

1 **Towards a Global Public Repository of Community Protocols to**  
2 **encourage Best Practices in Biomolecular Ocean Observing and**  
3 **Research**

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28 **Abstract**

29 Biomolecular ocean observing and research is a rapidly evolving field that uses omics approaches to describe  
30 biodiversity at its foundational level, giving insight into the structure and function of marine ecosystems over  
31 time and space. To achieve a global ocean biomolecular observing network (OBON) for the UN Decade of  
32 Ocean Science for Sustainable Development and beyond, research groups will need a system to effectively share,  
33 discover, and compare omic practices and protocols. While numerous informatic tools and standards exist, there  
34 is currently no global, publicly-supported platform specifically designed for sharing marine omics (or any omics)  
35 protocols across the entire value-chain from initiating a study to the publication and use of its results. Towards  
36 that goal, we propose the development of the Minimum Information for an Omic Protocol (MIOP), a  
37 community-developed guide of curated, standardized metadata tags and categories that will orient protocols in

38 the value-chain for the facilitated, structured, and user-driven discovery of suitable protocol suites on the Ocean  
 39 Best Practices System. Users can annotate their protocols with these tags, or use them as search criteria to find  
 40 appropriate protocols. Implementing such a curated repository is an essential step towards establishing best  
 41 practices. Sharing protocols and encouraging comparisons through this repository will be the first steps towards  
 42 designing a decision tree to guide users to community endorsed best practices.

## 43 **1 Introduction**

44 The term “omics” generally means studying anything holistically, and here we take a broad view of  
 45 biomolecular omics that includes, but is not limited to: quantitative target gene amplification (e.g.  
 46 qPCR, qNASBA etc.), (meta)barcoding, (meta)genomics, (meta)transcriptomics, (meta)proteomics,  
 47 and metabolomics; and field collection approaches that target organisms or parts thereof, including  
 48 single-celled organisms (microorganisms), as well as environmental DNA (eDNA). In the marine  
 49 realm, omic techniques are used to assess and monitor biodiversity, reveal population structure and  
 50 gene flow, and discover new compounds with applications in medicine and industry. Rapid advances  
 51 in omics research, and the declining cost of high-throughput sequencing technologies (Wetterstrand,  
 52 2020) support the increasing application of omics in marine microbiome research.

53 The recent expansion in marine omics has led to a proliferation of protocols specific to multiple  
 54 applications. However, these protocols are rarely shared publicly with sufficient detail to reliably  
 55 reproduce a study (Dickie et al., 2018). While the omics community has already achieved high  
 56 standards for sharing sequence data through the [International Nucleotide Sequence Database  
 57 Collaboration](#), these data often lack sufficient metadata and provenance information on the protocols  
 58 used (Dickie et al., 2018), undermining efforts to implement the Findable, Accessible, Interoperable  
 59 and Reusable ([FAIR](#)) data principles (Wilkinson et al., 2016). These limitations create challenges for  
 60 marine microbiome research and operations from individual labs up to global (meta)data analysis  
 61 efforts such as [MGnify](#) (Mitchell et al., 2020), which must identify data collected using comparable  
 62 methods, in order to integrate and re-use data for meta-analysis (Berry et al., 2020). Moreover, a lack  
 63 of protocol-sharing impedes the identification of comparable methods needed for global monitoring  
 64 efforts aiming to understand, and sustainably manage the changing marine ecosystem (Berry et al.,  
 65 2020).

66 Many projects are looking to develop best practices for omics research: standards organisations, such  
 67 as the [Genomic Standards Consortium](#)’s (GSC) [Genomic Biodiversity Interest Group](#), the  
 68 [Biodiversity Information Standards](#) (TDWG) and the [Biocode Commons](#) are working collaboratively  
 69 towards standards specifications for genomic observatories (Davies et al., 2012; Davies et al., 2014).  
 70 Large campaigns, such as the [Earth Microbiome Project](#) (Thompson et al., 2017), [TARA Oceans](#)  
 71 (Sunagawa et al., 2020), and the [Australian Microbiome Initiative](#) (AM; Bissett et al., 2016; Brown et  
 72 al., 2018; DOI:[10.4227/71/561c9bc670099](#)), have already developed standardised practices, and  
 73 innovative software enterprises, such as [protocols.io](#), are providing powerful solutions for sharing  
 74 protocols. Yet there is currently no global, publicly-supported infrastructure developed explicitly for  
 75 encouraging the exchange and harmonization of omic protocols, so these valuable contributions  
 76 remain fragmented and underutilized.

