUR* 24316 *EN* – *Joint Research Centre, Luxembourg: Office for Official Publications of the European Communities*: 82 pp.

Planque B, Loots C, Petitgas P, Lindstrom U, Vaz S. 2011. Understanding what controls the spatial distribution of fish populations using a multi-model approach. *Fisheries Oceanography*, 20: 1–17.

Poloczanska ES, Smith S, Fauconnet L, Healy J, Tibbetts IR, Burrows M, Richardson AJ. 2011. Little change in the distribution of rocky shore faunal communities on the Australian east coast

after 50 years of rapid warming. *Journal of Experimental Marine Biology & Ecology*. 400(1-2): 145-154.

Poloczanska ES, Brown CJ, Sydeman WJ, Kiessling W, Schoeman DS, Moore P, Brander K, Bruno JF, Buckley LB, Burrows MT, et al. 2013. Global imprint of climate change on marine life. *Nature Climate Change*. 3(10): 919-925.

Pontee N. 2013. Defining coastal squeeze: A discussion. *Ocean & Coastal Management*. 84: 204-207.

Pörtner HO, Karl D (Coordinating lead authors). 2014. Chapter 6. Ocean systems. IPCC WGII AR5 Chapter 6: 138 pp.

Pörtner HO, Peck MA. 2010. Climate change impacts on fish and fisheries: towards a cause and effect understanding. *Journal of Fish Biology*, 77: 1745–1779

Poulter B, Feldman RL, Brinson MM, Horton BP, Orbach MK, Pearsall SH, Reyes E, Riggs SR, Whitehead JC. 2009. Sea-level rise research and dialogue in North Carolina: creating windows for policy change. *Ocean & Coastal Management*. 52, 147–153.

Ramirez-Llodra E, Tyler PA, Baker MC, Bergstad OA, Clark MR, Escobar E, Levin LA, Menot L, Rowden AA, Smith CR, Van Dover CL. 2011. Man and the Last Great Wilderness: Human Impact on the Deep Sea. *PLoS ONE*, **6**: e22588.

Rasmussen E. 1973. Systematics and ecology of the Isefjord marine fauna (Denmark). *Ophelia*, 11(1): 1-507.

Reid PC, Valdés L. 2011. ICES status report on climate change in the North Atlantic. *ICES Cooperative Research Report*, **310**: 262 p.

Reid PC, Edwards M, Beaugrand G, Skogen M, Stevens D. 2003. Periodic changes in the zooplankton of the North Sea during the twentieth century linked to oceanic inflow. *Fisheries Oceanography* 12, 260-269.

Reid PC, Edwards M, Hunt HG, Warner AJ. 1998. Phytoplankton change in the North Atlantic. *Nature* 391, 546.

Reid PC, Johns DG, Edwards M, Starr M, Poulin M, Snoeijs P. 2007. A biological consequence of reducing Arctic ice cover: arrival of the Pacific diatom Neodenticula seminae in the North Atlantic for the first time in 800,000 years. *Global Change Biology* 13, 1910-1921.

Reiss H, Birchenough S, Borja A, Buhl-Mortensen L, Craeymeersch J, Dannheim J, Darr A, Galparsoro I, Gogina M, Neumann, H, et al., 2014 Benthos distribution modelling and its relevance for marine ecosystem management. *ICES Journal of Marine Science*, doi:10.1093/icesjms/fsu107

Rice J, Arvanitidis C, Borja A, Frid C, Hiddink J, Krause J, Lorance P, Ragnarsson SA, Skold M, Trabucco B. 2010. Marine Strategy Framework Directive – Task Group 6 Report Seafloor integrity. *EUR* 24334 *EN* – *Joint Research Centre, Luxembourg: Office for Official Publications of the European Communities*: 73 pp.

Richardson A, Brown CJ, Brander K, Bruno JF, Buckley L, Burrows MT, Duarte CM, Halpern, BS, Hoegh-Goldberg O, Holding J, et al., 2012. Climate change and marine life. *Biology Letters*. 8: 907-909

Riebesell U, Zondervan I, Rost B, Tortell PD, Zeebe RE, Morel FMM. 2000. Reduced calcification of marine plankton in response to increased atmospheric CO₂. *Nature* 407, 364-367.

Rijnsdorp A, Peck MA, Engelhard GH, Möllmann C, Pinnegar JK. 2009. Resolving the effect of climate change on fish populations. *ICES Journal of Marine Science*, 66: 1570–1583.

Roberts DA, Birchenough SNR, Lewis C, Sanders MB, Bolam T, Sheahan D. 2013. Ocean acidification increases the toxicity of contaminated sediments. *Global Change Biology*, 19: 340-351.

Rogers S, Casini M, Cury P, Heath M, Irigoien X, Kuosa H, Scheidat M, Skov H, Stergiou KI, Trenkel VM, et al. 2010. Marine Strategy Framework Directive – Task Group 4 Report Food Webs. *EUR 24343 EN – Joint Research Centre, Luxembourg: Office for Official Publications of the European Communities*: 55 pp.

Rombouts I, Beaugrand G, Fizzala X, Gaill F, Greenstreet SPR, Lamare S, Le Loc'h F, McQuatters-Gollop A, Mialet B, Niquil N, et al. 2013. Food web indicators under the Marine Strategy Framework Directive: From complexity to simplicity? *Ecological indicators*, vol. 29, p246-254 http://www.sciencedirect.com/science/article/pii/S1470160X12004402

Rombouts I, Beaugrand G, Dauvin J-C. 2012. Potential changes in benthic macrofaunal distributions from the English Channel simulated under climate change scenarios. *Estuarine*, *Coastal and Shelf Science*, **99**: 153-161.

Scavia D, Field JC, Boesch, DF, Buddemeier, RW, Burkett, V, Cayan DR, Fogarty, M; Harwell MA, Howarth RW, Mason C, et al. 2002. Climate Change Impacts on U. S. Coastal and Marine Ecosystems. *Estuaries*, Vol. 25, No. 2. (Apr., 2002), pp. 149-164.

