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## ORIGINAL INVESTIGATION

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# Rate of binding of various inhibitors at the dopamine transporter in vivo

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Abstract The rate of entry of drugs into brain is thought to be a factor in their abuse liability. In this investigation, we have examined the rate of entry and binding at dopamine transporters in mouse striatum for a variety of dopamine transporter inhibitors. The method utilized was based on measuring the displacement of <sup>3</sup>H-WIN 35,428 from striatal dopamine transporter sites in vivo at different times. Eleven cocaine analogs (RTI-31, RTI-32, RTI-51, RTI-55, RTI-113, RTI-114, RTI-117, RTI-120, RTI-121, WIN 35,065-2, and WIN 35,428) as well as other dopamine uptake site blockers (bupropion, nomifensine, and methylphenidate) were compared with (-) cocaine for their rates of displacement of <sup>3</sup>H-WIN 35,428 binding in vivo. The drugs that displayed the fastest occupancy rates were bupropion, (-) cocaine, nomifensine, and methylphenidate. RTI-51, RTI-121, RTI-114, RTI-117, RTI-120, RTI-32, RTI-55, and RTI-113, showed intermediate rates, whereas RTI-31, WIN 35,065-2, and WIN 35,428 exhibited the slowest rates of displacement. While many of the cocaine analogs have proven to be behaviorally and pharmacologically more potent than (-) cocaine, their rates of entry and binding site occupancy were slower than that for (-) cocaine.

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Earliest times of transporter occupancy by the different drugs were correlated (although weakly) with their degree of lipophilicity (r = 0.59; P < 0.02). Kinetic effects and metabolism of the compounds could complicate the interpretations of these data. There was no obvious correlation between rate of occupancy in this animal model and abuse liability in humans, which is consistent with the notion that other factors are critical as well.

Key words Dopamine transporter · Cocaine analogs · Mouse striatum

Abbreviations RTI-31  $3\beta$ -[4-Chlorophenyl]tropane- $2\beta$ carboxylic Acid Methyl Ester Hydrochloride RTI-32  $3\beta$ -[4-Methylphenyl]tropane- $2\beta$ -carboxylic Acid Methyl Ester Tartrate · RTI-51 3B-[4-Bromophenyl]tropane- $2\beta$ -carboxylic Acid Methyl Ester Tartrate RTI-55  $3\beta$ -[4-Iodophenyl]tropane- $2\beta$ -carboxylic Acid Methyl Ester Tartrate · RTI-113 3β-[4-Chlorophenyl]tropane- $2\beta$ -carboxylic Acid Phenyl Ester Hydrochloride · RTI-114  $3\beta$ -[4-Chlorophenyl]tropane-2β-carboxylic Acid Isopropyl Ester Hydrochloride ·  $3\beta$ -[4-Methylphenyl]tropane- $2\beta$ -carboxylic RTI-117 Acid Isopropyl Ester Hydrochloride · RTI-120 3β-[4-Methylphenyl]tropane- $2\beta$ -carboxylic Acid Phenyl Ester Hydrochloride  $\cdot$  *RTI-121* 3 $\beta$ -[4-Iodophenyl]tropane- $2\beta$ -carboxylic Acid Isopropyl Ester Hydrochloride  $\cdot$ *WIN-35,065-2*  $3\beta$ -[Phenyl]tropane- $2\beta$ -carboxylic Acid Methyl Ester Tartrate · WIN-35,428  $3\beta$ -[4-Fluorophenyl]-tropane- $2\beta$ -carboxylic Acid Methyl Ester Tartrate.

## Introduction

A neurochemical mechanism hypothesized to underlie at least some of the reinforcing effects of cocaine is an augmentation of dopaminergic transmission due to inhibition of dopamine reuptake (Wise and Bozarth

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1987; Koob and Bloom 1988; Kuhar et al. 1990, 1991; Robinson and Berridge 1993). It has further been suggested that abuse liability is greater in general for those drugs that enter the brain and occupy their receptors more rapidly (see Sellers et al. 1989 for discussion). It is also known that some dopamine transporter inhibitors enter the brain and occupy transporter binding sites at different rates (Pögün et al. 1991).

Previously, the dopamine transporter has been characterized in vitro and in vivo using a variety of radioligands (Carroll et al. 1992a; Wong et al. 1993; Boja et al. 1994). Radiolabeled WIN 35,065-2 (Ritz et al. 1990) and WIN 35,428 (Madras et al. 1989) have been shown to bind in vivo in animals and humans to the dopamine transporter, and the specific binding was also shown to be higher and longer-lasting than that of the parent compound, radiolabeled cocaine (Fowler et al. 1989; Scheffel et al. 1989; Wong et al. 1993). The increased binding was presumed to be the result of higher binding affinity and less vulnerability to metabolism because of the absence of an ester linkage between the phenyl and tropane moieties (Scheffel et al. 1989). In vivo competition studies with in vivo bound <sup>3</sup>H-WIN 35,428 have shown that cocaine enters the brain more rapidly than GBR 12909, and both of these compounds enter more rapidly than mazindol (Pögün et al. 1991). Of these compounds, cocaine is most reinforcing and mazindol is least reinforcing (Chait et al. 1987). Since the rate of transporter occupancy has potential importance, we have further explored this aspect of dopamine transporter binding for 11 cocaine analogs (i.e. RTI-31, RTI-32, RTI-51, RTI-55, RTI-113, RTI-114, RTI-117, RTI-120, RTI-121, WIN-35,065-2, and WIN 35,428) and three other uptake blockers (i.e. bupropion, nomifensine, and methylphenidate) that may or may not have abuse liability (Ritz et al. 1987; Bergman et al. 1989; Garris and Ben-Jonathan 1991; Schaefer and Michael 1992).