77 For marine ecosystems, the Intergovernmental Oceanographic Commission’s [Ocean Best Practices  
 78 System](#) (OBPS) provides a public repository for all ocean research methodological documentation  
 79 that can interlink protocols, standard specifications, and other guidelines. The OBPS seeks to support  
 80 continuous convergence of methods as they undergo community refinement to become best practices  
 81 (Hörstmann et al., 2021). In collaboration with the broader omics community, through the Omic  
 82 BON initiative (Buttigieg et al., 2019), we propose to develop a best practice system specific to

83 marine omics research, leveraging the framework of the OBPS to curate a global repository for  
84 marine omics protocols.

85 As part of the omics/eDNA session at the 4th OBPS [workshop](#), we discussed recommendations and  
86 community needs for an omics/eDNA specific best practices system. Recognizing an urgent need for  
87 the ocean omics community to get organized as the UN Decade of Ocean Science for Sustainable  
88 Development starts, we identified the demand for publishing protocols into a user-friendly decision  
89 tree framework. With such a framework we would aim to support protocol selection, increase  
90 protocol findability and improve recognition for protocol developers. In a series of focused follow-up  
91 meetings, we identified that an omics decision tree would require a library of constituent parts (the  
92 protocols) and framework to: (1) locate where the protocol fits within the entire omics workflow  
93 (outlined in section 2), and (2) organise protocols using focused descriptive terms (metadata tags),  
94 based on what the protocol does and how/why it is used (outlined in section 3).

## 95 **2 Ocean Omics Methodology Categories**

96 The typical omics workflow involves a series of protocols, which take a project from ideation,  
97 through to publication, and on to societal use. Protocols from each step in the omics workflow hold  
98 valuable information for different groups. For example, sample collection protocols may be most  
99 relevant to scientists/technicians in the field, whereas local stakeholders and indigenous communities  
100 may primarily engage with aspects of how the project and resulting data address and impact  
101 important ethical, legal, and societal issues. Documenting details and provenance for the entire  
102 marine omics workflow requires input from multiple parties, as each step of the workflow may be  
103 conducted by different individuals or groups. The omics OBPS therefore needs to identify these key  
104 methodological categories, to allow protocols and accompanying metadata to be uploaded in modules  
105 that link together to form the entire workflow.

106 We propose twelve protocol categories (Figure 1A) for ocean omics research and operations.  
107 Protocols and guidelines are assigned into these categories according to the purpose they serve<sup>1</sup>.  
108 Categories 5-12 outline methodological categories for operational activities used in the AM Initiative  
109 (van de Kamp et al., 2019). Categories 1-4 were identified to additionally cover cross-cutting  
110 documentation in the omics workflow: 1) Society, 2) Sampling/observational design, 3) Ethics and  
111 law, and 4) Data management.

- 112 1) Society - All workflows should begin and end with society; societal needs inform the question  
113 or purpose behind the research, and societal impacts show the value in the research once it has  
114 been completed.
- 115 2) Design and logistics - This category covers the practical logistics for implementing ocean  
116 omics research and operations, including the experimental/observational design formulated to  
117 address the societal priorities outlined in 1.
- 118 3) Ethics and law - A survey of workshop participants highlighted a need for guidance on  
119 sharing data and complying with important ethical and legal requirements (Simpson et al.  
120 2021). This category will include information on permits and permission required to obtain  
121 samples and release data. Collating and publishing this information will firstly provide  
122 examples for how previous projects have adhered to legal requirements/ethical principles and

<sup>1</sup> Currently, the protocol categories focus on genomics and transcriptomics but we expect this list to expand with further input from the broader omics community, particularly in areas such as proteomics and metabolomics.

