

Unbiased Molecular Dynamics of 11 min Timescale Drug Unbinding Reveals Transition State Stabilizing Residues

Samuel D. Lotz* and Alex Dickson*+

*Dept. Biochemistry & Molecular Biology and +Dept. of Computational Mathematics, Science and Engineering
Michigan State University, East Lansing, MI

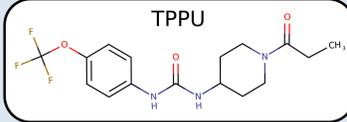


Connect:
salotz@salotz.info
ORCID: 000-0001-6159-615X
twitter: @real_salotz
github: @salotz
linkedin: in/salotz

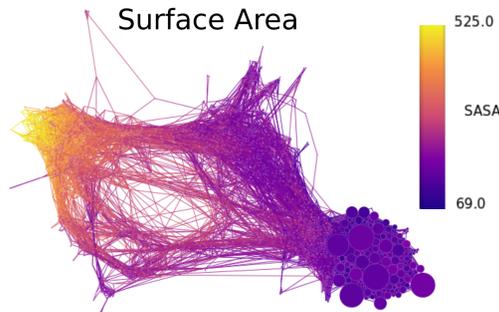


Introduction

Recent interest in drug binding kinetics (1) has motivated all atom molecular dynamics simulations of ligand (un)binding which has only become feasible due to increases in computational power and improvements in sampling algorithms. We report sampling of the unbinding of a drug-like inhibitor (TPPU) of the protein soluble epoxide hydrolase (sEH) with an experimentally determined residence time of 11 min. We conclude that **specific interactions with residues at the edges of the binding site are important stabilizers of the unbinding transition state**. The specific structural details of these interactions will be of interest to medicinal chemists designing drugs with longer residence times, which has been shown to improve efficacy. sEH is an important target for treating a number of diseases such as diabetic neuropathic pain.



Solvent Accessible Surface Area

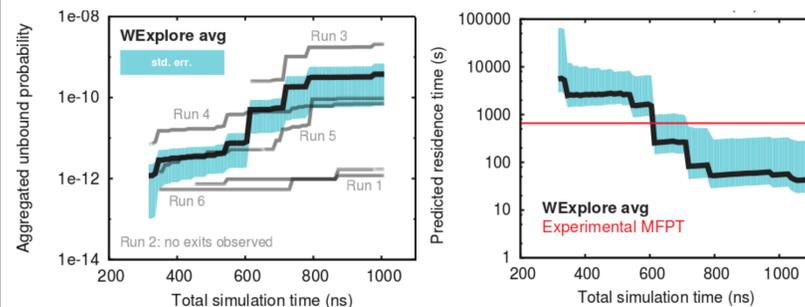


Solvent accessible surface area (SASA) reports the progress of ligand unbinding. The free energy of the network shows that the upper branch (P₁) is more probable than the lower. P₁ is the shortest path leaving the wide sEH binding site.

