

Tracking Cancer Cells with Microfluidic High Frequency DEP Cytometer Implemented on BiCMOS Lab-on-Chip Platform

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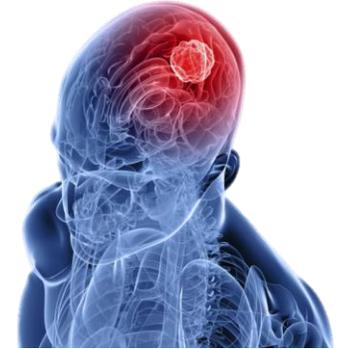
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Motivation

- Need for new therapeutic strategies dedicated to poor outcome diseases
 - ▶ *Tumor with high recurrence*
 - ▶ *Strong resistance to existing treatments*
 - ▶ *Highly heterogeneous brain tumors*



Ex: Glioblastoma:

Resulting efficiency from standard therapies is very low

- ➔
- ☹️ *Poor patient survival rate*
 - ☹️ *Frequent relapse*

Role of some hidden tumor-initiating cells ?

How fight them more efficiently?

What they look like?

How many are they?

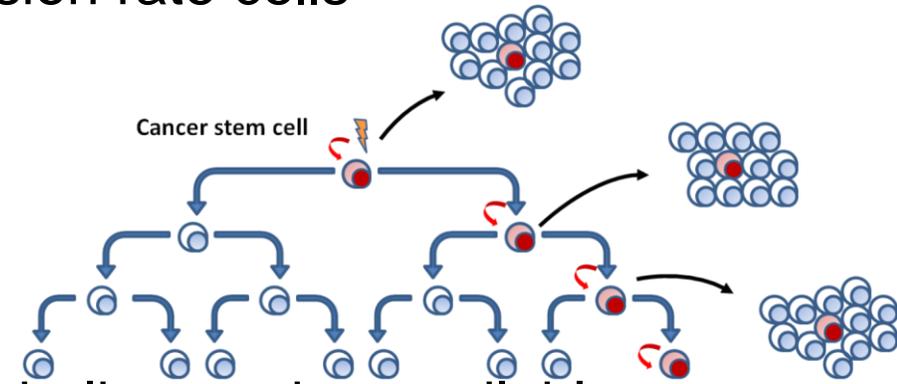
Where are they?

Motivation

- Need for alternative tools able to track such specific and rare cells

Cancerous Stem Cells: *Tumorigenic cells with ability to give rise to all tumor cell type*

- ▶ Quiescent cells: escape from therapies targeting high division rate cells
- ▶ Differentiation into multiple cell types (progenitors...)
- ▶ Self-renewal capabilities
- ▶ Low number, Hidden in the tumor
- ▶ Undifferentiated cells: No specificity: lacking for specific labeling marker available

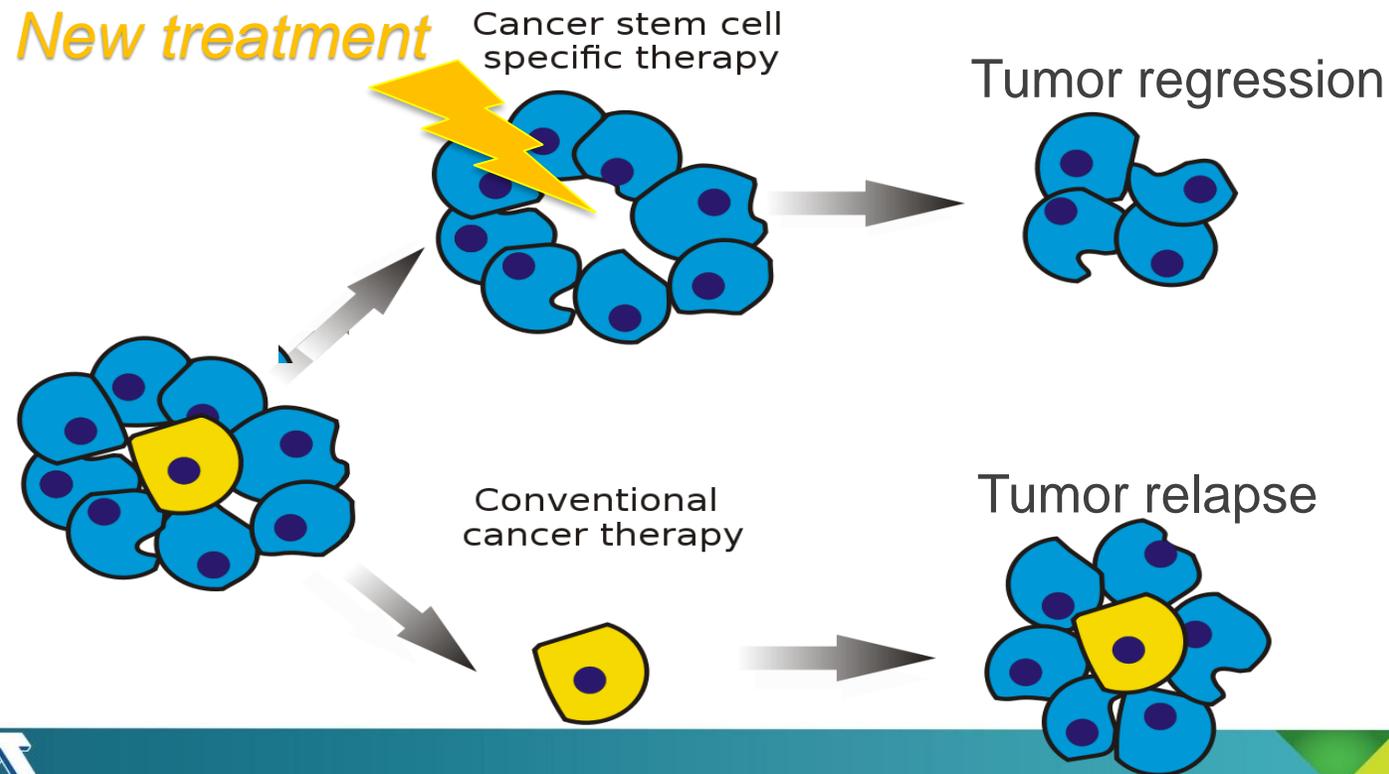


➡ Currently hypothesized to be the main cause of **relapse** and **metastasis**

Motivation

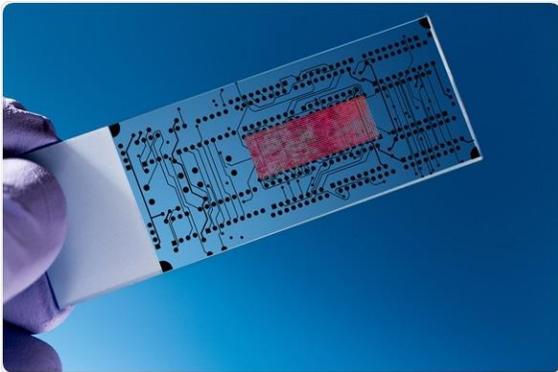
Tools able to identify CSC's in/outside the tumor might contribute to:

- help diagnosis and favor more appropriated treatment
- promote to the development of more efficient therapies



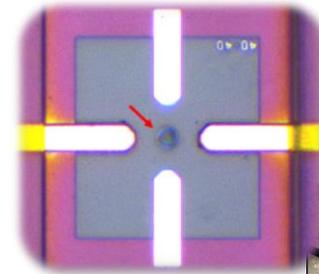
Our approach

- Developing new Generation of Lab-on-Chip for Cancerous Stem Cells identification and sorting using **Electromagnetic Waves**



Proposed concept: Exploit the penetration of **EM radiations** inside the cells to:

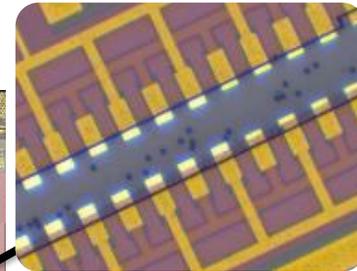
- **sense** their content
- **electromanipulate** them accordingly to their own dielectric property specificities