123 secondly stimulate discussion on how to facilitate adherence to these requirements and  
124 principles, perhaps through checklists, templates, or training materials.  
125 4) Data management - The data management plan (DMP) is designed to support all the  
126 downstream steps according to the ethics, legalities and societal needs identified in 1) - 3),  
127 while making sure that the (meta)data flows to the right stakeholders in society that we need  
128 to interface with. DMPs should be drafted prior to data collection and referred to throughout  
129 the workflow to ensure that quality assurance and quality checks take place, and that detailed  
130 information on (meta)data requirements for both short and long-term (meta)data storage is  
131 given. There is a growing body of tools and best practices surrounding DMPs, including  
132 principles for making them more machine-actionable, that should be leveraged in omic  
133 protocols and associated infrastructure (see Miksa et al., 2019). Publishing documentation on  
134 omics specific DMPs will increase transparency for funders by providing direct links to the  
135 protocols they refer to. Furthermore, collating examples of omics specific DMPs will provide  
136 insight into what the community needs from omics specific data management tools.

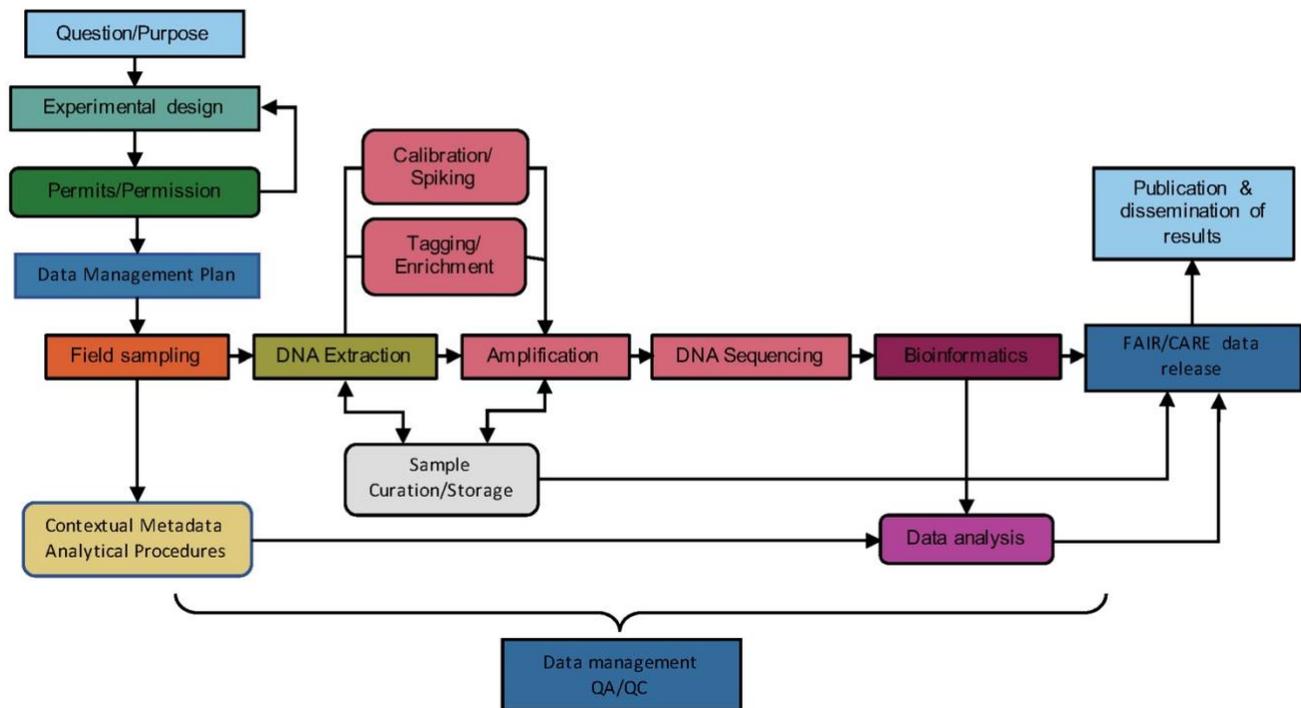
137 In Figure 1B, we give an example of a DNA metabarcoding workflow, where the colour of each step  
138 corresponds to a methodology category in Figure 1B. Protocols uploaded to OBPS can be assigned  
139 (tagged) to the relevant omics categories. The granularity of protocols uploaded to the OBPS may  
140 include individual uploads for sub-stages (i.e. Tagging/Enrichment within 4. Omics sequencing  
141 procedures), or single documents spanning multiple methodology categories (i.e. 3. Sample  
142 extraction & purification, through to 5. Bioinformatics). To accommodate these levels of granularity,  
143 each upload could be tagged with single or multiple methodology category and linked to those  
144 protocols pre- and succeeding it. The granular use of methodology categories will increase  
145 modularity within the omics workflow and facilitate the mixing and matching of methods from  
146 various projects.