#### Materials and methods

#### Drugs and radioactive materials

<sup>3</sup>H-WIN 35,428 was purchased from DuPont NEN (Boston, Mass). The cocaine analogs, RTI-31, RTI-32, RTI-51, RTI-55, RTI-113, RTI-114, RTI-117, RTI-120, RTI-121, WIN-35,065-2, and WIN 35,428 were obtained as previously reported (Carroll et al. 1992b, 1994). Bupropion hydrochloride and methylphenidate were obtained from RBI (Natick, Mass.), nomifensine maleate from Hoechst (Germany) and (–) cocaine from Sigma (St Louis, Mo.). Prior to each experiment, all drugs were freshly dissolved in 0.9% NaCl. For complete solubilization, the solutions were sonicated. Solubilization of (–) cocaine required acidification with 25 µl glacial acetic acid for each ml of saline.

Determination of in vivo competition ED<sub>50</sub> values

Male, CD-1 mice obtained from Charles River Laboratories, (Wilmington, Mass.), weighing approximately 28 g, were injected intravenously via a tail vein with 0.1 ml of the respective drug solution, ranging in concentration from 0.01 to 20 mg/kg. Controls received saline injections. Five minutes after intravenous injection of the drug, 2 µCi <sup>3</sup>H-WIN 35,428 (approximately 10.0 ng in a volume of 0.2 ml saline) was injected. Thirty minutes after tracer injection, the animals were killed by cervical dislocation. Thereafter, the brains were quickly removed and dissected on ice. The cerebellum, olfactory tubercles, corpus striata, and cerebral cortices (all cortical areas except the portion superior to the corpus striatum) were dissected, weighed, and placed into glass vials. After digestion of the tissues with Solvable (Dupont NEN Research Products), 10 ml of scintillation fluid 989 (NEN) was added to each vial, followed by measurement of the radioactivity in a Packard beta scintillation counter, keeping the counting error to 3% or less. Aliquots of the injectate were counted along with the samples. All data were expressed as percent of the injected dose per gram of tissue (% D/g). To calculate the ED<sub>50</sub> of bupropion, methylphenidate, nomifensine, RTI-121, RTI-117, RTI-114, RTI-120, and RTI-113, data from the descending portion of the striatal inhibition curves were analyzed, using logit transformation methodology.

Rate of displacement of <sup>3</sup>H-WIN 35,428 in vivo by various compounds

In order to obtain an estimate of the overall rate of entry of drugs into the brain and of occupancy of the DA transporter, the displacement of maximal <sup>3</sup>H-WIN 35,428 binding from striatal and olfactory binding sites was examined at different times after injection. The experimental design was essentially the same as that used previously to compare the dopamine transporter occupancy rates of mazindol and GBR 12,909 with that of (-) cocaine (Pögün et al. 1991).

Male, CD-1 mice obtained from Charles River Laboratories, (Wilmington, Mass.), weighing approximately 28 g were injected via the tail vein with 2  $\mu$ Ci <sup>3</sup>H-WIN 35,428 (approximately 10.0 ng) in a volume of 0.2 ml saline. Thirty minutes after tracer injection, a time at which in vivo binding was maximal and at apparent equilibrium, the various compounds were injected IV in a volume of 0.1 ml via a tail vein over 10 s. The drugs were administered in a dose equivalent to their ED<sub>50</sub> value determined in this or in previous studies. Controls received saline injections. At different times (30 s, 1, 2, 3, 5, 7, 10, 15, and 20 min) after injection of the drugs (or saline), the animals were killed by cervical dislocation, and dissections and measurements of radioactivity in the various tissue samples were performed.

For the analysis of the in vivo binding data, regional radioactivity levels were converted to percent dose per gram of tissue (%D/g). In addition, the %D/g data of each tissue region studied was divided by the %D/g of the cerebellum in order to obtain ratios of total to nonspecific binding. Data were analyzed by comparing [(tissue to cerebellum) -1] values for the various compounds. As described previously (Scheffel et al. 1989; Cline et al. 1992), these [(T/CB)-1] values provide an estimate of specific to nonspecific binding, based on the observation that dopamine uptake sites are highly concentrated in the striatum and relatively absent in the cerebellum. The time of initial occupancy of striatal DA transporters by each drug was determined as the time point at which a significant difference (P < 0.05) in in vivo binding from controls (saline injected) first occurred. Data were analyzed statistically by using the one-way classification of analysis of variance (ANOVA), with the post-hoc Dunnett's test.

Partition coefficients

Partition coefficients were determined by the chromatographic method of Minick and co-workers (1988), using an Econosil C-8 10  $\mu$ m (4.6  $\times$  10 cm) column and mobile phases which consisted of methanol containing 0.25 v/v octanol and water containing

0.02 M 4-morpholinepropanesulfonic acid (MOPS) and 0.15% n-decylamine. Utilizing different methanol-water mixtures (75:25, 65:35, 55:45, 45:55, 35:65, and 25:75 in the instances of RTI-32, WIN 35428; WIN 35065-2) at 2 ml/min, retention times were measured in triplicate for each compound as well as the reference compounds chloronitrobenzene, naphthalene and pyrene. The column void volume,  $t_0$ , was 1.568 min as determined by the retention time of urea detected at 220 nm. The average of the retention time was then converted to k' at each solvent composition and plotted versus the percentage MeOH contained in that solvent mixture. The y-intercept derived from the linear portion of the curve was calculated, and represents the log  $k'_{w}$  value. The average  $k'_{w}$  values for each compound obtained by this HPLC method were converted to the logarithm of the partition coefficients  $(log P_{hpic})$  through substitution into the following formula:  $\log P_{hplc} = 1.006k'_w + 0.006$ . This formula had been previously generated through the comparison of experimentally determined  $\vec{k}_{w}$  values with known logP<sub>octanol</sub> values for a series of standard compounds (Musachio 1992). The value for pyrene was 4.44, which compares well with reported values of 4.89 (Braumann 1986). HPLC was performed on a Waters Associates Model 510EF pump equipped with a Model 490 UV absorbance detector.