Individual Cell sensor



Prototype of microfluidic sensing platform on CMOS chip



Electromagnetic based Cytometer

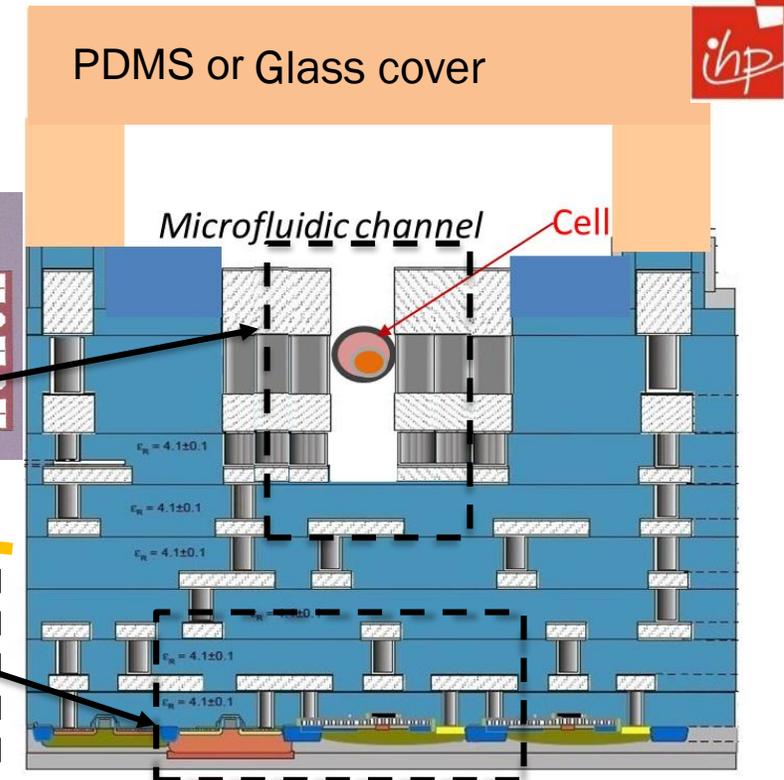
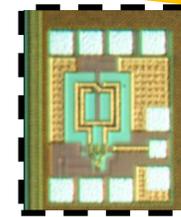
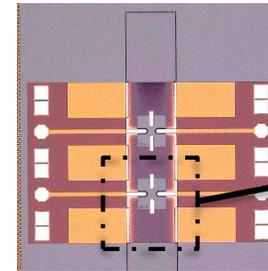
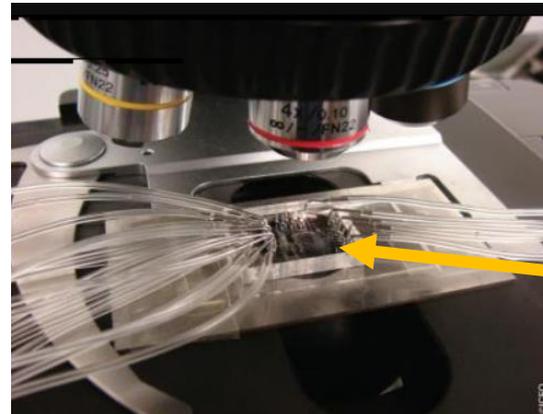
Taking advantage of BiCMOS technology

- ✓ Complete system integration with several electronic functions on the same chip
- ✓ Miniaturization of the complete device And coupling with microfluidics

► Mature technology able to quickly address a large market



► On the way to fully instrumented Lab-on-Chip



Outline

- Motivation and pursued approach
- How characterizing biological cells with high frequency DEP
- High frequency DEP Electromagnetic cytometer design
- First experimental results
- Conclusion and futures developments

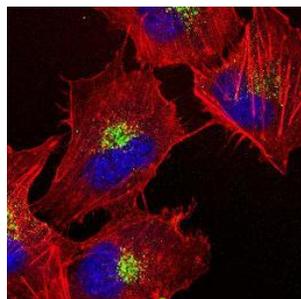
How biologists can study CSC's?



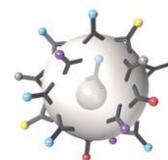
Optical microscopy



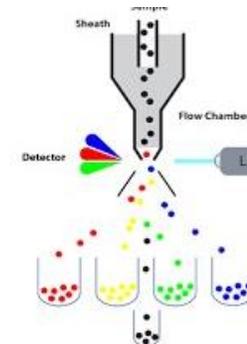
Staining



Fluorescence labeling



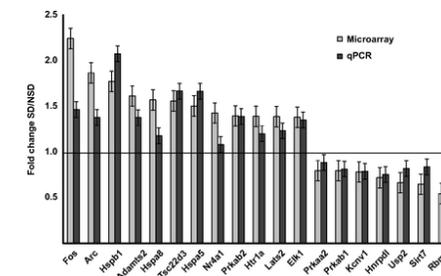
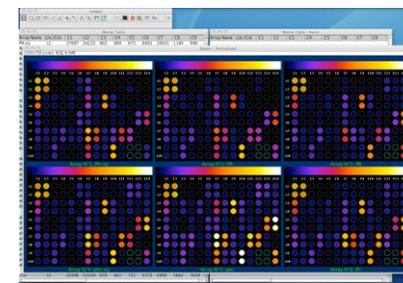
Flow cytometry



QPCR & Protein Array analysis

Drawback/ constrains:

- ✓ CSC's are rare -> require amplification of the population
- ✓ Specific label are lacking -> Cross coupling of generic markers
- ✓ Efficient functional tests exist (clonogenicity, animal drafting) but results are very long

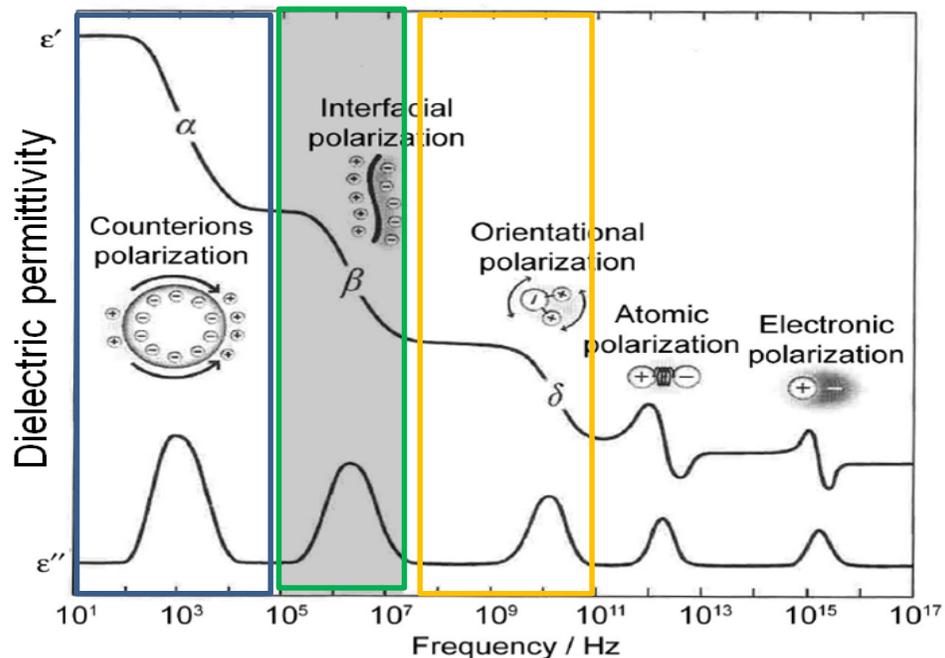


➡ Others approaches investigating intracellular specificities?

What about using EM field to characterize cells?

Depending the frequency EM field could interact with different cell constituents

- Low frequency -> Cell shape/ morphology/size influence
 - Mid frequency -> Plasma Membrane specificities
 - High frequency -> Intracellular content properties



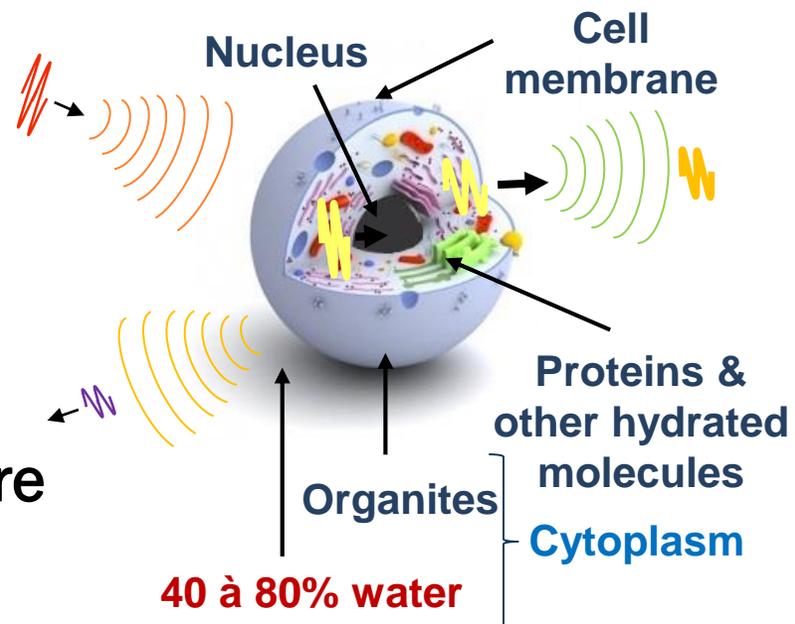
Own cell dielectric properties = A signature that can be specific



High frequency signal well suitable to access to cell interior properties and measure specificities