147 The interplay between the activities within and across the steps within a workflow - and how they  
148 bring value to the community and society - is complex and beyond the scope of this article; however,  
149 we have provided an initial perspective on this using the Porter's value chain approach (Porter, 1985;  
150 Supplementary Figure 1).

A

1. Society	Documenting societal inputs and outputs (e.g. mission statements (linked to broadly accepted societal priorities e.g. SDGs EOVs, EBVs. etc), stakeholder consultation outputs, associated publications/media, links to relevant data repositories, educational materials associated with the workflow)
2. Design and logistics	Documenting plans for the experimental/observational design (e.g. cruise schedules/reports, deployment regimes, statistical replication, logistics, costing/resources/material lists)
3. Ethics and law	Documenting permits, permissions and guidelines for obtaining samples and release data. (e.g., Internationally Recognized Certificate of Compliance (IRCC) code for sampling, or relevant Due Diligence documentation if an IRCC code is not required, for countries that have signed and ratified the Nagoya protocol (Nagoya Protocol, 2010)) checklists showing compliance to ethical principle (e.g. OCAP) for ethical sampling on indigenous lands, etc
4. Data management	Protocols for data and quality management. This will include data management plans with quality assurance and quality check measures to be undertaken throughout the workflow, as well as checklists and guidance documents for the compliance with FAIR/CARE data standards; and links to all data repositories to which the omics study (meta)data should be uploaded.
5. Sampling Collection Procedure	Protocols for the physical collection of samples to be used in omics research. Including both environmental samples (e.g. water, sediments, biofilms) and organismal samples (e.g. gut contents, bulk plankton, sponge).
6. Contextual Metadata Analytical Procedures	Protocols used for the measurement of additional parameters (e.g. temperature, salinity, pH, chlorophyll etc.)
7. Sample Extraction & Purification	Protocols for the extraction and purification of biological molecules (e.g. DNA, RNA, Proteins) from environmental and/or organismal samples.
8. Omics Sequencing Procedures	Protocols for sequencing nucleic acids. This stage may contain sub-stages that occur during the sample extraction and purification stage, if so, links between the two stages should be provided.
9. Bioinformatics	Bioinformatics pipeline for processing sequence data. This can include links to github, docker etc.
10. Quantitative amplification	Protocols for the quantitative amplification of DNA and RNA. Including quantitative PCR (qPCR), reverse transcriptase quantitative PCR (rt-qPCR), nucleic acid sequence-based amplification (NASBA), Recombinase Polymerase Amplification (RPA) etc.
11. Data Analysis	Protocols for analysis of data. This will include scripts for statistical tests, modelling and data visualisations.
12. Sample archiving/biobanking	Protocols for archiving physical samples at all stages of laboratory processing (e.g. whole environmental/organismal samples, DNA/RNA/protein extracts and/or amplicons)

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**Figure 1. A.** Proposed methodology categories to enhance exchange of ocean omics analysis knowhow. Protocols, guidelines, and other methodologies in some of these categories (such as Sample archiving/biobanking, Data Management, and Society) are cross-cutting and may apply at multiple points in the workflow. **B.** Example workflow for a DNA metabarcoding project. Colours correspond

155 to the methodology categories outlined in A and arrows indicate the order of the workflow. Square boxes show essential steps in a  
 156 metabarcoding workflow, whereas rounded boxes indicate non-essential steps. Data management and QA/QC are required throughout  
 157 the entire workflow.

### 158 3 Essential Metadata for Omics Protocols

159 The targeted discovery and reuse of protocols can be improved if protocols are effectively described  
 160 using standardised metadata terms on upload to OBPS and other platforms. Terms and checklists to  
 161 standardise metadata about primary sequence or biodiversity data already exist (GSC’s Minimum  
 162 Information about any (x) Sequence checklist (MIxS; Yilmaz et al., 2011) and TDWG’s Darwin  
 163 Core standard (Wieczorek et al., 2012)); however, no such standards have thus far been published for  
 164 metadata about omics protocols.