Free Energy

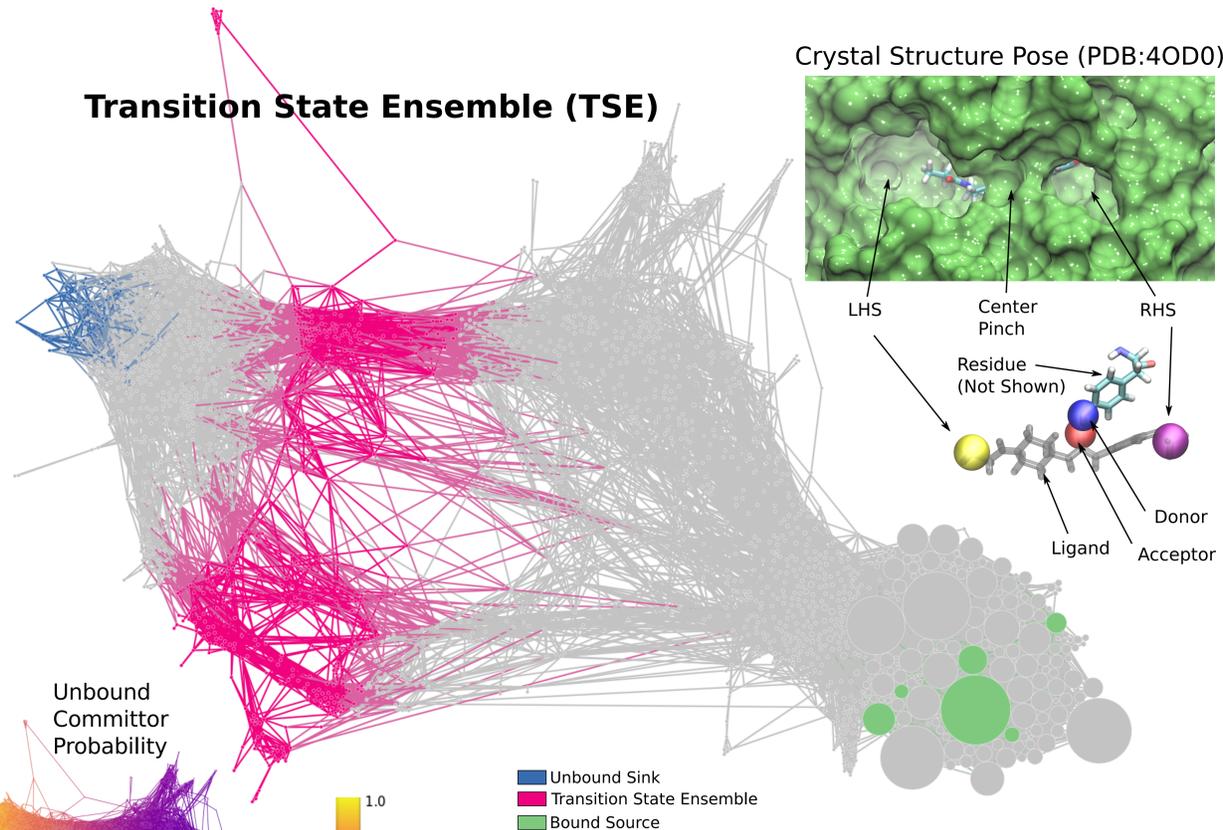


Residence Time

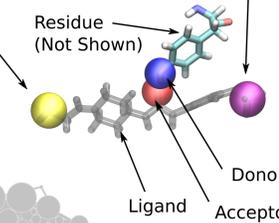
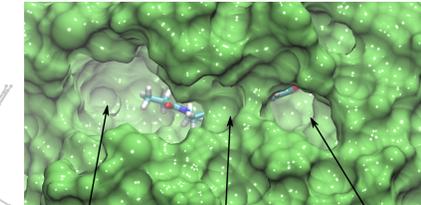


The sampling method used was WExplore (8) a variation on the weighted ensemble method of Huber and Kim (9). Six independent simulations were run accumulating a total of 7.0 microseconds of sampling in which 75 exit points were observed. Residence times (1/MFPT) were calculated from the time for each exit point. Our final value (42 s) is relatively close to the experimental value (11 min) given the timescales involved.

Transition State Ensemble (TSE)



Crystal Structure Pose (PDB:4OD0)