## Results

H<sub>3</sub>C

Various dopamine uptake inhibitors were selected for study. Of the available phenyltropane cocaine analogs, some methyl, isopropyl and phenyl esters (Fig. 1) as well as other uptake inhibitors including methylphenidate, bupropion and nomifensine were selected. Almost all of these compounds were more potent than cocaine in in vitro binding assays (Table 1). The potencies of the compounds in vivo in inhibiting <sup>3</sup>H WIN 35,428 binding were collected from previous publications or determined as part of this study (Table 1).



Fig. 1 Cocaine and cocaine analogs used to displace <sup>3</sup>H-WIN 35,428 binding to the dopamine transporter

**Table 1** Chemical and biological characteristics of the dopamine transporter inhibiting drugs. See materials methods for additional details. Data are mean of 3–11 determinations

Drug	In vivo ED <sub>50</sub> (µmol/kg)	In vitro IC <sub>50</sub> (nM)	Log P <sub>hplc</sub>	
WIN-35,428	0.24 <sup>b</sup>	15.7°	0.79	
WIN-35,065-2	0.61 <sup>b</sup>	23.0 <sup>d</sup>	0.81	
RTI-32	0.31°	1.7°	1.00	
RTI-31	0.18°	1.2°	1.08	
(-) Cocaine	23.1 <sup>b</sup>	89.0°	1.29	
RTI-117	2.61 <sup>a</sup>	6.45 <sup>a</sup>	1.26	
RTI-51	0.31°	1.8°	1.32	
Methylphenidate	11.12 <sup>a</sup>	59.8ª	1.36	
RTI-55	0.26 <sup>c</sup>	1.3°	1.57	
RTI-114	1.52 <sup>a</sup>	1.41 <sup>d</sup>	1.63	
RTI-120	1.91ª	3.27 <sup>d</sup>	2.03	
RTI-121	0.66 <sup>a</sup>	$0.4^{d}$	2.12	
RTI-113	1.23 <sup>a</sup>	1.98 <sup>d</sup>	2.44	
Bupropion	57.20 <sup>a</sup>	199 <sup>a</sup>	2.49	
Nomifensine	9.23ª	18.5 <sup>a</sup>	2.77	

<sup>a</sup> Determined during this study

<sup>b</sup> Scheffel et al. (1991)

<sup>c</sup> Boja et al. (1992)

<sup>d</sup> Carroll et al. (1992)

There was a significant correlation (r = 0.77, P < 0.0008) between log of in vivo binding potencies and log of in vitro binding potencies.

When determining the rate of entry, the transport inhibitors were injected into mice via the tail vein 30 min after injection of <sup>3</sup>H WIN-35,428. At that time, the striatal binding of <sup>3</sup>H WIN-35,428 in vivo is in apparent equilibrium and does not change significantly for the next 30 min or so (Fig. 2; Scheffel et al. 1991). All drugs were injected at their ED<sub>50</sub> doses (Table 1) in order to inject an equieffective dose so that rate of entry rather than potency would be the major factor in our measurements. The animals were killed at varying times after drug injection in order to assess the time at which significant displacement of previously bound <sup>3</sup>H WIN-35,428 occurred.

There was a variation in the times at which the drugs showed a significant displacement of <sup>3</sup>H WIN-35,428 (Figs 2 and 3, Table 2). Some drugs such as bupropion, cocaine, methylphenidate and nomifensine showed competition with <sup>3</sup>H WIN-35,428 at early times, that is, at 1 min and 3 min. On the other hand, drugs such as WIN 35,428 and WIN 35,065–2 did not show competition until much later, that is, at the measured time point of 15 min.

When some of the compounds studied in the in vivo binding competition experiments were injected into animals and assayed for the rate of onset of locomotor activity, it was clear that different analogs had different rates of onset that were generally in agreement with our findings here. However, because of variability, it was difficult to obtain statistically significant differences at early times. Also, the maximal effects for some drugs differed from those of other drugs, making comparisons of rates of onset more difficult (not



Fig. 2 Time course of <sup>3</sup>H-WIN 35,428 displacement by cocaine. Region/cerebellar ratios of <sup>3</sup>H-WIN 35,428 concentrations were determined at different times after injection of cocaine (23.07  $\mu$ mol/kg in 0.1 ml saline) or saline (0.1 ml). <sup>3</sup>H WIN 35,428 (2  $\mu$ Ci/mouse) was injected IV 30 min before IV administration of cocaine or saline. Region/cerebellar ratios defined as % D/g tissue divided by %D/g cerebellum. Data represent means ± SEM (n = 3 - 10). Abbreviations for regions: C. STR. corpus striatum, OLF. T. olfactory tubercles, CER. CTX. cerebral cortex

**Table 2** Comparison of ratios of specific to nonspecific binding [(Str/Cb)-1] of <sup>3</sup>H–WIN 35,428 at different times after intravenous administration of dopamine transporter blockers. Ratios presented

shown). Other investigators have reported different rates of onset of behavior for different compounds (see Discussion).

Lipid solubility is an important factor for drug entry into the CNS. Accordingly, partition coefficients ( $P_{hple}$ ) were determined using an HPLC method. The log  $P_{hple}$  varied from 0.79 to 2.77 (Table 1). When the initial occupancy time shown by these compounds as indicated in Table 2 was correlated to the log of the partition coefficient, a significant although weak relationship (P = 0.02, r = 0.59) was found (Fig. 4).

