

Primers

Reaction	Loci	Label	Direction	Sequence
Mix 1	Tle19	FAM	Forward	CACTGCTGTGATGTCTTTTCAGG
Mix 1	Tle19		Reverse	CTCTTAAGCTTCTGAAGTGTCAAGG
Mix 1	Tabi8	PET	Forward	GCCAACATCCTTCTCTGTAAATCCC
Mix 1	Tabi8		Reverse	AGCAGATGTAAGTGTGCCCTTTTG
Mix 1	Tle16	VIC	Forward	GTCTATTTCCTTCTAGGGATTGAG
Mix 1	Tle16		Reverse	GTATTTTTTGCTGCCATACGCTC
Mix 1	Tabi4	NED	Forward	GAAGTTCCTCATAGCTGGCTCTAAGAC
Mix 1	Tabi4		Reverse	TAGTGGTGAAAGACAGCTTGCTGG
Mix 1	Tabi1	FAM	Forward	GCTGAATTTGGACAACTCACCTC
Mix 1	Tabi1		Reverse	CTCCCAATCAGAAGCAGAAGCAC
Mix 2	Tbi104	FAM	Forward	CCCTGTGGTTCCTTGTTCTG
Mix 2	Tbi104		Reverse	GTTTTCTACGAGCCTCCCTTGGTGA
Mix 2	Tabi34	VIC	Forward	GGAAGCACGAGATGGTATTAC
Mix 2	Tabi34		Reverse	CCCACCAGAATCCTCCACAG
Mix 2	Tal6	VIC	Forward	GGATATATCTCAGTGGCCTAATGGC
Mix 2	Tal6		Reverse	CTCATGCATCATTGGATTAAGTGG
Mix 2	Tabi25	PET	Forward	CACTGCGTACCTAAAATCTCTGG
Mix 2	Tabi25		Reverse	CTGAAGTCTAGCACTGGAAGTCTG

Reactions

Mix 1	
Reagent	Volume (ul)
H2O	4.24
10 x PCR buffer	1.00
MgCl2 (25mM)	1.30
10 uM TLE19 FAM F	0.12
10 uM TLE 19 R	0.12
10 uM Tle 16 VIC F	0.12
10 uM Tle 16 R	0.12
10 uM TaBi 4 NED F	0.36
10 uM TaBi 4 R	0.36
10 uM TaBi 1 FAM F	0.12
10 uM TaBi 1 R	0.12
10 uM TaBi 8 PET F	0.36
10 uM TaBi 8 R	0.36
dNTPs (10mM)	0.20
Taq (2.5 U/ul)	0.10
DNA	1.00
Total	10.00

Mix 2	
Reagent	Volume (ul)
H2O	5.04
10 x PCR buffer	1.00
MgCl2 (25mM)	1.30
10 uM TaBi 25 PET F	0.30
10 uM TaBi 25 R	0.30
10 uM Tal 6 VIC F	0.12
10 uM Tal 6 R	0.12
10 uM TaBi 34 VIC F	0.12
10 uM TaBi 34 R	0.12
10 uM Tbi 104 FAM F	0.14
10 uM Tbi 104 R	0.14
dNTPs (10mM)	0.20
Taq (2.5 U/ul)	0.10
DNA	1.00
Total	10.00

Cycling Conditions

Mix 1
1. 95 C for 2 minutes
2. 95 C for 50 seconds
3. 56 C for 1 minute
4. 72 C for 1 minute
5. Repeat 2-4 34x
6. 72 C for 30 minutes
7. Hold 10 C

Mix 2
1. 95 C for 2 minutes
2. 95 C for 50 seconds
3. 58 C for 1 minute
4. 72 C for 1 minute
5. Repeat 2-4 34x
6. 72 C for 30 minutes
7. Hold 10 C