Smith H, Mazik K, Davey A, Nixon S, Codling I. 2010. *Developing best practice to underpin the UKMMAS: marine monitoring protocols database*. WRc Report to Defra. Contract ME4126.

Smits A, Klein Tank AMG, Konnen GP. 2005: Trends in storminess over the Netherlands, 1962-2002. *International Journal of Climatology*, 25, 1331-1344.

Snelgrove PVR, Butman CA. 1994. Animal-sediment relationships revisited: cause versus affect. *Oceanography & Marine Biology Annual review*. 32: 111-177.

Somero G. 2012. The physiology of global change: linking patterns to mechanisms. *Annual Review of Marine Science*. 4: 39-61.

Sorte CJB, Williams SL, Carlton JT. 2010. Marine range shifts and species introductions: comparative spread rates and community impacts. *Global Ecology and Biogeography*, 19: 303-316.

Southward AJ, Langmead O, Hardman-Mountford NJ, Aiken J, Boalch GT, Dando P, Genner MJ, Joint I, Kendall MA, Halliday NC, et al., 2005 Long-term oceanographic and ecological research in the western English Channel. *Advances in Marine Biology*. 47: 2-105.

Staudinger MD, Carter SL, Cross MS, Dubois NS, Duffy JE, Enquist C, Griffiths R, Hellmann J, Lawler JL, O'Leary JO, et al. 2013. Biodiversity in a changing climate: a synthesis of current and projected trends in the US. *Frontiers in Ecology and Environment*. 11(9): 465-473.

Stocker T, Dahe Q, Plattner GKE. 2013. Working Group I Contribution to the IPCC Fifth Assessment report Climate Change 2013: The Physical Science Basis. Final Draft Underlying Scientific-Technical Assessment IPCC: 2216 p.

Swaney DP, Humborg C, Emeis K, Kannen A, Silvert W, Tett P, Pastres R, Solidoro C, Yamamuro M, Henocque Y, Nicholls R. 2012. Five critical questions of scale for the coastal zone. *Estuarine, Coastal and Shelf Science*, 96: 9-21.

Swartenbroux F, Albajedo B, Angelidis M, Aulne M, Bartkevics V, BesadaV, Bignert A, Bitterhof A, Hallikainen A, Hoogenboom R, et al. 2010. Marine Strategy Framework Directive – Task Group 9 Report Contaminants in fish and other seafood. *EUR 24339 EN – Joint Research Centre, Luxembourg: Office for Official Publications of the European Communities*: 36 pp.

Sydeman WJ, Bograd SJ. 2009. Marine ecosystems, climate and phenology: introduction. *Marine Ecology Progress Series*, **393**: 185-188.

Tasker MLE. 2008. The effect of climate change on the distribution and abundance of marine species in the OSPAR Maritime Area. *ICES Cooperative Research Report*, **293**: 45 p.

Tasker ML, Amundin M, Andre M, Hawkins A, Lang W, Merck T, Scholik-Schlomer A, Teilmann J, Thomsen F, Werner S, Zakharia M. 2010. Marine Strategy Framework Directive – Task Group 11 Report Underwater noise and other forms of energy. *EUR 24341 EN – Joint Research Centre, Luxembourg: Office for Official Publications of the European Communities*: 55 pp.

Taylor JA, Murdock AP, Pontee NI. 2004. A macroscale analysis of coastal steepening around the coast of England and Wales. *The Geographical journal*. 170(3): 179-188

Teal L, de Leeuw JJ, van der Veer HW, Rijnsdorp AD. 2008. Effects of climate change on growth of 0-group sole and plaice. *Marine Ecology Progress Series*. 358: 219-230.

Teal LR, van Hal R, van Kooten T, Ruardij P, Rijnsdorp AD. 2012. Bio-energetics underpins the spatial response of North Sea plaice (Pleuronectes platessa L.) and sole (Solea solea L.) to climate change. *Global Change Biology*, 18(11) 3291-3305.

Teixeira H, Berg T, Fürhaupter K, Uusitalo L, Papadopoulou N, Bizsel KC, Cochrane S, Churilova T, Heiskanen A-S, Uyarra M, et al. 2014. Existing biodiversity, non-indigenous species, food-web and seafloor integrity GES indicators. DEVOTES Deliverable 3.1. 198pp + 2 Annexes. http://www.devotes-project.eu/wp-content/uploads/2013/10/Deliverable-1.3-Monitoring_networks-31-oct-2013.pdf

Tett P, Gowen R, Painting S, Elliott M, Forster R, Mills D, Bresnan E, Capuzzo E, Fernandes T, Foden J, et al. 2013. Framework for understanding marine ecosystem health. *Marine Ecology Progress Series* 494: 1–27 + suppl. material.

Thorner J, Kumar L, Smith SDA. 2014. Impacts of Climate-Change-Driven Sea Level Rise on Intertidal Rocky Reef Habitats Will Be Variable and Site Specific. *PLoS ONE*, 9: e86130.

Tomczak MT, Dinesen GE, Hoffmann E, Maar M, Støttrup JG. 2013. Integrated trend assessment of ecosystem changes in the Limfjord (Denmark): Evidence of a recent regime shift? *Estuarine, Coastal and Shelf Science*, **117**: 178-187.

Travers M, Shin Y-J, Jennings S, CuryP. 2007. Towards end-to-end models for investigating the effects of climate and fishing in marine ecosystems. *Progress in Oceanography*, **75**: 751-770.

Turner K, Schaafsma M, Elliott M, Burdon D, Atkins J, Jickells T, Tett P, Mee L, van Leeuwen S, Barnard S, et al. 2014. UK National Ecosystem Assessment Follow-on. Work Package Report 4: Coastal and marine ecosystem services: principles and practice. UNEP-WCMC, LWEC, UK.

US Congress. 2002. An Act to establish a Commission on Ocean Policy and for other purposes. PL106-256, S.2327, US Congress, Washington DC, pp5.