## Discussion

A rapid rate of delivery of drug to receptors in brain is thought to be a significant factor in abuse liability. While this makes intuitive sense in that a rapid onset of action would be most desirable by a drug abuser. this notion has been supported experimentally as well. For example, rapid delivery of drug to brain and receptors provides optimal conditions for reinforcement, drug self-administration and human subjective response (Arendt et al. 1983; Griffiths et al. 1984a,b; Busto and Sellers 1986; Ator and Griffiths 1987; Griffiths and Wolf 1990; de Wit et al. 1992, 1993; Mumford et al. 1993). Also, prodrugs, that is, compounds which must be converted to an active form and therefore act relatively more slowly, appear to have lesser abuse liability than the ultimate products (Jaffe et al. 1983; Griffiths et al. 1984a, b). It is further postulated that routes of administration that allow for a more rapid entry of drug are more reinforcing; this is thought to be a major factor in the increased abuse liability of "crack" cocaine which is smoked as compared to insufflated cocaine (Johanson and Fischman 1989; Johanson and Schuster 1995). As rate of entry is a

are mean  $\pm$ SEM (n = 3-11). Reading from left to right, the first time point to show significance (\*) is the time at which significant occupancy occurs

Druge	0.5 min	1 min	) min	2 min		7	10 min	15	20
Diugs	0.5 mm	1 11111	2 11111	5 11111	Jimin	/ 11110	10 mm	13 min	20 min
Bupropion	$2.2 \pm 0.1$	$1.9\pm0.1^{*}$	$1.9 \pm 0.1^{*}$	$1.9 \pm 0.1^{*}$	$1.6 \pm 0.1^{*}$		$1.1 \pm 0.1^{*}$	$0.99 \pm 0.1^{*}$	· · · ·
Nomifensine	$2.6 \pm 0.1$	$2.2 \pm 0.1$	$2.2 \pm 0.2$	$2.2 \pm 0.1^{*}$	$1.4 \pm 0.1^{*}$		$1.3 \pm 0.03^{*}$	$1.1 \pm 0.1^{*}$	
(-) Cocaine	$2.7 \pm 0.1$	$2.5 \pm 0.1$	$2.3 \pm 0.3$	$1.8\pm0.1^{*}$	$1.6 \pm 0.1^{*}$		$1.2\pm0.04^{*}$	$0.96 \pm 0.1^{*}$	
Methylphenidate	$2.2 \pm 0.2$	$2.3 \pm 0.2$	$2.3 \pm 0.1$	$1.8\pm0.1^*$	$1.7 \pm 0.1^{*}$		$1.2\pm0.1^{*}$	$0.93 \pm 0.1^{*}$	
RTI51	$2.4 \pm 0.1$	$2.3 \pm 0.2$	$2.0 \pm 0.1$	$2.2 \pm 0.2$	$1.8\pm0.1^{*}$		$1.4\pm0.2^*$	$1.4 \pm 0.1^{*}$	
RTI-121	$2.4 \pm 0.1$	$2.4 \pm 0.1$	$2.4 \pm 0.1$	$2.4 \pm 0.1$	$2.0\pm0.1^{*}$		$2.1 \pm 0.1^{*}$	$1.7 \pm 0.1^{*}$	$1.6 \pm 0.1^{*}$
RTI-114	$2.5 \pm 0.1$	$2.5 \pm 0.04$		$2.2 \pm 0.2$	$2.3 \pm 0.1$	$1.9 \pm 0.1^{*}$	$1.8\pm0.1^{*}$	$1.3 \pm 0.1^{*}$	
RTI-117	$2.5 \pm 0.1$	$2.4 \pm 0.04$	$2.1 \pm 0.1$	$2.0 \pm 0.1$	$2.1 \pm 0.1$	$1.8 \pm 0.02^{*}$		$0.92 \pm 0.1^{*}$	
RTI-120	$2.5 \pm 0.1$	$2.4\pm0.1$	$2.4 \pm 0.1$	$2.3 \pm 0.3$	$2.4 \pm 0.1$	$1.9 \pm 0.2^{*}$	$2.0\pm0.1^*$	$1.6 \pm 0.1^{*}$	
RTI-32	$2.3 \pm 0.1$	$2.5 \pm 0.1$	$2.4 \pm 0.1$	$2.6\pm0.04$	$2.4 \pm 0.04$		$1.3\pm0.2^{*}$	$0.86 \pm 0.1^{*}$	
RTI-55	$2.5 \pm 0.1$		$2.6 \pm 0.1$	$2.7 \pm 0.1$	$2.2 \pm 0.1$		$1.9\pm0.2^{*}$	$2.0 \pm 0.2^*$	
RTI-113	$2.5 \pm 0.1$	$2.6 \pm 0.2$	$2.2 \pm 0.3$	$2.4 \pm 0.2$	$2.4 \pm 0.1$	$2.7 \pm 0.1$	$1.7\pm0.2^{*}$	$1.4 \pm 0.1^*$	
RTI-31	$2.3 \pm 0.1$	$2.4 \pm 0.1$	$2.6 \pm 0.1$	$2.3 \pm 0.2$	$2.4 \pm 0.1$		$2.3 \pm 0.03$	$1.6 \pm 0.1^{*}$	
WIN-3065-2	$2.4 \pm 0.1$	$2.5 \pm 0.1$	$2.6 \pm 0.1$	$2.5 \pm 0.1$	$2.1 \pm 0.01$		$2.1 \pm 0.1$	$1.4 \pm 0.02^*$	$1.7 \pm 0.2^{*}$
WIN-35,428	$2.6 \pm 0.1$	$2.6\pm0.1$	$2.6 \pm 0.1$	$2.6 \pm 0.1$	$2.4\pm0.2$		$2.6 \pm 0.1$	$2.1 \hspace{0.2cm} \pm \hspace{0.2cm} 0.2^{*}$	$1.8\pm0.1^*$

\*Values indicate at least a P < 0.05 value of significance compared to saline injected controls ( $n = 201, 2.6 \pm 0.34$ )