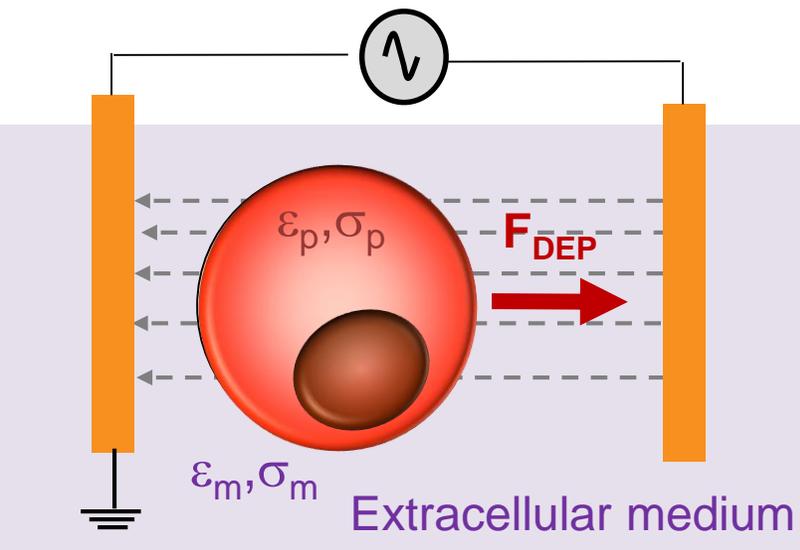
Dielectric spectroscopy allows non destructive & label free characterization



Dielectrophoresis vs Dielectric Spectroscopy approach

DEP relies on the fact that EM fields generate forces that can move cells

Basic DEP theory $F_{DEP} = 2\pi\epsilon_m r^3 \text{Re}[K(\omega)] \nabla|E_{rms}|^2$ ← Related to the E field gradient intensity



$-1 < \text{Re}[K(\omega)] < 1$

$$K(\omega) = \left(\frac{\epsilon_p^* - \epsilon_m^*}{\epsilon_p^* + 2\epsilon_m^*} \right)$$

$$\epsilon_p^* = \epsilon_p - j \frac{\sigma_p}{\omega}$$

Claussius-Mossotti factor

Complex permittivity of the particle

$\text{Re}[K(\omega)] < 0$

Repulsive force

$\text{Re}[K(\omega)] > 0$

Attractive force

➔ Cells can be individually electromanipulated accordingly their own dielectric properties

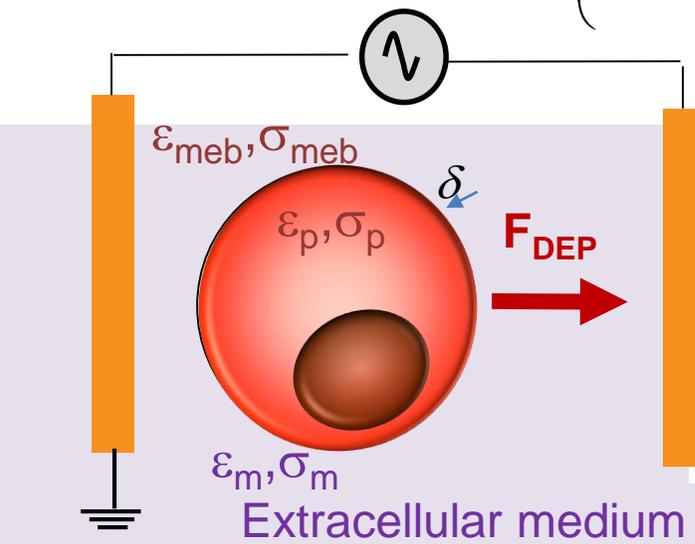
Dielectrophoresis vs Dielectric Spectroscopy approach

Claussius-Mossotti factor also depends on the DEP signal frequency

Few basics

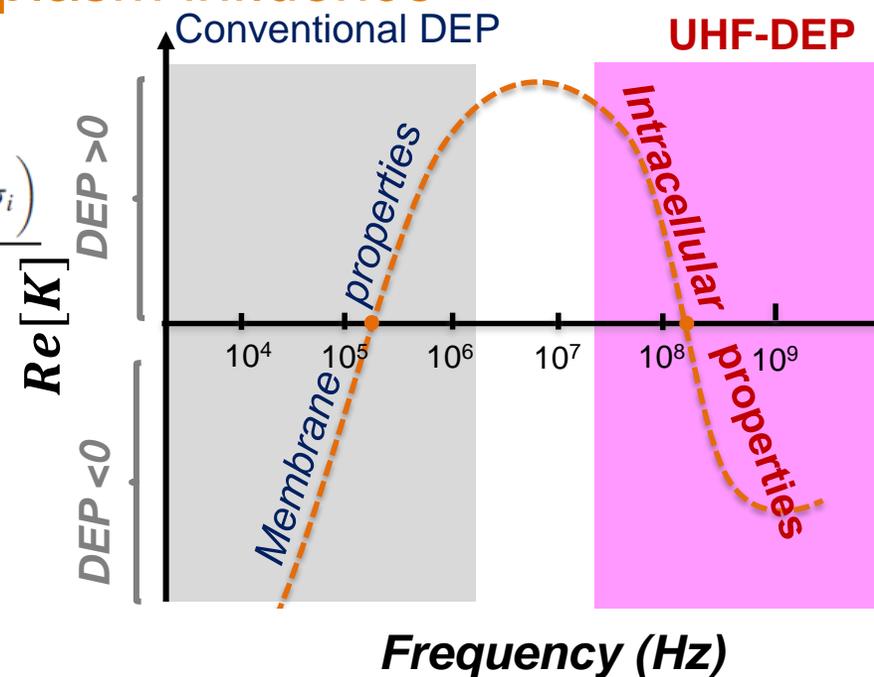
$$\text{Re}[K(\omega)] = \frac{(\epsilon_p - \epsilon_m)(\epsilon_p + 2\epsilon_m) + \frac{1}{\omega^2}(\sigma_p - \sigma_m)(\sigma_p + 2\sigma_m)}{(\epsilon_p + 2\epsilon_m)^2 + \frac{1}{\omega^2}(\sigma_p + 2\sigma_m)^2}$$

As $\omega \nearrow$, plasma membrane influence \searrow
& cytoplasm influence \nearrow



$$\frac{\epsilon_p}{\epsilon_0} = \frac{\left(\frac{\epsilon_m}{\epsilon_0} \frac{\epsilon_i}{\epsilon_0} - \frac{\sigma_m \sigma_i}{\omega^2 \epsilon_0^2}\right) \left(\frac{\epsilon_m}{\epsilon_0} + \frac{\delta}{R} \frac{\epsilon_i}{\epsilon_0}\right) + \frac{1}{\omega^2 \epsilon_0^2} \left(\sigma_m \frac{\epsilon_i}{\epsilon_0} + \frac{\epsilon_m}{\epsilon_0} \sigma_i\right) \left(\sigma_m + \frac{\delta}{R} \sigma_i\right)}{\left(\frac{\epsilon_m}{\epsilon_0} + \frac{\delta}{R} \frac{\epsilon_i}{\epsilon_0}\right)^2 + \frac{1}{\omega^2 \epsilon_0^2} \left(\sigma_m + \frac{\delta}{R} \sigma_i\right)^2}$$

$$\sigma_p = \frac{-\left(\frac{\epsilon_m}{\epsilon_0} \frac{\epsilon_i}{\epsilon_0} - \frac{\sigma_m \sigma_i}{\omega^2 \epsilon_0^2}\right) \left(\sigma_m + \frac{\delta}{R} \sigma_i\right) + \left(\sigma_m \frac{\epsilon_i}{\epsilon_0} + \frac{\epsilon_m}{\epsilon_0} \sigma_i\right) \left(\frac{\epsilon_m}{\epsilon_0} + \frac{\delta}{R} \frac{\epsilon_i}{\epsilon_0}\right)}{\left(\frac{\epsilon_m}{\epsilon_0} + \frac{\delta}{R} \frac{\epsilon_i}{\epsilon_0}\right)^2 + \frac{1}{\omega^2 \epsilon_0^2} \left(\sigma_m + \frac{\delta}{R} \sigma_i\right)^2}$$




 DEP also allows characterizing cell dielectric properties Cell DEP spectral signature