Valle M, Chust G, del Campo A, Wisz MS, Olsen SM, Garmendia JM, Borja Á. 2014. Projecting future distribution of the seagrass Zostera noltii under global warming and sea level rise. *Biological Conservation*, **170**: 74-85.

Valle M, van Katwijk MM, de Jong DJ, Bouma TJ, Schipper AM, Chust G, Benito BM, Garmendia JM, Borja Á. 2013. Comparing the performance of species distribution models of Zostera marina: Implications for conservation. *Journal of Sea Research*, **83**: 56-64.

Verny J, Grigentin C. 2009. Container shipping on the Northern Sea Route. *International Journal of Production Economics*, 122(1), 107-117.

Webb TJ, Tyler EHM, Somerfield PJ, 2009. Life history mediates large-scale population ecology in marine benthic taxa. *Marine Ecology Progress Series*, **396**: 293-306.

Weisse R, von Storch H, Feser F. 2005. Northeast Atlantic and North Sea storminess as simulated by a regional climate model 1958-2001 and comparison with observations. *Journal of Climate*, 18, 465-479.

Wernberg T, Thomsen MS, Tuya F, Kendrick GA. 2011. Biogenic habitat structure of seaweeds change along a latitudinal gradient in ocean temperature. *Journal of Experimental Marine Biology and Ecology*, **400**: 264-271.

Whitfield AK, Elliott M, Basset A, Blaber SJM, West RJ. 2012. Paradigms in estuarine ecology – the Remane diagram with a suggested revised model for estuaries: a review. *Estuarine*, *Coastal & Shelf Science*, 97: 78-90.

Williamson P, Turley C, Brownlee C, Findlay HS, Ridgwell A, Schmidt DN, Schroeder DC, Blackford J, Tyrrell T, Pinnegar JK. 2013. Impacts of ocean acidification. *MCCIP Science Review* 2013, 34-48, doi:10.14465/2013.arc05.034-048.

Wittmann AC, Pörtner HO. 2013. Sensitivities of extant animal taxa to ocean acidification. *Nature Climate Change*; DOI: <u>10.1038/nclimate1982</u>

Wong PP, Losada IJ (Coordinating lead authors). 2014. Chapter 5. Coastal Systems and Low-Lying Areas. IPCC WGII AR5 Chapter 5: 85 pp.

Yamanaka T, Raffaelli D, White PCL. 2013. Non-Linear Interactions Determine the Impact of Sea-Level Rise on Estuarine Benthic Biodiversity and Ecosystem Processes. *PLoS ONE*, **8**: e68160.

Zhang K, Douglas BC, Leatherman SP. 2004. Global warming and coastal erosion. *Climatic Change*. 64: 41-58

Zondervan I, Zeebe RE, Rost B, Riebesell U. 2001. Decreasing marine biogenic calcification: A negative feedback on rising atmospheric pCO₂. *Global Biogeochemical Cycles* 15, 507-516.

Table 2 Main Topics relating to the marine consequences of climate change and the way in which they influence the Good Environmental Status Descriptors D1-D11 (cross refer to Figures 2-10; see text and Tables S1 and S2 for Descriptor titles)

Descriptor	D1	D2	D3	D4	D5	D6	D7	D8	D9	D10	D11
Topics											
I Altered temperature	✓	1	1	1		1					
regime – species re-											
distribution and											
community response											
II Altered temperature	✓	1	1	1	1	1					
regime – individual											
physiological/phenological											
response											
III Increased relative sea-	V		1	1		1	1				
level rise - physiographic											
changes											
IV Increased climate	1			/		/	1				
variability effects on											
coastal hydrodynamics											
V Changes to large scale	V		V	1	1	1	V	1	1		
climatic patterns due to											
land run-off											
VI Increased relative sea-	~			1		1	1				
level rise changing											
estuarine hydrodynamics											
VII Increased ocean	V		1	V		V		V	V		
acidification and seawater											
physico-chemical changes											
VIII Loss of polar ice	~	1	1	1	1		1			1	~
cover and global transport											
repercussions						<u> </u>					_
Sum categories	8	3	6	8	3	7	5	2	2	1	1

Table 3 The Rate of change in latitudinal location of representative groups

Organism	Rate of change	Reference
Phytoplankton	469.9 (±115.3) km dec ⁻¹	Poloczanska et al. (2013)
Invertebrate zooplankton	142.1 (±27.8) km dec ⁻¹	Poloczanska et al. (2013)
Copepods	≈500 km dec ⁻¹	Calculated from Beaugrand <i>et al.</i> (2002)
Intertidal biota	50 km dec ⁻¹	Helmuth <i>et al.</i> (2006)
Bony fish	277.5 (±76.9) km dec ⁻¹	Poloczanska et al. (2013)
Plaice (North Sea)	-3.96 m (depth) dec ⁻¹ (1980-	Dulvy et al. (2008)
	2004)	Engelhard et al. (2011)
	142 km NE (1913-2007)	
Sole (North Sea)	+7.64 m (depth) dec ⁻¹ (1980-	Dulvy et al. (2008)
	2004)	Engelhard et al. (2011)
	93 km SE (1913-2007)	

Table 1. Biodiversity-related qualitative Marine Strategy Framework Directive Descriptors and indicators which may be affected by climate change, together with the cause, evidence, precise examples and references