Fig. 3 Rate of inhibition of in vivo transporter binding by various drugs that display different occupancy rates. Bupropion and (-) cocaine are two of the drugs that displayed the fastest occupancy times, i.e. 1 min and 3 min, respectively. RTI-121 and RTI-55 are two of the inhibitors that displayed intermediate occupancy times, i.e. 5 min and 10 min, respectively. RTI-31 and WIN 35,428 are two of the uptake blockers that displayed slowest occupancy times, i.e. 15 min for both. Data are means  $\pm$  SEM (*n* = 3 – 11) of (striatum/cerebellum - 1). The asterisks show the earliest time point at which a significant difference occurred (P < 0.05)



significant factor in the abuse liability of drugs, we have attempted to develop a way of quantifying how quickly drugs enter the brain and occupy their relevant receptors. Recently based on in vivo receptor binding competition, we showed that cocaine enters the brain more rapidly than GBR 12909 which in turn enters more rapidly than mazindol (Pögün et al 1991).

What are potential errors or effects that would affect the interpretation of our data? One factor is that the kinetics of the tracer (<sup>3</sup>H WIN35428) is different from that of most of the other compounds. In other words, WIN35428 has slower entry (and presumably exit) than cocaine and most of the other compounds tested. The net result of this would be that the kinetics of agents with rapid entry would appear to be slower than they actually are. Thus, cocaine's occupancy time could be faster than 3 min, the time at which whole tissue <sup>3</sup>H WIN35428 was reduced below control. This would not be an issue with compounds entering at same rate or more slowly than that of the tracer. Additional factors that could confound our interpretation include competition with endogenous dopamine, and rapid production of metabolites of competing drugs which are also competitors and have different potencies and rates of entry. Corrections for these factors can be made in future studies as we learn to assess their significance.

The notion that drug action can be based on the rate of drug-receptor combination has been proposed by Paton (1961). The theory has been successful in explaining many but not all pharmacological observations (Goldstein et al. 1974 pp. 104–106). A potential rationale why rapid rate is important has to do with desensitization of receptor by agonist; a rapid rate of occupancy would stimulate a maximal number of

Fig. 4 Correlation between initial occupancy time of the dopamine transporter blocking drug and its lipophilicity as expressed by the log of the partition coefficient (log P<sub>bplc</sub>). *M.P.* methylphenidate, *BUP* bupropion, *NOM* nomifensine, *WIN* WIN 35428, *WIN–2* WIN 35065–2



receptors before their response was lessened by desensitization. Several investigators have shown that dopamine receptors are desensitized by exposure to agonists (Memo et al. 1982; Roseboom and Gnegy 1989; Barton and Sibley 1990; Barton et al. 1991). An additional mechanism that could be involved is (slow acting?) feedback regulatory systems. As dopamine in the synapse rises, autoreceptor actions and inhibitory circuits tend to return dopamine to lower levels. Fast entering compounds might cause higher levels of dopamine in the synapse before feedback mechanisms could take effect. Whether or not such mechanisms have validity, or what additional factors might be important, will require more investigation.

As in our previous study (Pögün et al. 1991), cocaine displacement of <sup>3</sup>H WIN 35,428 was significant at 3 min after intravenous injection. In human studies with tracer doses of <sup>11</sup>C-cocaine, specific binding (i.e. striatal radioactivity greater than cerebellar radioactivity) was not detected until about 2 1/2 min after injection (Fowler et al. 1989; J. Fowler personal communication); these findings are in good agreement with those presented here. But cocaine was not the fastest entering drug; bupropion was effective in producing significant displacement at 1 min after injection. Nomifensine and methylphenidate were as fast as cocaine in displacing in vivo <sup>3</sup>H WIN 35428 binding. Of the compounds examined, WIN 35428 and WIN 35065-2 occupied receptors at the slowest rate, i.e., 15 min. The slower entry time of these latter two compounds compared to cocaine has a behavioral counterpart in that onset of behavior was slower than that of cocaine (Spealman et al. 1977, 1989). The remaining compounds entered at intermediate rates, i.e. between 5 and 15 min. According to our previous data, GBR 12,909 and

mazindol appear to occupy transporters at about 5–10 min after IV injection (Pögün et al. 1991).

Are these findings compatible with other data on abuse liability? Not obviously so. Bupropion, nomifensine and methylphenidate enter and occupy drug binding sites as rapidly as or faster than cocaine yet are not abused like cocaine. Bupropion and nomifensine are self-administered by sub-human primates and occasion cocaine-appropriate responding by rats (Spyraki and Fibiger 1981; Bergman et al. 1989; Lamb and Griffiths 1990). However, there is no evidence of abuse of these compounds by humans (Griffith et al. 1983; Miller and Griffith 1983). Methylphenidate also shows potential for abuse in animal studies (Bergman et al. 1989), and in this case, there is some evidence for abuse by humans, although it is not as big a problem as cocaine (Haglund and Howerton 1982; Parran and Jasinski 1991; Chait 1994). Thus, for the drugs producing the fastest displacement, there is little evidence of abuse liability in human populations, although some evidence for abuse liability does exist. This suggests that rate of entry is not the only significant factor in abuse liability which is obviously the case.

The compounds displacing <sup>3</sup>H WIN 35428 most slowly, nonradioactive WIN 35428 and WIN 35065–2, also may have abuse liability in that they are self-administered in animals (Spealman et al. 1977; Spealman and Kelleher 1981). This suggests that all of the other compounds studied would be self-administered in that they inhibit dopamine transport and enter the brain more rapidly than WIN 35,428 or WIN 35,065–2. Thus a much slower entry time (longer than 15 min) may be needed to preclude abuse liability on the basis of rate of entry alone. Also, a confounding factor in existing studies is that animals are trained to self-administer cocaine before being switched to other drugs (Lamb and Griffiths 1990); this may facilitate the self-administration of the other compounds and make a valid comparison of abuse liability between cocaine and the other compounds more difficult. The lack of abuse of some of these compounds in humans may be due to several factors, such as lack of drug availability or the presence of aversive effects of the drugs, such as those reported for mazindol in monkeys (Bergman et al. 1989). Obviously, lack of abuse by humans at present does not mean that abuse will not occur in the future.