165 Here we present initial suggestions for the Minimum Information for an Omic Protocol (MIOP), a set  
 166 of ten metadata categories which could correspond to ten key decision tree questions asked to  
 167 identify the relevant protocol for any project. The ten MIOP categories (Table 1) consist of five novel  
 168 categories (methodology category, purpose, resources, analysis, target) and five categories already  
 169 used in the GSC’s MIxS (project, geographic location, broad-scale environmental context, local  
 170 environmental context, and environmental medium). Each category is linked to a set of predefined  
 171 keywords (metadata terms) from existing vocabularies or ontologies; except for the ‘project’  
 172 category, which contains project names, affiliations, and contact details and the ‘methodology  
 173 category’ outlined in section 2 (Figure 1A). Omics users would then select the most appropriate  
 174 keywords for each category, assigning the terms as metadata for the protocol. This will improve the  
 175 FAIRness of our protocol data, by allowing consequent users to search the protocol database using  
 176 the same set of keywords; thereby, limiting the proliferation of descriptive keywords (e.g., mapping  
 177 synonyms) and increasing the findability of protocols.

178 **Table 1.** Description of keyword categories for protocol metadata and the terminologies (controlled vocabularies, thesauri, ontologies)  
 179 containing the relevant keywords. Terms would be added at upload and additional metadata would accumulate as the protocols are used  
 180 in different settings (e.g., Geographic Locations; see 4.2 learning from failed practices). EFO: Environmental Factor Ontology, OBI:  
 181 Ontology for Biomedical Investigations, NCIT: NCI Thesaurus, GAZ: Gazetteer, ENVO: Environment Ontology, UBERON: Uber-  
 182 anatomy ontology, NCBITaxon: NCBITaxon ontology.

Categories	Terminology/Ontology	Description
Methodology category	Methodology category (see Figure 1A.)	Methodology category which the uploaded protocol belongs to. This links to the associated methodology categories which precede and succeed it in the workflow, to facilitate the linking of protocols into entire workflows, while keeping granularity and flexibility. This will enable the mixing and matching of protocol modules from various uploaded workflows.
Project	N/A	Details about the project (e.g. Name, Affiliation, website). May also includes a field for tagging any projects that protocols are compliant with (e.g. Earth Microbiome Project/TARA Oceans). Once submitted the relevant PI may be notified and could choose to endorse or reject the protocol as compliant with their project.
Purpose	EFO, OBI	Terms to describe the purpose of the omics research. (e.g. time series design [OBI:0500020] or taxonomic diversity assessment by targeted gene survey [OBI:0001960])
Resources	EFO, NCIT	Terms to identify the key resources needed to complete the protocol (e.g. Illumina MiSeq [EFO:0004205], centrifuge [OBI:0400106])
Analyses	EFO, OBI, NCIT	Terms to describe the types of analyses used in the protocol (e.g. amplicon sequencing assay [OBI:0002767] or polymerase chain reaction [OBI:0002692])
Geographic Location	GAZ	Geographic location/s in which the protocol has been used (e.g. Hawaii Ocean Time-series Site [GAZ:00187530], Western English Channel Sampling Stations [GAZ:00187525])

Broad-scale environmental context (former Biome)	ENVO	Biome/s in which the protocol was successfully used (e.g. oceanic epipelagic zone biome [ENVO:01000033])
Local environmental context (former Feature)	ENVO, UBERON	Environmental feature/s targeted using the protocol (e.g. seasonal thermocline [ENVO:01000107])
Environmental medium (former Material)	MIXS environmental packages; ENVO	Identify the environmental or organismal material from which the biological molecule (e.g. DNA/RNA/Protein) was extracted (e.g. ocean water [ENVO:00002151])
Target	NCIT, NCBITaxon, EFO	Identify the target taxa, gene and/or molecule for the protocol (e.g. Polaribacter [NCBITaxon:1642819], 16S Mitochondrial Ribosomal RNA [NCIT:C131261]).

183

184 **4 Discussion**

185 OBPS provides a neutral, global public repository for ocean community practices. It is a stable and  
 186 persistent foundation that can host protocols themselves, or link to other protocol tools and  
 187 functionalities that can (and should) continue to be developed by other organizations including the  
 188 private sector. The primary function of Omics OBPS would be to publish and archive omics  
 189 protocols to enhance their global visibility and discoverability, and provide stable links to the entire  
 190 workflow of protocols. Expanding and improving the functionality of the OBPS for omics protocols  
 191 will help the community mature by providing a structured system in which context-based best  
 192 practices can be discovered and identified. A transparent and structured process for handling our  
 193 omics protocols will be an essential step towards operationalizing omics observing.