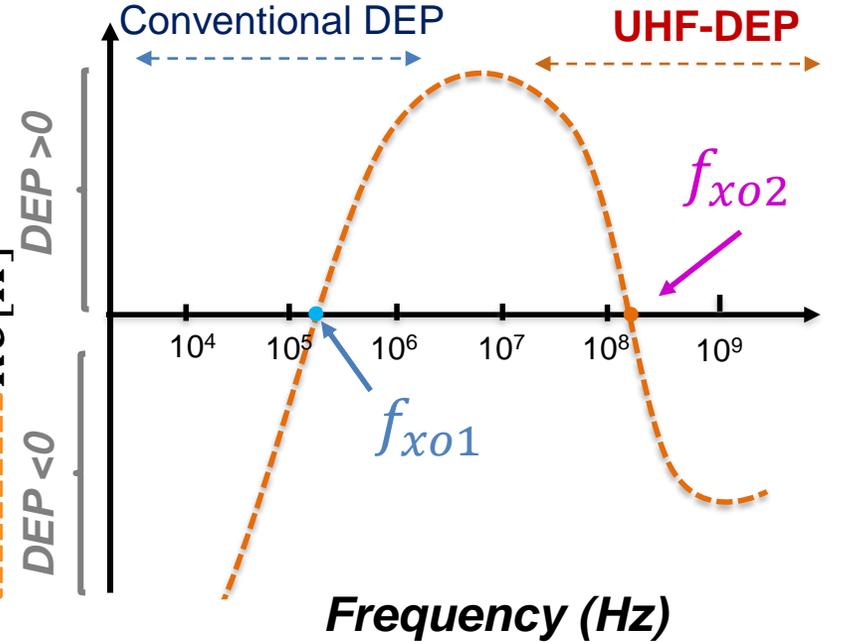
How taking advantage of Cell DEP spectral signature

Once cells are suspended in low σ medium

Two *crossover frequencies* appears $\rightarrow \text{Re}[K]=0$

$$f_{x01} = \frac{\sqrt{2}}{2\pi r C_{mem}} \sigma_m \leftarrow \text{Dominated by membrane dielectric properties}$$

$$f_{x02} = \frac{1}{2\pi} \sqrt{\frac{\sigma_{int}^2 - \sigma_{int}\sigma_m - 2\sigma_m^2}{2\varepsilon_m^2 - \varepsilon_{int}\varepsilon_m - \varepsilon_{int}^2}} \leftarrow \text{Depend on intra vs extra cellular dielectric properties}$$

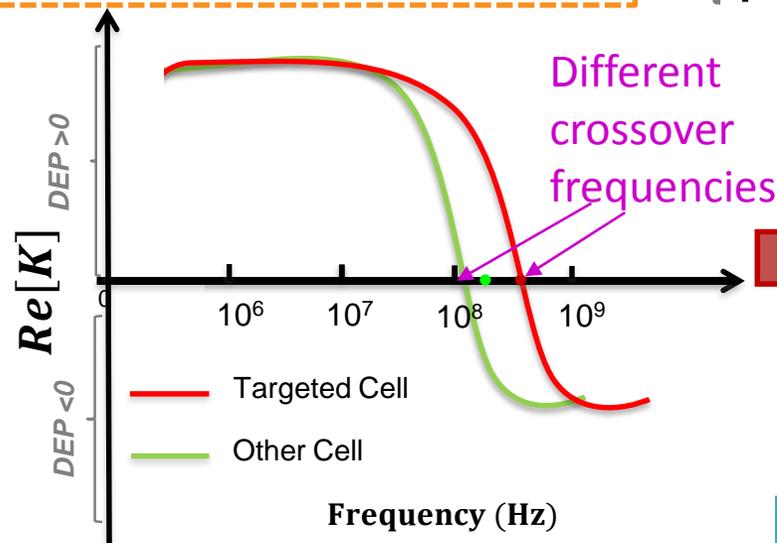


Different cells



Vs

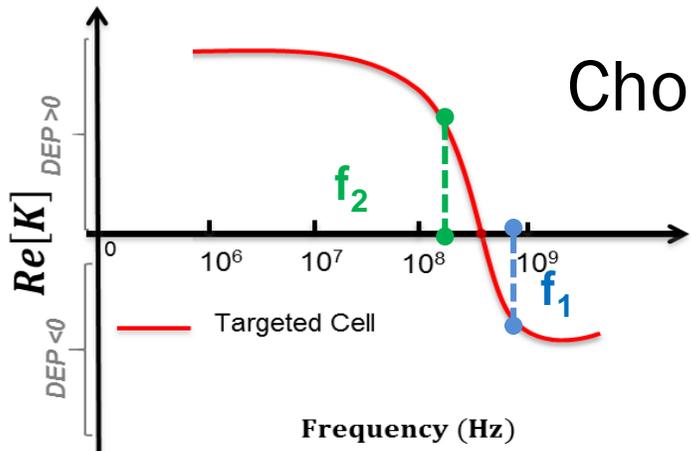
Different spectral signatures



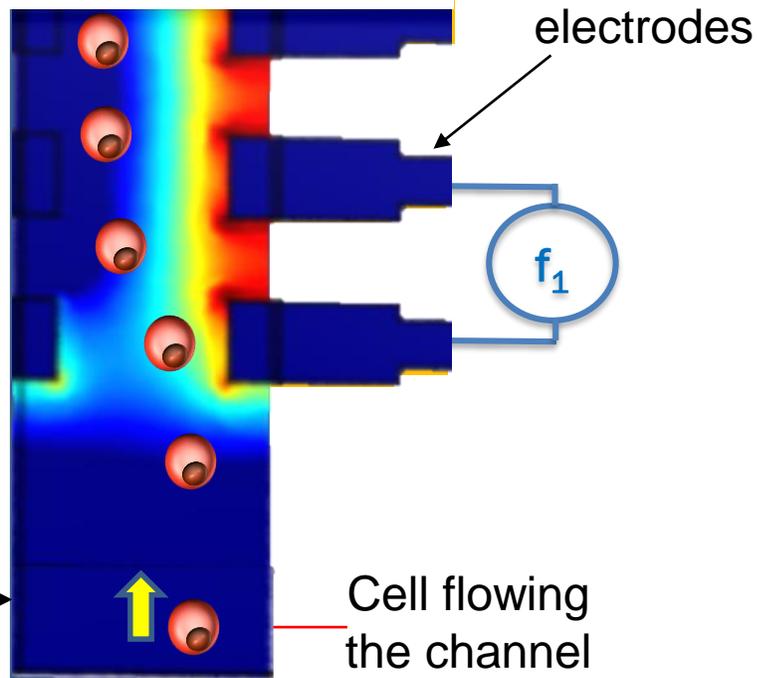
Characterize cells to identify their DEP cross over frequencies as discriminant specificities

