

# XCMS Workshop

Alignments with OBI-Warp  
Annotation using CAMERA

Ralf Tautenhahn

The Scripps Research Institute

5/31/2009

# OBI-Warp

John T. Prince and Edward M. Marcotte

## **Chromatographic Alignment of ESI-LC-MS Proteomics Data Sets by Ordered Bijective Interpolated Warping**

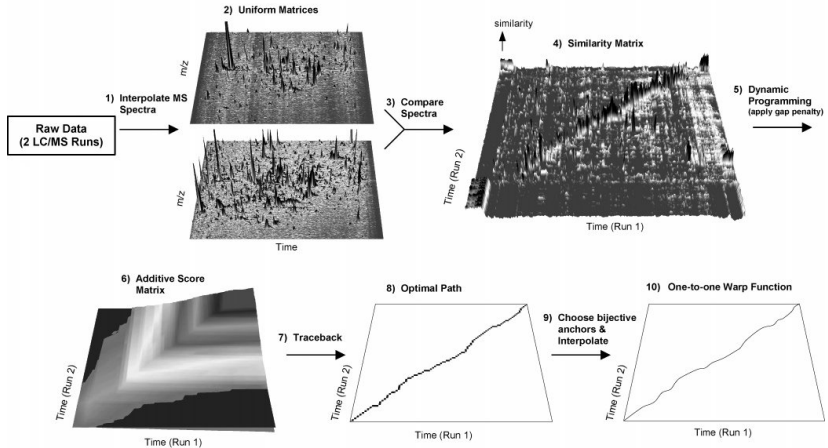
*Analytical Chemistry*, **2060** 78 (17), 6140-6152

`obi-warp.sourceforge.net`

- Retention time correction based on spectra similarity
- Doesn't rely on detected feature
- No initial grouping needed
- Potentially useful for samples with high retention time deviations
  
- Recently integrated into XCMS
- CAVE: still experimental!

# OBI-Warp

## 1) Alignment by OBI-Warp



# Retention time correction using OBI-Warp

```
library(xcms)
cdfpath <- system.file("cdf", package = "faahKO")
cdffiles <- list.files(cdfpath, recursive = TRUE, full.names = TRUE)
xset <- xcmsSet(cdffiles)
```

## OBI-Warp

```
xrc <- retcor(xset,
  method="obiwarp")
```

## LOESS

```
xg <- group(xset)
xrc <- retcor(xg)
```

Additional parameters see `?retcor.obiwarp`

# Retention time correction using OBI-Warp

```
library(xcms)
cdfpath <- system.file("cdf", package = "faahKO")
cdffiles <- list.files(cdfpath, recursive = TRUE, full.names = TRUE)
xset <- xcmsSet(cdffiles)
```

## OBI-Warp

```
xrc <- retcor(xset,
  method="obiwarp")
```

## LOESS

```
xg <- group(xset)
xrc <- retcor(xg)
```

final grouping

```
x <- group(xrc)
```

Additional parameters see `?retcor.obiwarp`

# Retention time correction using OBI-Warp

```
library(xcms)
cdfpath <- system.file("cdf", package = "faahKO")
cdffiles <- list.files(cdfpath, recursive = TRUE, full.names = TRUE)
xset <- xcmsSet(cdffiles)
```

## OBI-Warp

```
xrc <- retcor(xset,
  method="obiwarp")
```

## LOESS

```
xg <- group(xset)
xrc <- retcor(xg)
```

final grouping

```
x <- group(xrc)
```

OR

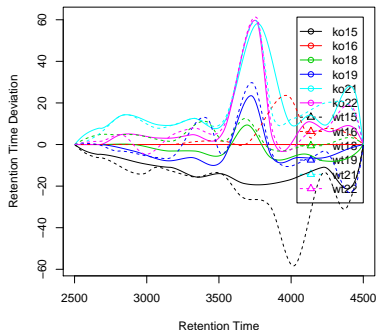
```
x <- group(xrc, method="nearest")
```

Additional parameters see `?retcor.obiwarp`

# Example: faahKO dataset

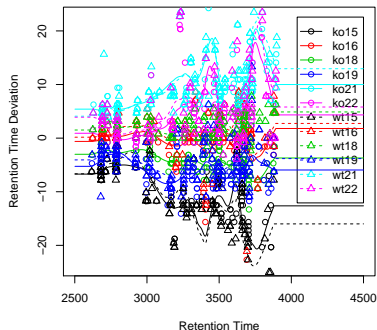
## OBI-Warp

Retention Time Deviation vs. Retention Time



## LOESS

Retention Time Deviation vs. Retention Time



# Alignment using the “nearest” algorithm

- alternative to `group.density` (default)
- inspired by the alignment algorithm of `mzMine`
- main difference: no binning necessary
- usage

```
xsg <- group(xs,method="nearest", mzCheck=0.2,  
             rtCheck=15, kNN=10)
```

- *mzCheck* maximum tolerated distance for *m/z*
- *rtCheck* maximum tolerated distance for RT.
- *kNN* number of nearest neighbors to check

`?group.nearest`

To use this method, the R-package *RANN* has to be installed.