194 Increasing protocol transparency, through detailed publication on OBPS, also means that simple cited  
 195 protocol strings can become a core component of methods sections in publications. Those strings can  
 196 then be harvested by machines to generate a graph of “what came before” and “what came after”.  
 197 When used with the decision tree recommendations this process could point out the most recent  
 198 protocol development to users and would essentially provide the decision-tree resource we are aiming  
 199 for. Such an approach enables ‘practices’ (which might be defined as ‘protocol strings’) to emerge  
 200 from how protocols are actually being used in the community. Assessment of which of these  
 201 practices represent a “best” practice in a given context is a distinct challenge, but not a unique one in  
 202 knowledge sectors. Peer endorsement and citation metrics are two commonly employed ranking  
 203 mechanisms that could also be applied here.

204 **4.1 Learning from community preferences**

205 Community-use metrics offer a way to capture the community’s preference for certain protocols. We  
 206 suggest that metrics such as times cited, user upvotes, and number of associated data records all be  
 207 recorded and used to rank lists of relevant protocols. Combined with the MIOP-based grouping into  
 208 methodology categories, this process will help accelerate the identification of potential best practices  
 209 within each category. Narrowing down the list of relevant protocols will additionally provide the  
 210 basis for more targeted and rigorous scientific comparisons between multiple potential best practices  
 211 for a given scientific endeavour. Outputs of such comparisons may offer further information about  
 212 the superiority of certain protocols, and could be considered in addition to the more general

213 community-use metrics<sup>2</sup>. Furthermore, focusing on these community driven best practices will help  
 214 to reveal protocols that are effective and convenient for a broad range of research facilities. This in  
 215 turn can reduce literature biases toward novel state of the art practices, which may not be feasible for  
 216 mainstream use.

217 **4.2 Learning from failed practices**

218 During the initial workshop, participants outlined a desire for a best practice system to include ‘failed  
 219 practices’ and flag when a protocol may limit or eliminate a range of downstream applications. While  
 220 this type of functionality would not be immediately addressed by implementing MIOP metadata,  
 221 there would be potential for users to provide feedback for protocols using MIOP metadata and  
 222 boolean operators. For example, if a protocol, originally designed for seawater, was used with  
 223 freshwater samples, the user could upload additional MIOP metadata using ‘AND freshwater’ if the  
 224 protocol was successful or ‘NOT freshwater’ if unsuccessful. Thereby, broadening the findability of  
 225 successful protocols and documenting potential limitations to be aware of. Documenting these failed  
 226 attempts has the potential to save both time and resources.

227 **4.3 Promoting collaborative omic networks**

228 MIOP may additionally promote collaboration between groups. For example, the ‘Project’ category  
 229 is an administrative metadata field that will describe the project (study or program) for which the  
 230 protocol was developed, including contact details and affiliated institution. To create links between  
 231 similar projects and facilitate collaboration, it would be possible to introduce an option to tag a  
 232 protocol as compliant with pre-existing projects. In such cases, a notification could be sent to the PI  
 233 of the lead project, allowing them to add or reject the protocol to their list of compliant protocols.  
 234 Protocols linked this way could form overarching protocol concepts, which may contain a variety of  
 235 versions and accepted, cross-comparable protocols that include minor adaptations to make them  
 236 suitable in different circumstances.

237 An endorsement process for a global observation network has already been developed by Global  
 238 Ocean Observing System (GOOS) in cooperation with OBPS, to encourage standardised methods for  
 239 global observations and for reporting on GOOS’ Essential Ocean Variables (EOVs) (Miloslavich et  
 240 al., 2019; Hermes, 2020). To gain this endorsement, protocols will have to undergo a rigorous  
 241 community review process that will be strengthened if there is a large source of omics protocols to  
 242 compare with on the OBPS. Standardised practices and official endorsements are likely to become  
 243 increasingly valuable as countries begin to use legislation to make biodiversity targets legally  
 244 binding. Any omic method used to measure biodiversity impacts will need to undergo legal scrutiny  
 245 if it is used as evidence of a country/organisation meeting or failing to meet biodiversity targets.  
 246 Therefore, protocols officially endorsed through international programmes, such as GOOS, are likely  
 247 to hold more sway legally. Broad participation from the omics community in open sharing and  
 248 reviewing of protocols on the OBPS will help to ensure that community endorsed best practices are  
 249 representative of the wider community needs and not only focussed on expensive state of the art  
 250 methodologies.