Combined electromanipulation & characterization

Choice of appropriated DEP signal frequency is important

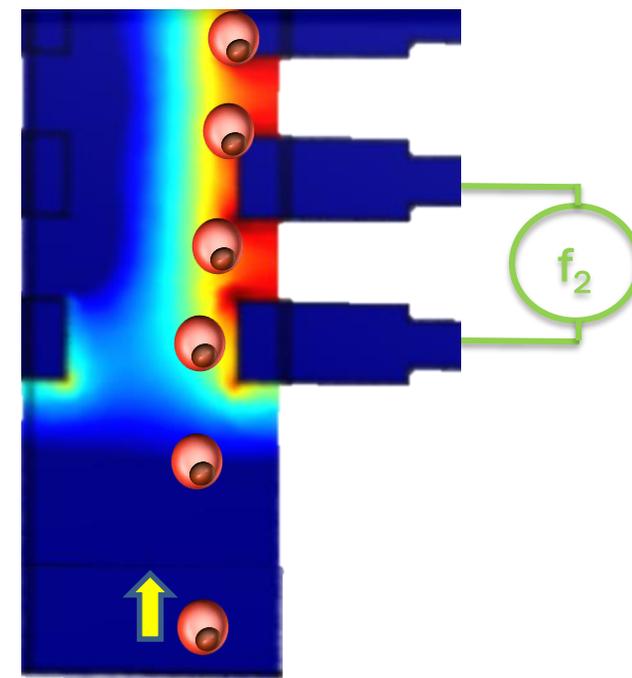


Negative DEP



Cell is *repelled* from electrode

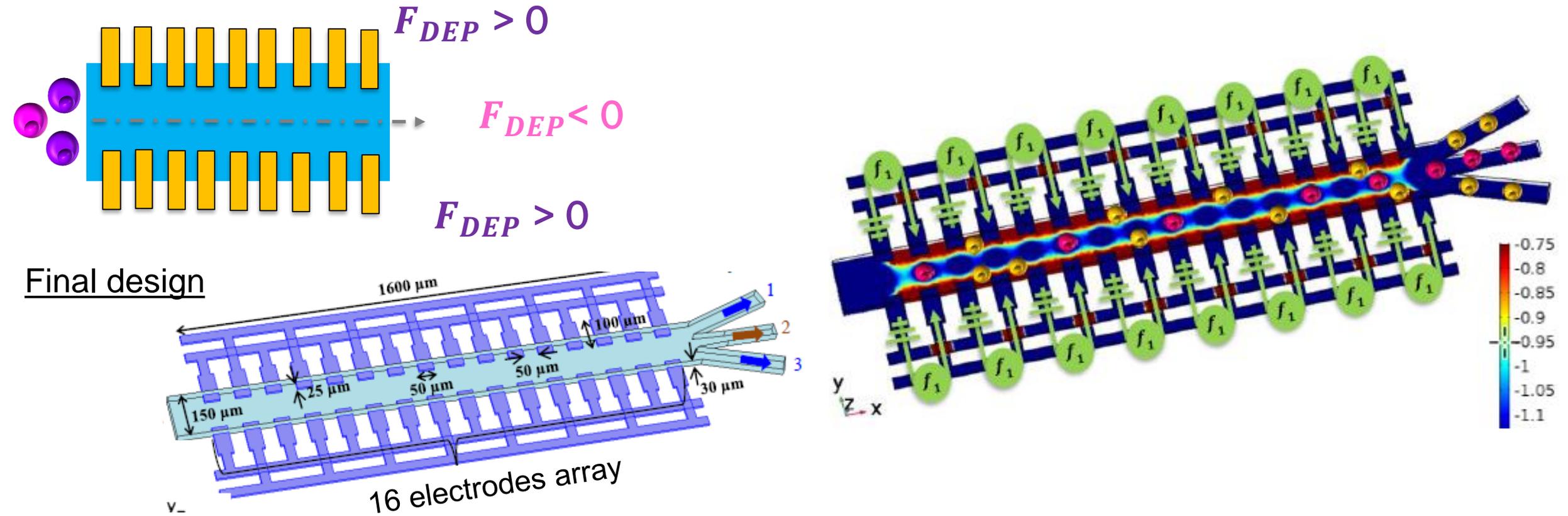
Positive DEP



Cell is *attracted* by electrodes

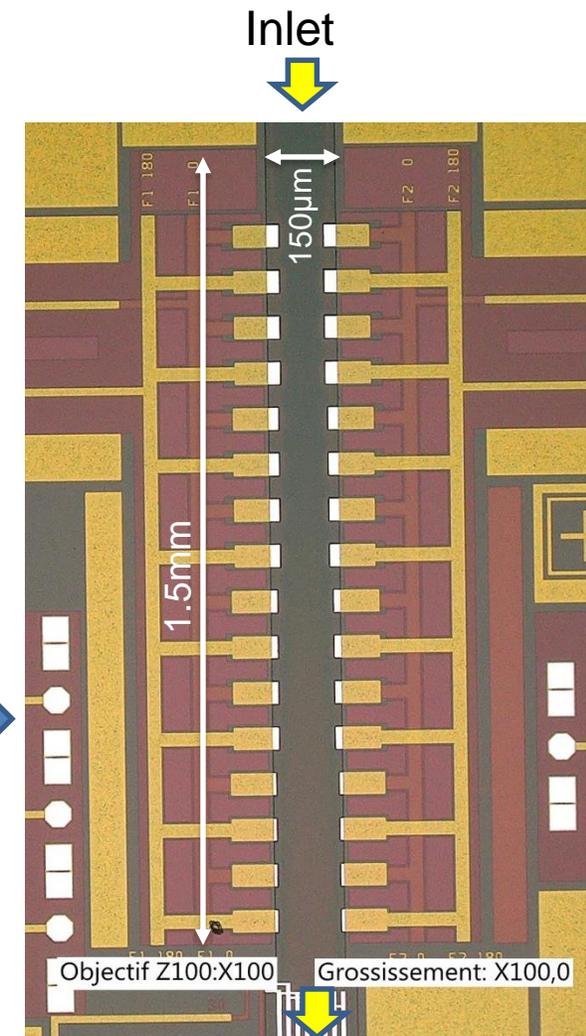
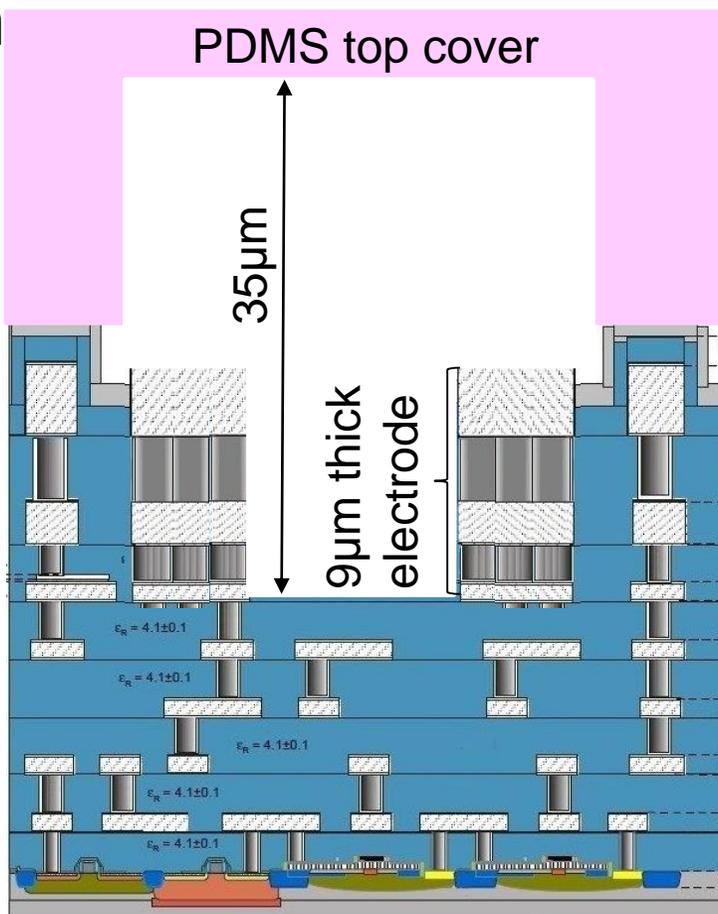
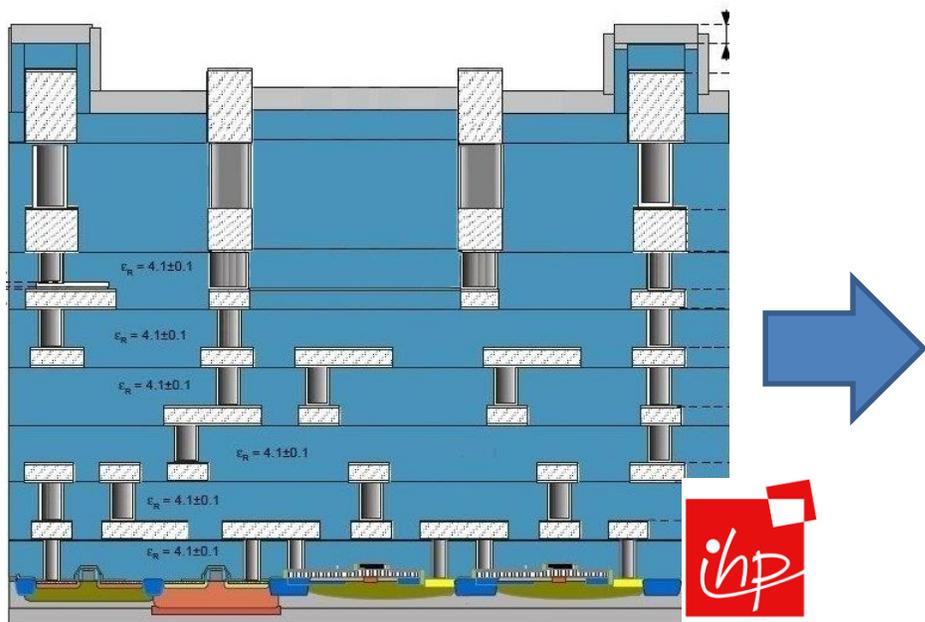
Proposed High Frequency DEP cytometer design

- Double Array of electrode allowing progressive cell trajectory deviation
- Optimization of electrode penetration inside channel and periodicity