After IV injections of cocaine, anecdotal reports by human subjects suggest that a "rush" occurs in typically less than a minute, which is followed by a period of mainly euphoria and arousal. Our findings that injected cocaine takes 3 min to occupy significant levels of transporters suggest that dopamine transporter inhibition mediates the later effects of cocaine rather than the earlier effects. This is supported by direct evidence where the dopamine receptor blocker, haloperidol, reduced drug liking scores elicited by cocaine (Sherer et al. 1989) but had no effect on "rush", Thus, the "rush" may be (partly) peripherally mediated or may be due to action at sites in brain where very low levels of cocaine are effective. However, the finding that cocaine appeared to occupy significant sites at 3 min might be difficult to interpret because of differences in kinetics between tracer and competitor as described above.

It has also been proposed that dopamine transporter inhibiting drugs with a slow rate of entry into brain and a slow onset of action may be useful as treatment medications (Rothman 1990). Balster and Schuster (1973) showed that increasing the duration of a cocaine infusion decreased the response rate of animals. If slow entry drugs are indeed useful, then RTI-31, WIN 35428 and WIN35065–2, which were the slowest entering drugs, may be candidates as treatment medications. Other compounds in the RTI series may be candidates as well. We have not explored the duration of action in these studies. However, it is feasible to do this by similar techniques. For example, we have recently published data regarding the occupancy of serotonin transporters by in vivo competition (Scheffel et al. 1994).

As lipid solubility is one factor which affects entry of a drug into brain, the relative lipophilicity of the drugs under study were determined by a high performance liquid chromatographic method (Minick et al. 1988) (Table 1). As expected, values found for the structurally related cocaine analogs followed two internally consistent trends. First, the log  $P_{hplc}$  increased as the alkoxy portion of the ester moiety changed from methyl to isopropyl to phenyl substitution, reflecting the donation of the additional hydrocarbon fragment. Secondly, the log  $P_{hplc}$  further increased as the substituent on the phenyl ring changed from hydrogen to halogen (Table 1; Fig. 1). Because the other drugs studied come from diverse chemical classes, direct comparisons of the values obtained with these analogs could not be made. The weak correlation (r = 0.59) between rate of entry and log P<sub>hplc</sub> suggests that other or additional pharmacodynamic factors may be involved as well.

In conclusion, our results show that closely related compounds can have quite different rates of entry into brain and occupancy of transporter binding sites. Compounds that enter as fast as or faster than cocaine do not show the same level of abuse as cocaine in humans. This could be due to the fact that these data with mice are not applicable to humans because of pharmacokinetic or other differences. Nevertheless, rates of entry into brain must be at least a partial factor in abuse liability. Rate of entry is also a factor in consideration of developing a substitute or surrogate agonist as a medication for cocaine abuse. The approach used here provides quantitative data and may suggest some candidates for treatment medications.

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### References

- Arendt RM, Greenblatt DJ, de Jong RH, Bonin JD, Abernethy DR et al. (1983) In vitro correlates of benzodiazepine cerebrospinal fluid uptake, pharmacodynamic action and peripheral distribution. J Pharmacol Exp Ther 227:98–106
- Ator NA, Griffiths RR (1987) Self-administration of barbiturat and benzodiazapines: review. Pharmacol Biochem Behav 27:391–398
- Balster RL, Schuster CR (1973) Fixed interval schedule of cocaine reinforcement:effect of dose and infusion duration. J Exp Anal Behav 20:119–129
- Barton AC, Sibley DR (1990) Agonist-induced desensitization of D1-dopamine receptors linked to adenylyl cyclase activity in cultured NS20Y neuroblastoma cells. Mol Pharmacol 38: 531-541
- Barton AC, Black L, Sibley DR (1991) Agonist-induced desensitization of D2 dopamine receptors in human Y-79 retinoblastoma cells. Mol Pharmacol 39:650–658
- Bergman J, Madras BK, Johnson SE, Spealman RD (1989) Effects of cocaine and related drugs in non-human primates. III Selfadministration by squirrel monkeys. J Pharmacol Exp Ther 251:150–155
- Boja JW, Cline EJ, Carroll FL, Lewin AH, Philip A, Dannals R, Wong D, Scheffel, Kuhar MJ (1992) High potency cocaine analogs: neurochemical, imaging, and behavioral studies. The neurobiology of drug and alcohol addiction. Ann NY Acad Sci 654:282–291
- Boja JW, Vaughan R, Patel A, Shaya EK, Kuhar MJ (1994) The dopamine transporter. In:Niznik H (ed) Dopamine receptors and transporters. Marcel Dekker, New York, pp 611–644
- Braumann MS, (1986) Determination of hydrophobic parameters by reversed-phase liquid chromotography: theory, experimental techniques, and application in studies on quantitative structureactivity relationships. J Chromatogr 373:191–225
- Brücke T, Kornhuber J, Angelberger P, Asenbaum S, Frassine H, Podreka I (1993) SPECT imaging of dopamine and serotonin transporters with [<sup>123</sup>I]  $\beta$ -CIT. Binding kinetics in the human brain. J Neural Transm [Gen Sect] 94:137–146
- Busto U, Sellers EM (1986) Pharmacokinetic determinants of drug abuse and dependence. Clin Pharmacokinet 11:144-153