251 **4.4 Machine readability**

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<sup>2</sup> In certain cases (e.g., for contributing to a standardised global sampling scheme) it may not be about which method is “best”, but about which method delivers reliable results while being applicable throughout all regions of the ocean and inclusive of lower capacity research activities.

252 Machine readable tracking of protocol versions presents an opportunity to visually map the  
 253 progression of protocols by linking all versions to a “concept”, as implemented in [Zenodo](#) and  
 254 GitHub. Like software, omic protocols may be updated, corrected, and improved necessitating forms  
 255 of version control and tracking, such as the use of semantic versioning (Preston-Werner, 2021;  
 256 Hörstmann et al. 2020). Implementing this would help to increase recognition for the  
 257 scientists/technicians/students involved in protocol development through citable documentation of  
 258 their contributions.

259 Machine-readable and machine-actionable protocols are becoming more important as autonomous  
 260 technologies evolve. Devices such as the Environmental Sample Processor (ESP) and the Robotic  
 261 Cartridge Sampling Instrument (RoCSI) are currently being used and developed for autonomous  
 262 collection, preservation, and *in-situ* analysis of omics samples (Yamahara et al., 2019; National  
 263 Oceanography Centre, 2021). Eventually, smart sensing platforms using these technologies will be  
 264 able integrate data from various sensors and satellites to implement adaptive sampling regimes or  
 265 extraction protocols based on real-time environmental observations (Witt et al. 2020). To reach this  
 266 goal a variety of protocols will need to be translated into a machine actionable format using common  
 267 workflow language. A systematic review of protocols will help to devise such machine actionable  
 268 formats and protocol templates may help to bridge the gap between lab-based protocol development  
 269 and *in-situ* autonomous use.

## 270 **5 Conclusion**

271 Multiple groups within the omics community are actively developing best practices for their field. To  
 272 ensure that all these efforts are effectively utilized, a concerted and community wide effort will be  
 273 needed to gather and organise these practices. By harnessing the OBPS infrastructure and further  
 274 developing the MIOP metadata we can: 1) allow protocols to be searched for within a decision tree  
 275 framework; 2) establish a system that encourages the systematic review of protocols; and 3) reveal  
 276 community preferences through the accumulation of community use data. Taking these steps toward  
 277 a structured and global public repository of omics protocols will increase transparency and streamline  
 278 biomolecular ocean observing research to foster the collaborative networks needed to achieve global  
 279 scale biodiversity observations.

## 280 **6 Conflict of Interest**

281 The authors declare that the research was conducted in the absence of any commercial or financial  
 282 relationships that could be construed as a potential conflict of interest.