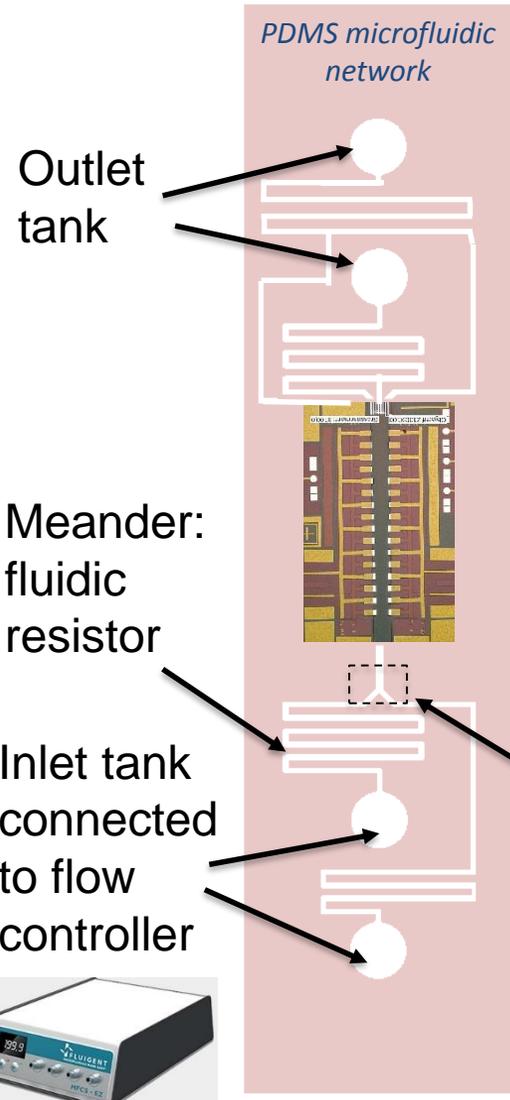


Implementation on BiCMOS technology

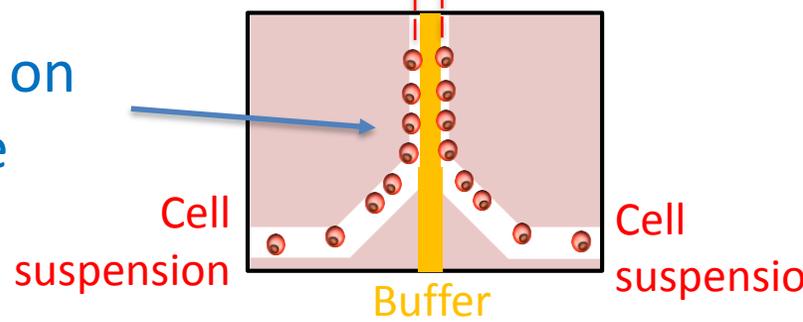
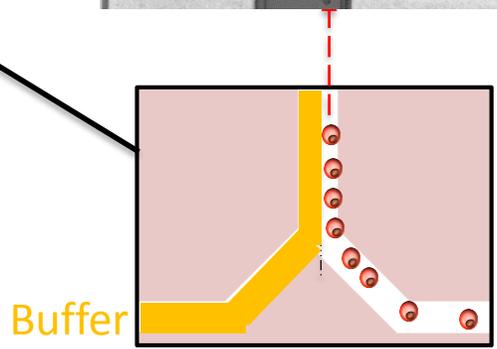
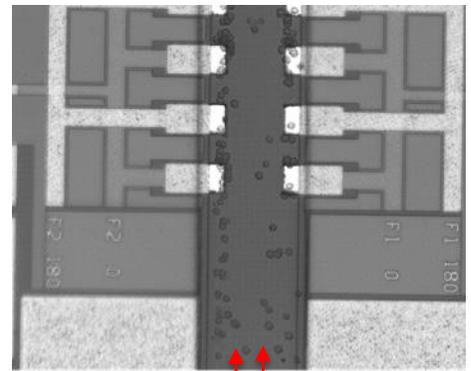
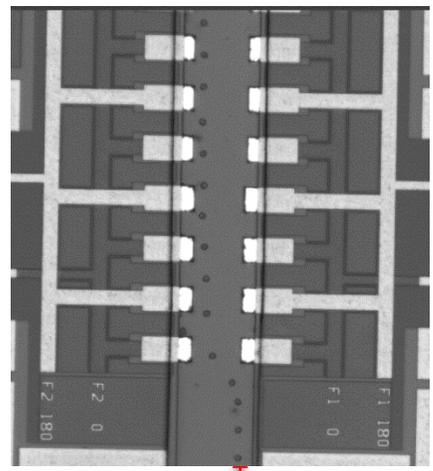
Starting with the BEOL stack IHP SG25H4 process
 + Post process μ channel etch
 + PDMS packaging



Microfluidic network implementation



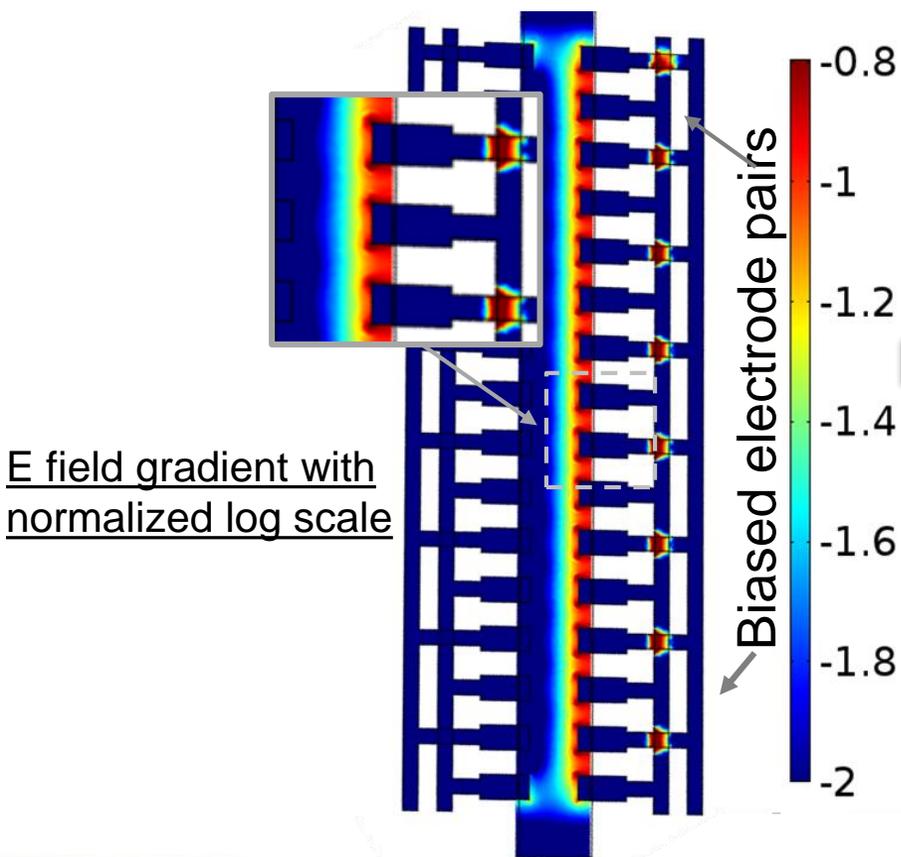
- Outside chip fluidic network for optimal cell injection/collect and velocity control
 - Flow focusing techniques to improve DEP effect



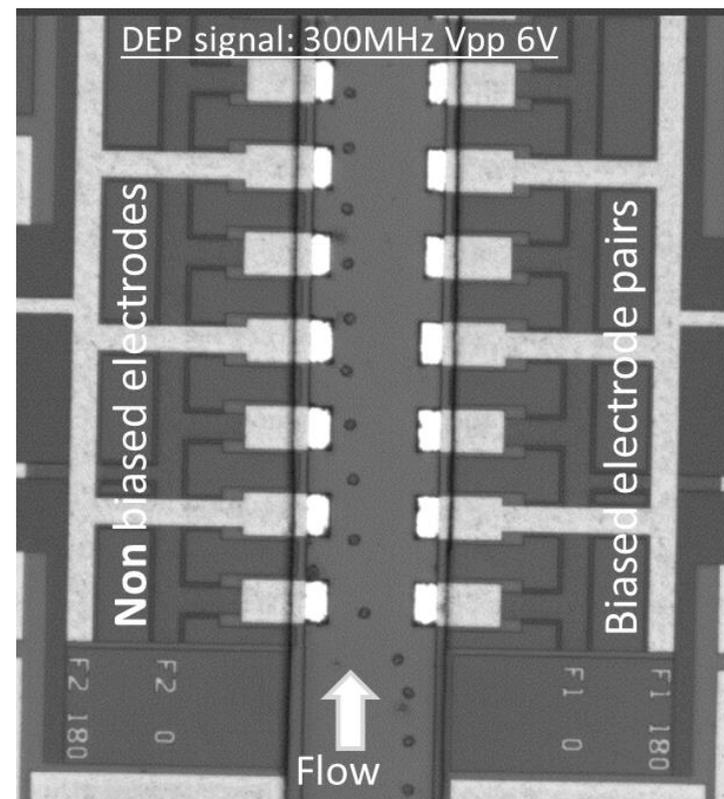
Working principle experimental validation

Test on LN 18 cells with single biased electrode array

- ✓ DEP signal frequency set to $DEP < 0$
- ✓ Cell velocity slow to 0.1mm/s



Cell is well quickly
repealed far the high
intensity E field
region to finally travel
in low intensity one