DESCRIPTOR	CRITERIA	INDICATOR	CAUSE OF	PRECISE	REFERENCES
			CHANGE AND	EXAMPLE	
			IMPACT		
1: Biological	1.1 Species distribution	1.1.1 Distributional range	Shift of species	Zostera noltii	Alcock, 2003; Chust et
diversity			distribution, especially	northward	al., 2013b; Duarte et al.,
			at the margins of its	distributional will shift	2013a; Nicolas <i>et al.</i> ,
			distributional range	of 888 km in the	2011; Poloczanska et
			(i.e. change of area	suitable habitat of the	al., 2013; Reid and
			occupied by a species)	species, and a retreat of	Valdés, 2011; Rombouts
				southernmost	et al., 2012; Tasker,
				populations (Valle et	2008; Valle <i>et al.</i> , 2014
				al., 2014)	
		1.1.2 Distributional pattern within the	Boreal species can be	In the North Sea six	Cheung <i>et al.</i> , 2013a, b;
		latter	refuge in deeper	fish species, including	Poloczanska <i>et al.</i> ,
		Tattor	waters, temperate	plaice	2013; Reid and Valdés,
			species can extend to	(Pleuronectes platessa)	2011
			deeper waters	and cuckoo ray	2011
				(Leucoraja naevus),	
				moved deeper with	
				warming but did not	
				change in latitude	
				(Perry et al., 2005)	
		1.1.3 Area covered by the species (for	Reduction of the area	Reduction in Calanus	Birchenough et al.,
		sessile/benthic species)	for boreal species,	finmarchicus available	2011; García <i>et al</i> .,
			increase for temperate	overwintering habitat	2013; Poloczanska et
			and subtropical	northwest of Scotland	al., 2013; Valle et al.,
				(Beaugrand, 2009)	2013
	1.2 Population size	1.2.1 Population abundance and/or	Described increases	Reduction in	Chust <i>et al.</i> , 2013a;
		biomass	and decreases,	abundance and biomass	Finney et al., 2010;

		depending on the resilience of the species, and are generally non-linear Reduction in population size if characteristics of the area becomes sub-optimal	of fish in lower latitudes, increase of biomass at high latitudes (Cheung et al., 2013a, b), reduction in zooplankton and phytoplankton biomasses of 11% and 6%, respectively (Chust et al., 2014a).	Hemery <i>et al.</i> , 2008; Munday <i>et al.</i> , 2013; Nye <i>et al.</i> , 2009; Poloczanska <i>et al.</i> , 2013; Reid and Valdés, 2011; Tasker, 2008
1.3 Population condition	1.3.1 Population demographic characteristics	Early signal of climate change, since it affects first to individual demography Changes to recruitment patterns due to temperature threshold changes; reduction in juvenile stages; changes to spawning thresholds due to temperature change		Albouy et al., 2014; Poloczanska et al., 2013; Tasker, 2008
	1.3.2 Population genetic structure	Better or worst adaptation to change Inflow of sibling species; genetic changes due to aquaculture changes,		Chust et al., 2013a
1.4 Habitat distribution	1.4.1 Distributional range	Shift of habitat (both bed and water column) distribution	Important habitat distributional range shift for macroalgae Pelvetia canaliculata	Martínez et al., 2012; Duarte <i>et al.</i> , 2013a; Reid and Valdés, 2011; Valle <i>et al.</i> , 2014

			(Neiva et al., 2014)	
	1.4.2 Distributional pattern	Boreal habitats can be refuge in deeper waters, temperate habitats can extend to	Changes in macroalgae (Lima <i>et al.</i> 2007)	Reid and Valdés, 2011
1.5 Habitat extent	1.5.1 Habitat area	deeper waters Reduction of the area for boreal species,	Reduction >80 habitat extent in intertidal	Duarte <i>et al.</i> , 2013a; Reid and Valdés, 2011;
	1.5.2 Habitat volume, where relevant	increase for temperate and subtropical No information;	pools and boulders (Thorner <i>et al</i> . 2014)	Valle et al., 2013
	1.5.2 Habitat volume, where relevant	habitat volume as a water mass affected by changes to thermohaline conditions changing through climate change		
1.6 Habitat condition	1.6.1 Condition of the typical species and communities	Reduction of habitat- forming species Change to community composition (but not necessarily guilds and traits represented)	The shift to small bodied, shallow burrowers with opportunistic life histories could caused a reduction or loss of habitat forming species and those that create habitat heterogeneity (Caswell and Frid, 2013)	Hoegh-Guldberg and Bruno, 2010
	1.6.2 Relative abundance and/or biomass, as appropriate	Increase of dominance (by number or biomass) change due to changing species		

		1.6.3 Physical, hydrological and chemical conditions	distributions and relative proportion in the community Many evidences of sea temperature increase, acidification increase, and sea-level rise Changes to temperature regime, storminess, salinity changes due to run-off		Stocker et al., 2013
	1.7 Ecosystem structure	1.7.1 Composition and relative proportions of ecosystem components (habitats, species)	Important changes in structure and function Changes to relative amounts of different communities in the ecosystem; changes to bentho-pelagic coupling	Significant changes in structure and lower diversity in response to reduced pH and increasing temperature. Molluscs are the most affected and annelids the less (Hale <i>et al.</i> , 2011)	Hoegh-Guldberg and Bruno, 2010; Reid and Valdés, 2011; Sydeman and Bograd, 2009; Tomczak <i>et al.</i> , 2013; Yamanaka <i>et al.</i> , 2013
2: Non- indigenous species	2.1 Abundance and state of non-indigenous species, in particular invasive species	2.1.1 Trends in abundance, temporal occurrence and spatial distribution of non-indigenous species	Increase of abundance for invasive species; increase in migration of species from outside the area	There is high confidence in which invasive subtropical species will increase at temperate latitudes (Wong and Losada, 2014)	Bremner, 2008; Dutertre et al., 2010; Hellmann et al., 2008; Lejeusne et al., 2010; Mendoza-González et al., 2013; Occhipinti-Ambrogi, 2007; Otero et al., 2013; Tasker, 2008
	2.2 Environmental impact of invasive non-indigenous sp.	2.2.1 Ratio between invasive non-indigenous species and native species	Ratio will increase; increasing naturalisation of non- native species	Introductions and shift are faster in marine than in terrestrial systems (Sorte <i>et al.</i> , 2010), this likely	Occhipinti-Ambrogi, 2007; Otero et al., 2013)