- Carroll FI, Lewin AH, Boja JW, Kuhar MJ (1992a) Cocaine receptor:biochemical characterization and structure-activity relationships of cocaine analogues at the dopamine transporter. J Med Chem 35:969–981
- Carroll FI, Abraham P, Lewin AH, Parham KA, Boja JW, Kuhar MJ (1992b) isopropyl and phenyl esters of  $3\beta$ -(4-Substituted phenyl)tropan- $2\beta$ -carboxylic acids. Potent and selective compounds for the dopamine transporter. J Med Chem 35:2497–2500
- Carroll FI, Mascarella SW, Kuzemko MA, Gao Y, Abraham P, Lewin AH, Boja JW, Kuhar MJ (1994) Synthesis, ligand binding, and QSAR (CoMFA and Classical) study of 3β-(3'-Substituted phenyl)-, 3β-(4'-Substituted phenyl)-, and (3',4'-Disubstituted phenyl)tropane-2β-carboxylic acid methyl esters. J Med Chem (in press)
- Chait LD (1994) Reinforcing and subjective effects of methylphenidate in humans. Behav Pharmacol 5:281-288
- Chair LD, Uhlenhuth EH, Johanson CE (1987) The reinforcing and subjective effects of several anorectics in normal human volunteers. J Pharmacol Exp Ther 242:777–783
- Cline EJ, Scheffel U, Boja JW, Carroll FI, Katz JL, Kuhar MJ (1992) Behavioral effect of novel cocaine analogs: a comparison with in vivo receptor binding potency. J Pharmacol Exp Ther 260[3]:1174–1179
- de Wit H, Bodker B, Ambre J (1992) Rate of increase of plasma drug level influences subjective response in humans. Psychopharmacology 107:352–358
- de Wit H, Dudish S, Ambre J (1993) Subjective and behavioral effects of diazepam depend on its rate of onset. Psychopharmacology 112:324–330
- Farde L, Halldin C, Muller L, Suhara T, Karlsson P, Hall H (1994) PET study of  $[^{11}C]\beta$ -CIT binding to manoamine transporters in the monkey and human brain. Synapse 16:93–103
- Fowler JS, Volkow ND, Wolf AP, Dewey SL, Schyler DJ, MacGregor RR, Hitzemann R, Logan J, Bendriem B, Gatley SJ, Christman D (1989) Mapping cocaine binding sites in human and baboon brain in vivo. Synapse 4:371–377
- Garris PA, Ben-Jonathan N (1991) Effects of reuptake inhibitors on dopamine release from the stalk-median eminence and posterior pituitary in vitro. Brain Res 556:123–129
- Griffiths RR, Wolf B (1990) Relative abuse liability of different benzodiazepines in drug abusers. J Clin Psychopharmacol 10:237-243
- Griffith JD, Carranza J, Griffith C, Miller LL (1983) Bupropion: clinical assay for amphetamine-like abuse potential. J Clin Psychiatry 44:206–208
- Griffiths RR, McLeod DR, Bigelow GE, Liebson IA, Roache JD (1984a) Relative abuse liability of diazepam and oxazepam: behavioral and subjective dose effects. Psychopharmacology 84:147-154
- Griffiths RR, McLeod DR, Bigelow GE, Liebson IA, Toache JD, Nowowieski P (1984b) Comparison of diazepam and oxazepam: preference, liking and extent of abuse. J Pharmacol Exp Ther 229:501–508
- Haglund RMJ, Howerton LL (1982) Ritalin:consequences of abuse in a clinical population. Int J Addict 17[2]:349–356
- Innis R, Baldwin R, Sybirska E, Zea Y, Laruelle M, Al-Tikriti M, Charney D, Zoghbi S, Wisniewski G, Hoffer P, Wang S, Millius R, Neumeyer J (1991) Single photon emission computed tomography imaging of monoamine reuptake sites in primate brain with [<sup>123</sup>I] CIT. Eur J Pharmacol 200:369–370
- Jaffe JH, Ciraulo DA, Nies A, Dixon RB, Monroe LL (1983) Abuse potential of halazepam and of diazepam in patients recently treated for acute alcohol withdrawal. J Clin Pharmacol Ther 34:623–630
- Johanson CE, Fischman MW (1989) The pharmacology of cocaine related to its abuse. Pharmacol Rev 41:3-52
- Johanson CE, Schuster CR (1995) Cocaine. In: Bloom FE, Kupfer DJ (eds) Generation of progress. (in press)
- Kilbourn MR, Mulholland GK, Sherman PS, Pisani T (1991) In vivo binding of the dopamine uptake inhibitor [<sup>18</sup>F]GBR