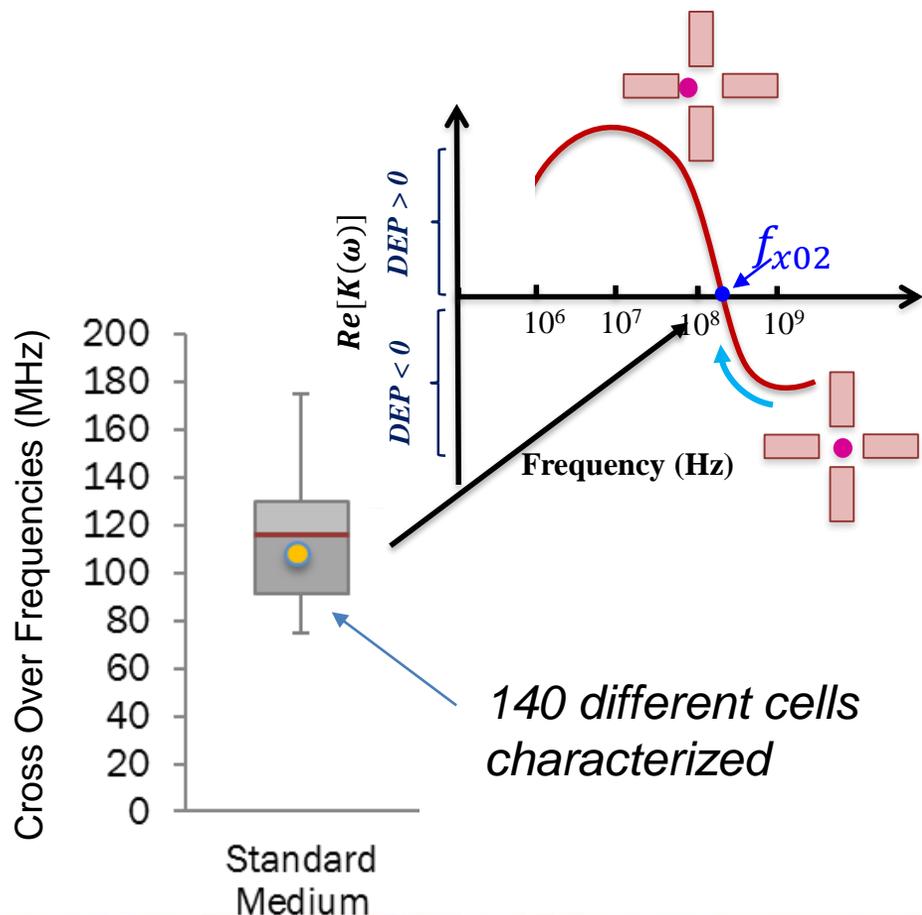


Stacked image of LN 18cell trajectory deviation

DEP characterization of GBM cell line

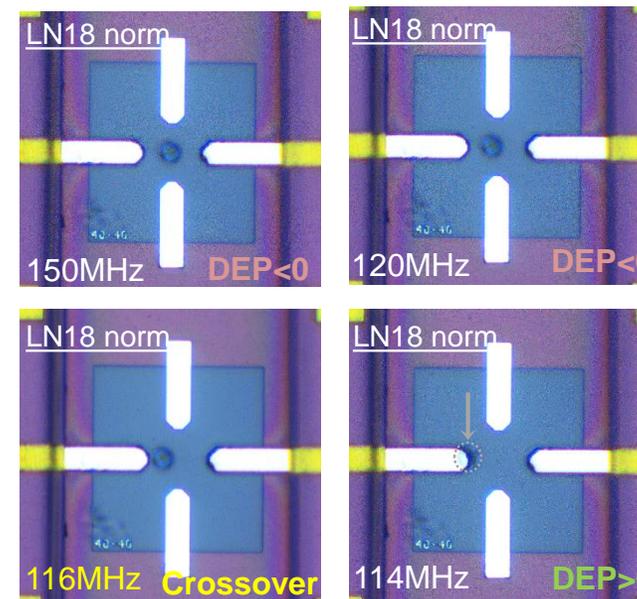
LN 18 cells cultured in conventional culture medium

➤ Specific quadrupole sensor for measuring crossover frequency



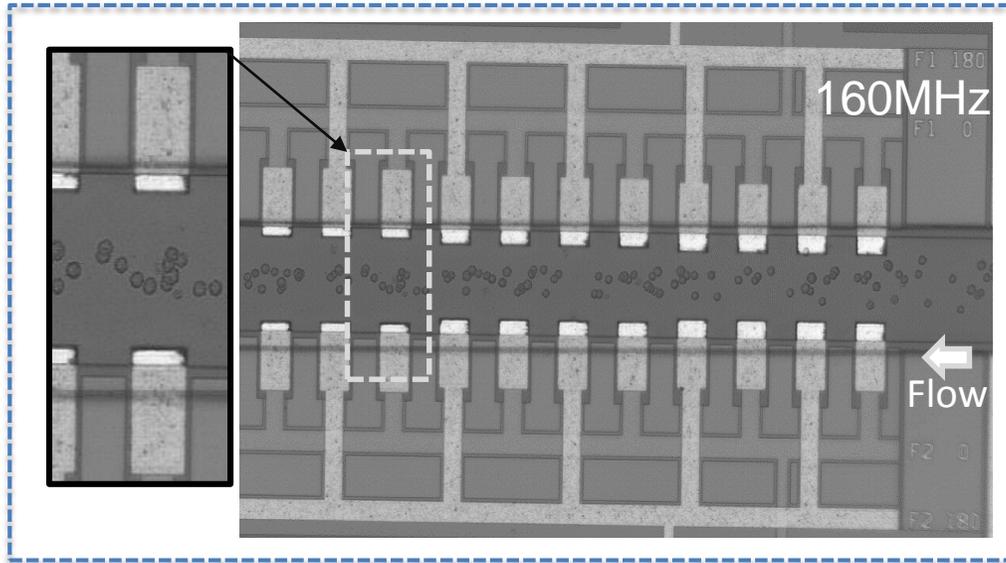
Methodology:

- 1) Cells are trapped in DEP < 0
- 2) Frequency is tuned until finding positive DEP

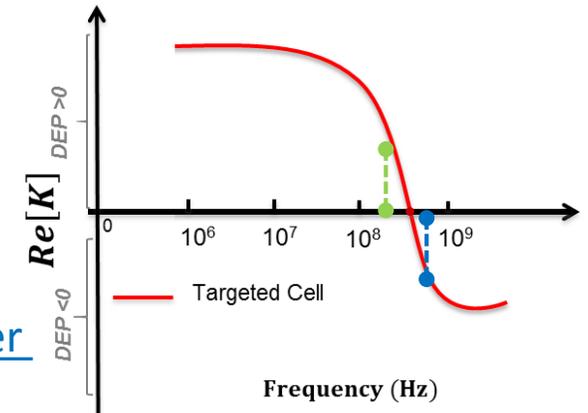


Discrimination of Glioblastoma Cancer Stem Cells by Measuring Their UHF-Dielectrophoresis Crossover Frequency, IMBIOC 2018

Sorting test on LN18 cells



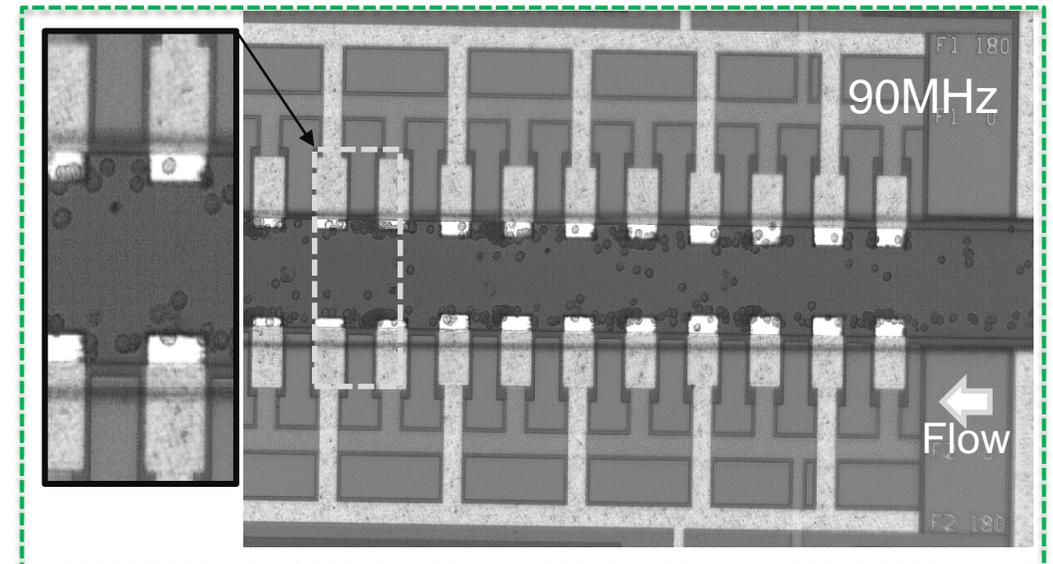
For $F_{DEP} > F_{cross-over}$



All cells are concentrated in microchannel center where E field intensity is the lowest