				producing increasing ratios	
		2.2.2 Impacts of non-indigenous invasive species at the level of species, habitats and ecosystem	Increase of vulnerability of species and habitats, impacts on ecosystem functionality, changes due to bio-engineer properties of invasive species		Birchenough <i>et al.</i> , 2011; Bremner, 2008; Occhipinti-Ambrogi, 2007; Ramirez-Llodra <i>et al.</i> , 2011
4: Food webs	4.1 Productivity of key species or trophic groups	4.1.1 Performance of key predator species using their production per unit biomass	Key top-predator species will be more vulnerable (e.g. cod, cetaceans, seabirds) Changes to energetic of certain species as shown by P/B ratios, result of physiological changes due to temperature changes		Albouy et al., 2014; Jennings and Brander, 2010; Niiranen et al., 2013; Ottersen et al., 2010
	4.2 Proportion of selected species at the top of food webs	4.2.1 Large fish (by weight)	Lower size, loss of many large fish; results of bioenergetic changes	Fish will have smaller body weight and local extinction/decreased abundance of larger-bodied species (Cheung <i>et al.</i> , 2013a, b)	Albouy et al., 2014; Jennings and Brander, 2010; Ottersen et al., 2010
	4.3 Abundance/distribution of key trophic groups/species	4.3.1 Abundance trends of functionally important selected groups/species	Loss of feeding links, simplification of food webs, changes in the up and down control of the food webs, change from phytoplankton to		Albouy et al., 2014; Cury et al., 2008; Defeo and Castilla, 2012; Drinkwater et al., 2010; Eriksson Wiklund et al., 2009; Hoegh-Guldberg

6: Seafloor integrity	6.1 Physical damage, having regard to substrate characteristics	6.1.1 Type, abundance, biomass and areal extent of relevant biogenic substrate	Interactions between human pressures and climate change, increasing damage or reducing resilience		and Bruno, 2010; Niiranen et al., 2013; Ottersen et al., 2010; Rombouts et al, 2013; Travers et al., 2007 Borja et al., 2013b; Hewitt and Thrush, 2010; Wernberg et al., 2011
		6.1.2 Extent of the seabed significantly affected by human activities for the different substrate types	Interactions between human pressures and climate change, increasing damage or reducing resilience	Exploited macroalgae beds or beds affected by discharges are less resilient to climate change than unaffected macroalgae (Borja <i>et al.</i> , 2013b)	Borja et al., 2013b; Hewitt and Thrush, 2010; Wernberg et al., 2011
	6.2 Condition of benthic community	6.2.1 Presence of particularly sensitive and/or tolerant species	Changes in the presence of sensitive species (especially temperature-sensitive)	Warming induces stratification, oxygen consumption and community shift toward lower species richness and increase of hypoxia-tolerant (Pörtner and Karl, 2014) and opportunistic species (Caswell and Frid, 2013)	Otero <i>et al.</i> , 2013; Tasker, 2008
		6.2.2 Multi-metric indices assessing benthic community condition and	Changes in current reference conditions	From the previous example, it is expected	Nicolas et al., 2011

functionality, such as species diversity and richness, proportion of opportunistic to sensitive species	and potential shifts in results from those indices (especially those using structural metrics)	that the ratio sensitive/opportunistic species will be reduced and, as such, the ecological status will be worst	
6.2.3 Proportion of biomass or number of individuals in the macrobenthos above specified length/size	Reduction of richness and biomass in some cases		Beukema, 2002; Birchenough <i>et al.</i> , 2011; Junker <i>et al.</i> , 2012; Yamanaka <i>et al.</i> , 2013
6.2.4 Parameters describing the characteristics of the size spectrum of the benthic community	Probable changes in body size spectra, towards a reduction	Expected reduction in body size, as shown in palaeo records (Caswell and Frid, 2013)	Levin et al., 2009; Webb et al., 2009

Figure(s)

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Figure 1: Figure 1: A conceptual model of the implementation of the MSFD (inner blue circle) together with the areas for caution as the result of global climate change (red boxes) (see text).

Figure 2 Primary drivers and consequences of marine global climate change (cross-referring to other figures)

Figure 3 Species re-distribution and community response due to altered temperature regime (MSFD Descriptor denoted in brackets, see text)

Figure 4 Physiological and phonological responses due to an altered temperature regime leading to ecosystem effects (MSFD Descriptor denoted in brackets, see text)

Figure 5 Physiographic changes due to increased relative sea level leading to ecosystem effects (MSFD Descriptor denoted in brackets, see text)

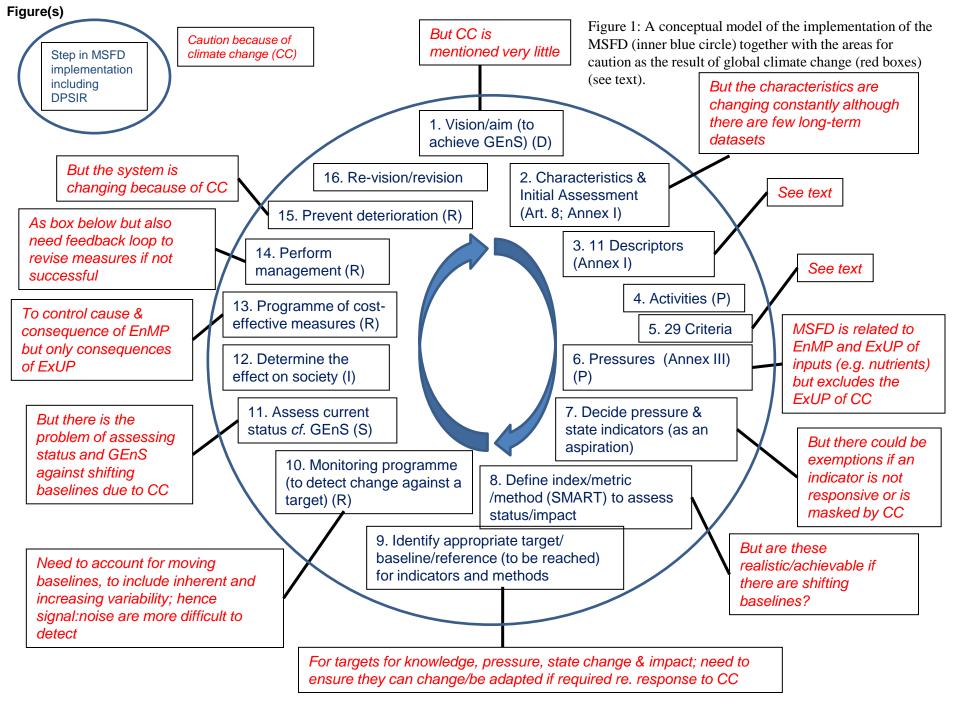
Figure 6 Coastal hydrodynamic changes due to increased climate variability leading to ecosystem effects (MSFD Descriptor denoted in brackets, see text)