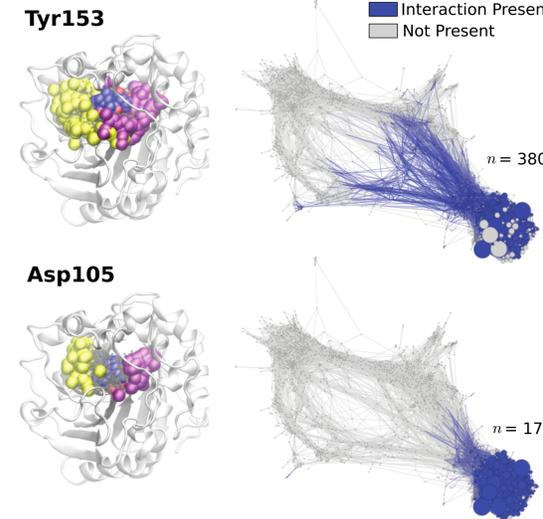


Unbound Committor Probability

Blue Unbound Sink
Pink Transition State Ensemble
Green Bound Source

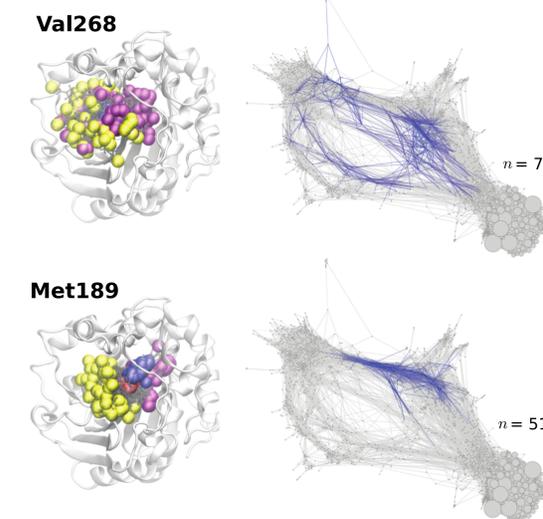
Committer probabilities are calculated using the source and sink nodes shown above (5). The transition state is defined as the nodes with a committor value between 0.4 and 0.6.

Bound State Stabilizing Interactions

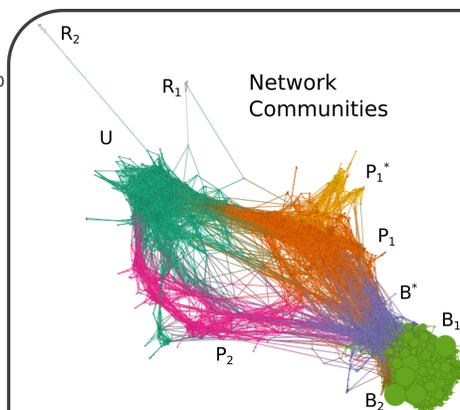


Hydrogen bonds in the native state play different roles. Asp105 only stabilizes the fully bound state, while Tyr153 stabilizes a larger set of states, facilitating outward movement of the ligand.

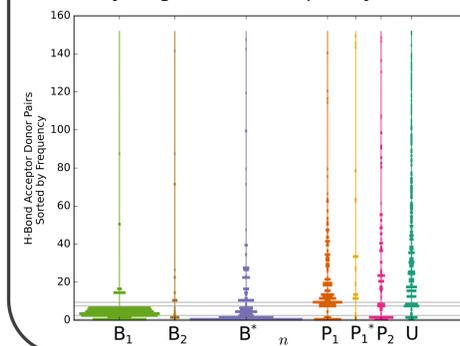
Transition State Stabilizing Interactions



Interactions with specific ligands stabilize different ligand exit pathways at the overall transition state. Met189 stabilizes movement along the RHS of the binding site (P₁) and is the highest frequency interaction in P₁ coincident with the TSE. Val268 stabilizes ligand poses at the center of the binding site straddling both P₁ and P₂. **Met189 is likely a stabilizer of bottlenecks that determine (un)binding kinetics for inhibitors of sEH.**



Hydrogen Bond Frequency Patterns



Using a network modularity metric the network was divided up into distinct communities. The main communities are B₁ (bound), B* (exit branchpoint), P₁ (path 1), P₂ (path 2), and U (unbound). R₁ and R₂ were found to have a reversal in ligand orientation. Profiling hydrogen bonds of each community shows that **in the bound state a handful of interactions are present in high frequency, while in B* fewer interactions are stabilizing (Tyr153)**. In P₁ interactions are more diverse but occur less frequently and are distinct from all other communities. The unbound ensemble has a large number of low frequency interactions as would be expected. Gray bars indicate interactions highlighted in the boxes to the right.

Literature Cited

- Lee, K. S. et al. J Med Chem, 57(16), 7016-7030 (2014).
- Casasnovas, R. et al., JACS, (2017).
- Spagnuolo et al., Journal of the American Chemical Society, 139(9), 3417-3429 (2017).
- Copeland, R. A., Nat Rev Drug Discov, 15(2), 87-95 (2016).
- Beauchamp, K. A., et al., Journal of Chemical Theory and Computation, 7(10), 3412-3419 (2011).
- Swinney, D. C., Pharmaceutical Medicine, 22(1), 23-34 (2008).
- Lu, H., & Tonge, P. J., Current Opinion in Chemical Biology, 14(4), 467-474 (2010).
- Dickson, A., & Brooks, C. L., J Phys Chem B, 118(13), 3532-3542 (2014).
- Huber, G. A., & Kim, S., Biophys J, 70(1), 97-110 (1996).