For $F_{DEP} < F_{cross-over}$

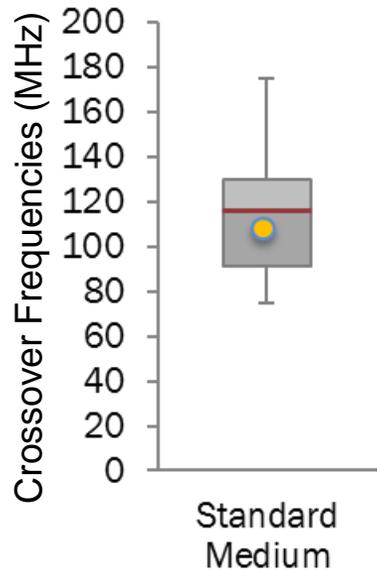
As expected cells are distributed on the edge of the channel attracted and trapped by high intensity E field areas



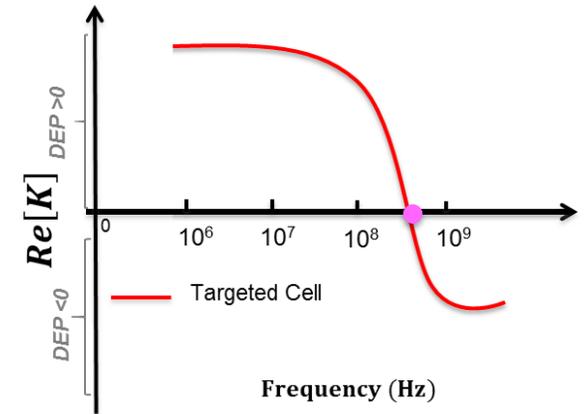
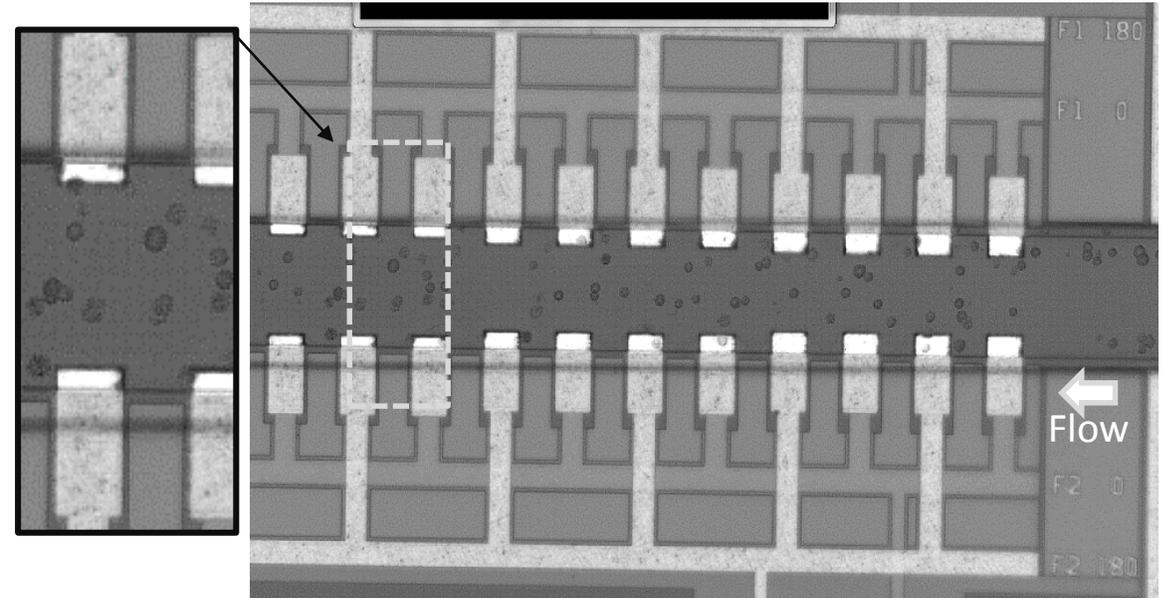
Sorting test on LN18 cells

For $F_{DEP} \sim F_{cross-over}$ (median value for LN18 population)

Cell spatial distribution is much disperse (some repealed in the center, others attracted to the channel edge)



Effect of the cell heterogeneity in the cell line



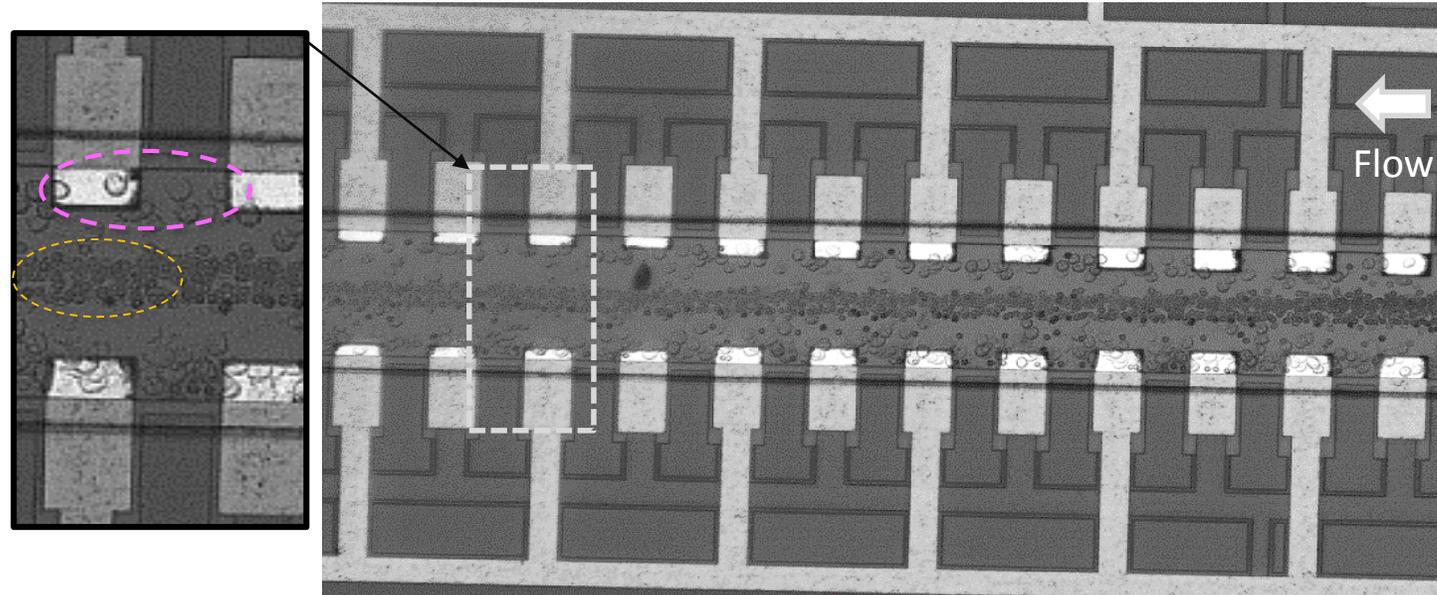
Operating too close $F_{cross-over}$ might not allow efficient cell sorting

Mixing Particle solution sorting capability

Mixing of LN 18 cells and polystyrene beads

- 50MHz DEP signal:
- all polystyrene beads react in $DEP < 0$
 - most of LN18 cells react in $DEP > 0$
 - very few cells seem not be deviated (dead or damaged cells?)

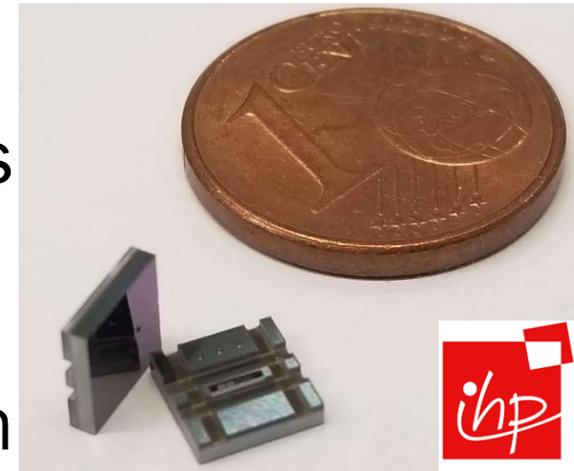
LN18 Cells
7 μ m polystyrene beads



➔ At the sensor output particle distribution testifies of its good sorting ability

Conclusions & prospect

- High frequency DEP could be a powerful tool for cell analysis
- A first prototype has been designed and fabricated based on the coupling of hydrofluidic and selective DEP forces
- HF DEP cytometer working principle validated
- Successful sorting of particles mixtures
 - > cell mixtures in coming weeks
- Technological development still on going improving
...integration



Acknowledgement

Thanks you for your attention



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Project partners:



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UNIVERSITÀ
DEGLI STUDI
DI PADOVA



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