Figure 7 Land-based discharges and run-off due to regional climate perturbations leading to ecosystem effects (MSFD Descriptor denoted in brackets, see text)

Figure 8 Estuarine hydrodynamic changes due to increased relative sea levels leading to ecosystem effects (MSFD Descriptor denoted in brackets, see text)

Figure 9 Physico-chemical water changes due to decreased pH and increased CO₂ levels leading to ecosystem effects (MSFD Descriptor denoted in brackets, see text)

Figure 10 Global transport repercussions due to loss of polar ice-cover leading to ecosystem effects (MSFD Descriptor denoted in brackets, see text)



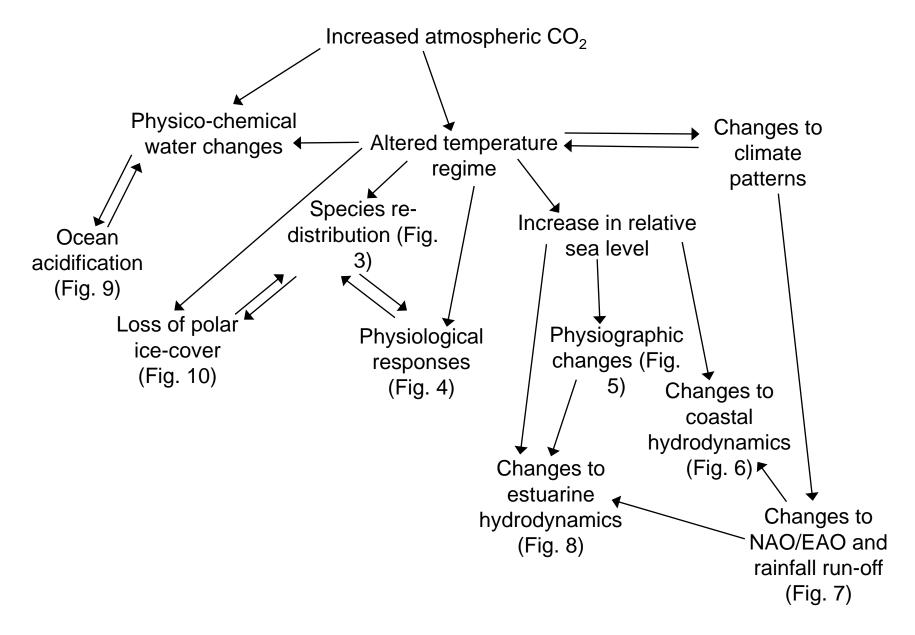


Figure 2 Primary drivers and consequences of marine global climate change (cross-referring to other figures)

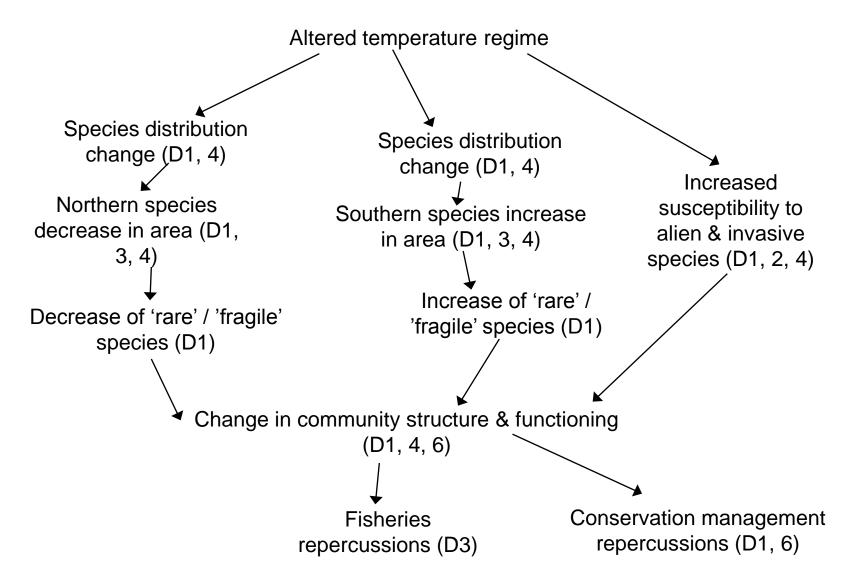


Figure 3 Species re-distribution and community response due to altered temperature regime (MSFD Descriptor denoted in brackets, see text)