## 283 **7 Author Contributions**

284 All authors contributed to the discussion and wrote the manuscript. AW, CM and RS constructed the  
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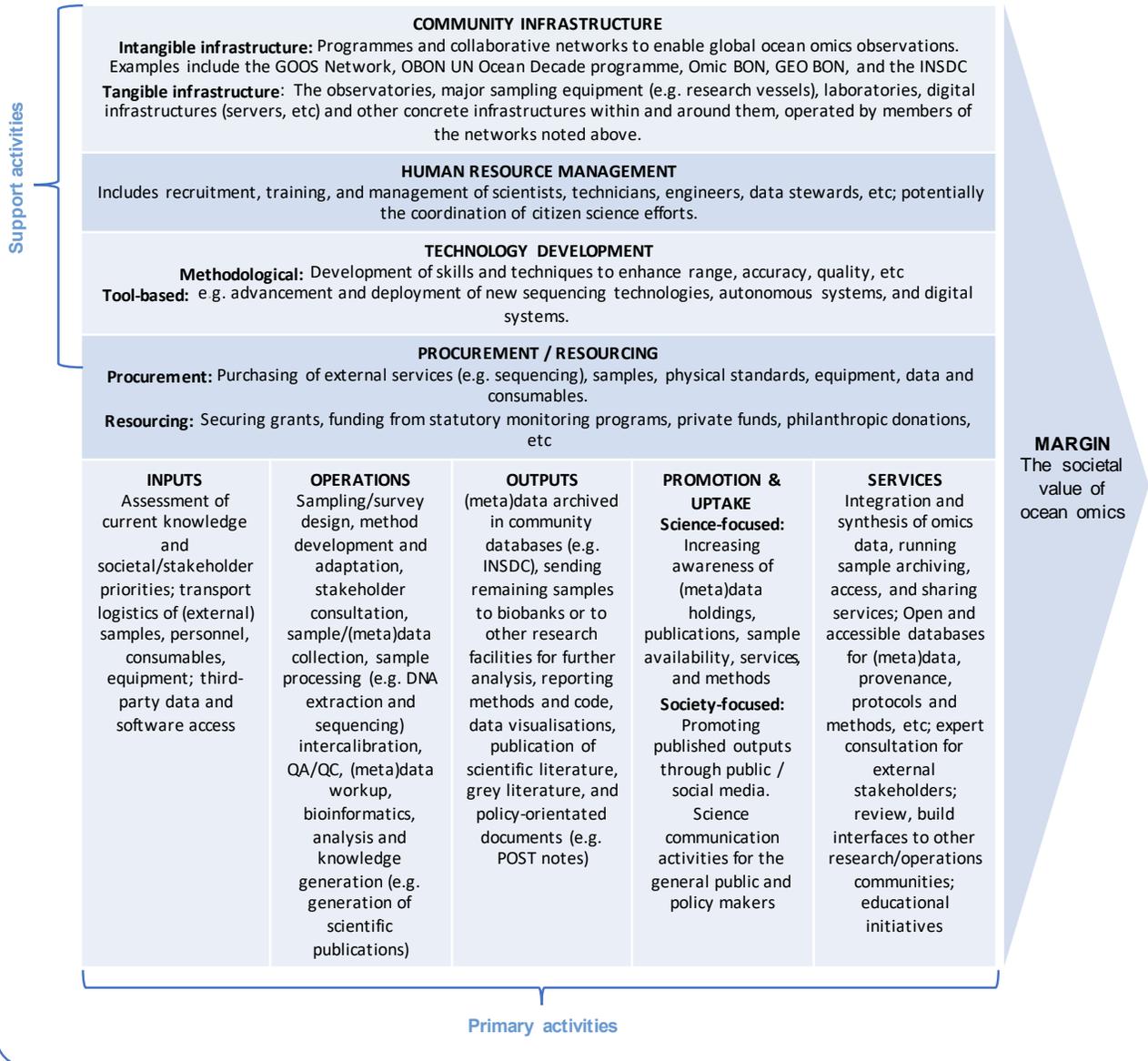
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- 415 **11 Supplementary Material**

**Ocean Omics Value Chain**

Applying Porter’s value chain concept to ocean omics research can help to systematically identify activities that will increase the value of ocean omics research to society at large: A well-functioning ocean omics value chain will generate more downstream value than the costs needed to operate it. This is the basis of justifying increases or decreases in investment in any activity along or across the chain. We believe such mechanisms will help the omics community coherently reflect on the activities needed to advance our methods while ensuring a healthy cost/benefit ratio, especially as we interface with other ocean communities and their own value chains. For example, when considering the uptake of new technologies (e.g. new sequencing technologies), value chain analysis helps to consider whether uptake is worth the disruption to the continuity of long-term ocean omics observatories. The process helps to evaluate what value is compromised (i.e. Are the time series broken? Are the POST notes valid anymore?) and what needs co-investment to mitigate it (e.g. intercalibration across space and time). Essentially answering “is it worth it?” at scale, or should it be a more isolated test by a smaller community (e.g. a scientific team/pilot study).



416

417 **Supplementary Figure 1.** We frame part of our perspective in an adaptation of Porter’s (1985) value chain, where a product passes  
 418 through all portions of the chain, gaining value from each activity. The language used in Porter’s value chain relates to commercial  
 419 business activities, this adaptation gives examples for the types of omic research activities that could apply to each category. Category  
 420 headings have been adapted to fit omics operations and research, as follows: Firm infrastructure → Community infrastructure,  
 421 Procurement → Procurements / Resourcing, Inbound logistics → Inputs, Outbound logistics → Outputs, Marketing & Sales →  
 422 Promotion & Uptake (note that original headings would be appropriate for omics-focused businesses). It should also be noted that

423 value within omics operations and research does not only refer to monetary transactions (for example, procurement may be facilitated  
424 by credit on scientific publications).

425