13119 in MPTP-treated C57BL/6 mice. Nucl Med Biol 18[7]:803-806

- Koob GF, Bloom FE (1988) Cellular and molecular mechanisms of drug dependence. Science 242:715–723
- Kuhar MJ, Sanchez-Roa PM, Wong DF, Dannals RF, Grigoriadis DE, Lew R, Milberger M (1990) Dopamine transporter:biochemistry, pharmacology, and imaging. Eur Neurol Suppl 30[1]:15–20
- Kuhar MJ, Ritz MC, Boja JW (1991) The dopamine hypothesis of the reinforcing properties of cocaine. Trends Neurol Sci 14: 299–302
- Kuikka JT, Bergstrom KA, Ahonan A, Lansimies E (1994) The dosimetry of iodine-123 labelled  $2\beta$ -(4-iodophenyl)tropane. Eur J Nucl Med 21:53–56
- Lamb RJ, Griffiths RR (1990) Self-administration in baboons and the discriminative stimulus effects in rats of bupropion, nomifensine, diclofensine, and imipramine. Psychopharmacology 102[2]:183–190
- Laruelle M, Baldwin R, Malison RT, Zea-Ponce Y, Zoghbi SS, Al-Tikriti MS, Sybirska EH, Zimmerman RC, Wisniewski G, Neumeyer JL, Milius RA, Wang S, Smith EO, Roth RH, Charney DS, Hoffer PB, Innis RB (1993) SPECT imaging of dopamine and serotonin transporters with [ $^{123}I$ ] $\beta$ -CIT pharmacological characterization of brain uptake in non-human primates. Synapse 13:295–309
- Madras BK, Spealman RD, Fahey MA, Neumeyer JL, Saha JK, Milius RA(1989) Cocaine receptors labeled by  $[^{3}H] 2\beta$  carbomethoxy- $3\beta$ -(4-flourophenyl)tropane. Mol Pharmacol 36: 518–524
- Memo M, Lovenberg W, Hanbauer I (1982) Agonist induced subsensitivity of adenylate cyclase coupled with a dopamine receptor in slices from rat corpus striatum. Proc Natl Acad Sci 79:4456-4460
- Miller L, Griffith J (1983) A comparison of bupropion, dextroamphetamine, and placebo in mixed substance abusers. Psychopharmacology 80:199-205
- Minick DJ, Frenz JH, Patrick MA, Brent DA, (1988) A comprehensive method for determining hydrophobicity constants by reversed-phase high-performance liquid chromotography. J Med Chem 31:1923–1933
- Mumford GK, Evans SM, Fleishaker JC, Griffiths RR (1993) Alprazolam absorption kinetics affects abuse liability. Clin Pharmacol Ther 55[2]:PI-39
- Musachio JL (1992) Novel radioiodinated neuroreceptor ligands. PhD Thesis, The Johns Hopkins University, Baltimore
- Parren TV, Jasinski DR (1991) Intravenous methylphenidate abuse. Prototype for prescription drug abuse. Arch Int Med 151:781–783
- Pögün S, Scheffel U, Kuhar MJ (1991) Cocaine displaces [<sup>3</sup>H]WIN35,428 binding to dopamine uptake sites in vivo more rapidly than mazindol or GBR 12909. Eur J Pharmacol 198:203-205
- Ritz MC, Lamb RJ, Goldberg SR, Kuhar MJ (1987) Cocaine receptors on dopamine transporters are related to self-administration of cocaine. Science 237:1219–1223
- Ritz MC, Boja JW, Zaczek R, Carroll FI, Kuhar MJ (1990) <sup>3</sup>H WIN 35,065–2 A ligand for cocaine receptors in striatum. J Neurochem 55:1556–1562
- Robinson T, Berridge KC (1993) The neural basis of drug craving: an incentive-sensitization theory of addiction. Brain Res Rev 18:247–291
- Roseboom PH, Gnegy ME (1988) Acute in vivo amphetamine produces a homologous desensitization of dopamine receptor-coupled adenylate cyclase activities and decreases agonist binding to the D1 site. Mol Pharmacol 34:148–156
- Rothman RB (1990) High affinity dopamine reuptake inhibitors as potential cocaine antagonists: a strategy for drug development. Life Sci 46:PL17-21
- Schaefer GJ, Michael RP (1992) The effects of amphetamine and nomifensine on intracranial self-stimulation discrimination behavior in rats. Pharmacol Biochem Behav 41:391–397

- Scheffel U, Boja JW, Kuhar MJ (1989) Cocaine receptors: in vivo labeling with <sup>3</sup>H- (-) cocaine, <sup>3</sup>H-WIN 35,065–2, and <sup>3</sup>H-WIN 35,428. Synapse 4:390–392
- Scheffel U, Pögün S, Stathis M, Boja JW, Kuhar MJ (1991) In vivo labeling of cocaine binding sites on dopamine transporters with [<sup>3</sup>H]WIN 35,428. J Pharmacol Exp Ther 257[3]:954–958
- Scheffel U, Kim S, Cline EJ, Kuhar MJ (1994) Occupancy of the serotonin transporter by fluoxetine, paroxetine, and sertraline:in vivo studies with [<sup>125</sup>I]RTI-55 Synapse 16:263–268
- Sellers EM, Busto M, Kaplan HL (1989) Pharmacokinetic and pharmacodynamic drug interactions: implications for abuse liability testing. In: Fischman MW, Mello MK (eds) testing for abuse liability in humans. NIDA Research Monograph #92, pp 287-306
- Shaya EK, Scheffel U, Dannals RF, Ricaurte GA, Carroll FI, Wagner HN, Kuhar MJ, Wong DF (1992) In vivo imaging of dopamine reuptake sites in the primate brain using single photon emission computed tomography (SPECT) and iodine-123 lalbeled RTI-55. Synapse 10:1169–1172
- Sherer MA, Kumor KM, Jaffe JH (1989) Effects of intravenous cocaine are partially attenuated by haloperidol. Psychiatry Res 27:117–125

- Spealman RD, Kelleher RT, (1981) Self-administration of cocaine derivatives by squirrel monkeys. J Pharmacol Exp Ther 216:532-536
- Spealman RD, Goldberg SR, Kelleher RT, Goldberg DM, Charlton JP (1977) Some effects of cocaine and two cocaine analogs on schedule-controlled behavior of squirrel monkeys. J Pharmacol Exp Ther 202[3]: 500–509
- Spealman RD, Madras BK, Bergman J (1989) Effects of cocaine and related drugs in non-human primates. II. Stimulant effects on schedule-controlled behavior. J Pharmacol Exp 261:142–149
- Spyraki C, Fibiger HC (1981) Intravenous self-administration of nomifensine in rats: implications for abuse potential in humans. Science 212:1167–1168
- Wise RA, Bozarth MA (1987) A psychomotor stimulant theory of addiction. Psychol Rev 94:469-492
- Wong DW, Yung B, Dannals RF, Shaya ES, Ravert HT, Chen CA, Chan B, Folio T, Scheffel U, Ricaurte G, Neumeyer JL, Wagner HN, Kuhar MJ (1993) In vivo imaging of baboon and human dopamine transporters by positron emission tomography using [<sup>11</sup>C]WIN 35.428. Synapse 15:130–142