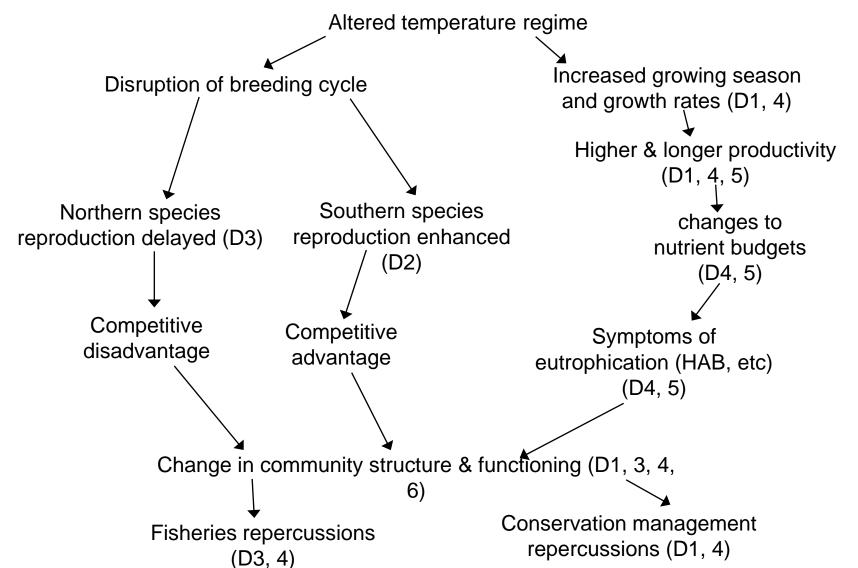


Figure 4 Physiological and phonological responses due to an altered temperature regime leading to ecosystem effects (MSFD Descriptor denoted in brackets, see text)

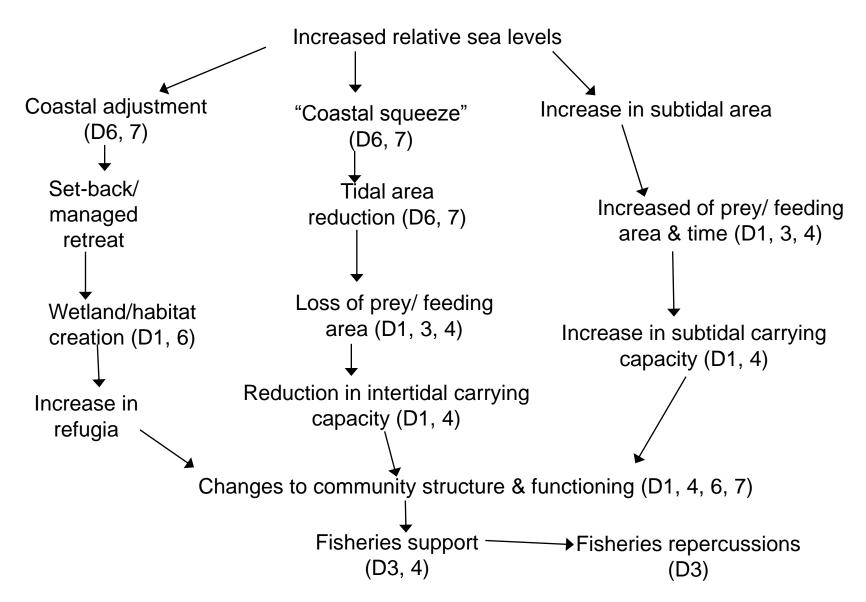


Figure 5 Physiographic changes due to increased relative sea level leading to ecosystem effects (MSFD Descriptor denoted in brackets, see text)

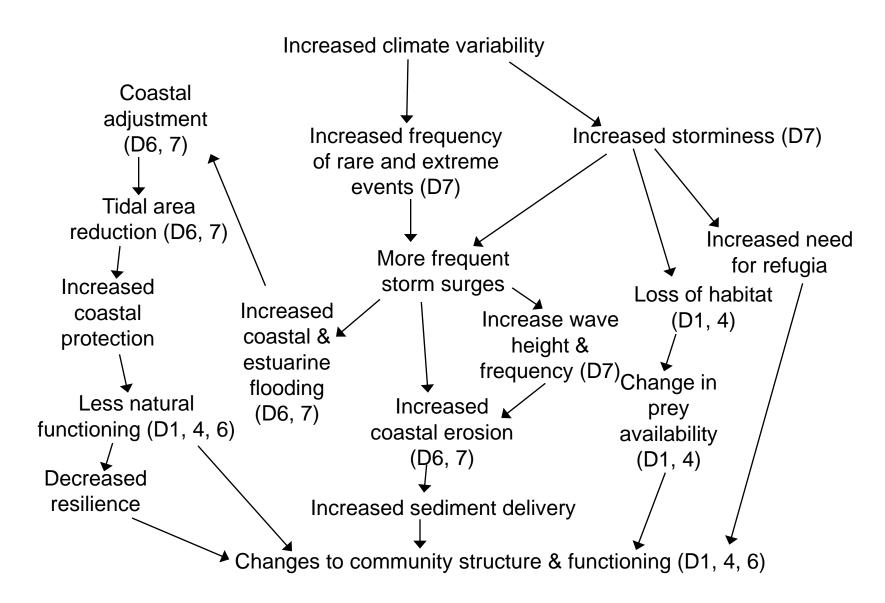


Figure 6 Coastal hydrodynamic changes due to increased climate variability leading to ecosystem effects (MSFD Descriptor denoted in brackets, see text)

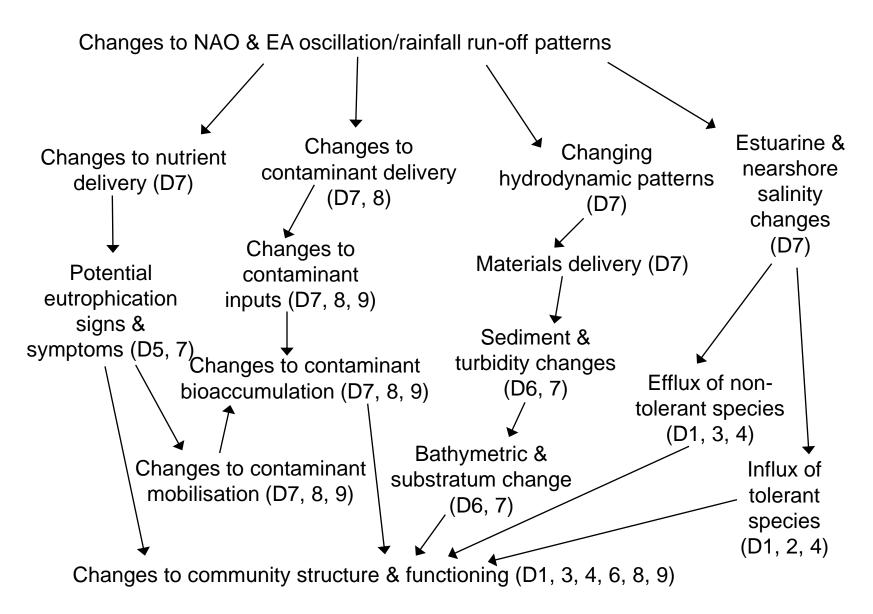


Figure 7 Land-based discharges and run-off due to regional climate perturbations leading to ecosystem effects (MSFD Descriptor denoted in brackets, see text)

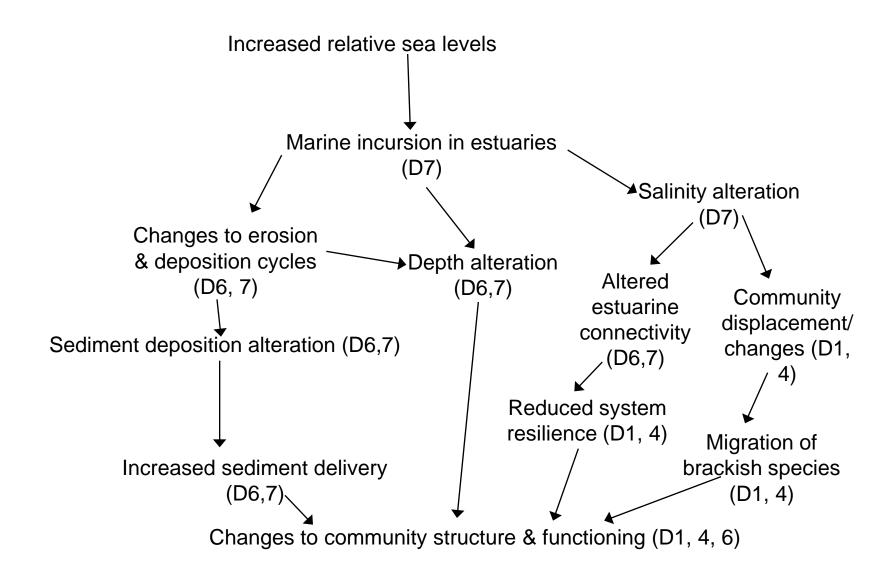


Figure 8 Estuarine hydrodynamic changes due to increased relative sea levels leading to ecosystem effects (MSFD Descriptor denoted in brackets, see text)

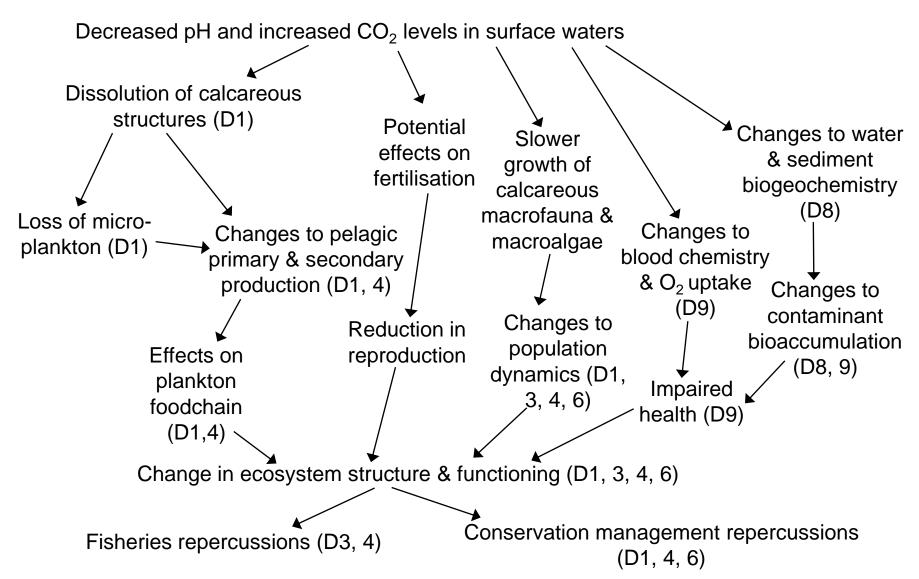


Figure 9 Physico-chemical water changes due to decreased pH and increased CO₂ levels leading to ecosystem effects (MSFD Descriptor denoted in brackets, see text)

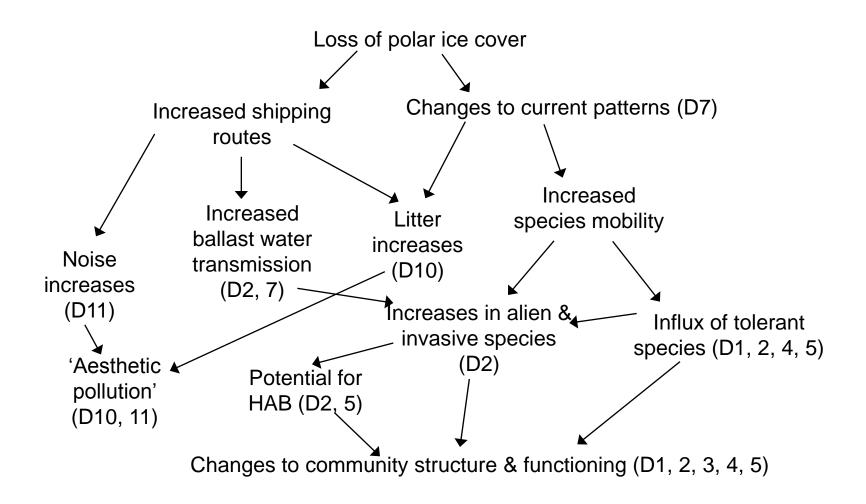


Figure 10 Global transport repercussions due to loss of polar ice-cover leading to ecosystem effects (MSFD Descriptor denoted in brackets, see text)