<http://rforge.net/RANN/>



# Annotation of LC/MS Features

<i>m/z</i>	Retention time	Intensity
135.05	280.43	97554
153.06	280.43	4207
175.04	280.43	7468
197.02	280.76	1015
117.04	278.07	995
145.03	278.07	20534
149.06	278.07	3561
177.05	278.07	31096
195.06	278.07	1925
217.05	277.74	1704
233.01	278.07	3541
427.07	278.07	1897
621.13	278.07	435
	⋮	
	⋮	

# Annotation of LC/MS Features

<i>m/z</i>	Retention time	Intensity	Compound No.
135.05	280.43	97554	1
153.06	280.43	4207	1
175.04	280.43	7468	1
197.02	280.76	1015	1
117.04	278.07	995	2
145.03	278.07	20534	2
149.06	278.07	3561	2
177.05	278.07	31096	2
195.06	278.07	1925	2
217.05	277.74	1704	2
233.01	278.07	3541	2
427.07	278.07	1897	2
621.13	278.07	435	2
⋮			

- Grouping of all features belonging to the same compound

# Annotation of LC/MS Features

<i>m/z</i>	Retention time	Intensity	Ion	Compound No.
135.05	280.43	97554		1
153.06	280.43	4207	[M+H] <sup>+</sup>	1
175.04	280.43	7468	[M+Na] <sup>+</sup>	1
197.02	280.76	1015		1
117.04	278.07	995		2
145.03	278.07	20534		2
149.06	278.07	3561		2
177.05	278.07	31096		2
195.06	278.07	1925	[M+H] <sup>+</sup>	2
217.05	277.74	1704	[M+Na] <sup>+</sup>	2
233.01	278.07	3541	[M+K] <sup>+</sup>	2
427.07	278.07	1897	[2M+K] <sup>+</sup>	2
621.13	278.07	435	[3M+K] <sup>+</sup>	2

⋮

- Grouping of all features belonging to the same compound
- Detect charge state
- Assignment of ion species

# Annotation of LC/MS Features

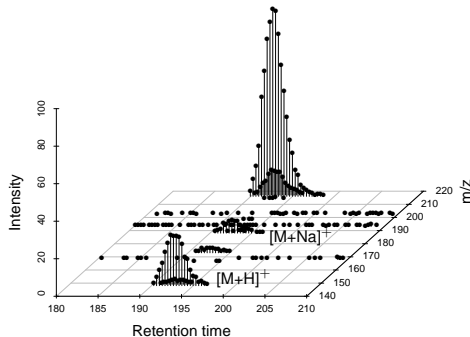
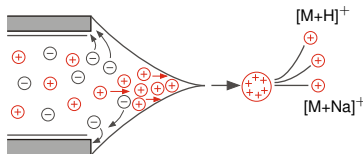
<i>m/z</i>	Retention time	Intensity	Ion	Molecular mass	Compound No.
135.05	280.43	97554			1
153.06	280.43	4207	[M+H] <sup>+</sup>	} <i>M</i> = 152.05	1
175.04	280.43	7468	[M+Na] <sup>+</sup>		1
197.02	280.76	1015			1
117.04	278.07	995			2
145.03	278.07	20534			2
149.06	278.07	3561			2
177.05	278.07	31096			2
195.06	278.07	1925	[M+H] <sup>+</sup>	} <i>M</i> = 194.05	2
217.05	277.74	1704	[M+Na] <sup>+</sup>		2
233.01	278.07	3541	[M+K] <sup>+</sup>		2
427.07	278.07	1897	[2M+K] <sup>+</sup>		2
621.13	278.07	435	[3M+K] <sup>+</sup>		2
	⋮				

- Grouping of all features belonging to the same compound
- Detect charge state
- Assignment of ion species
- Calculate molecular mass

→ Database search, molecular formula calculation

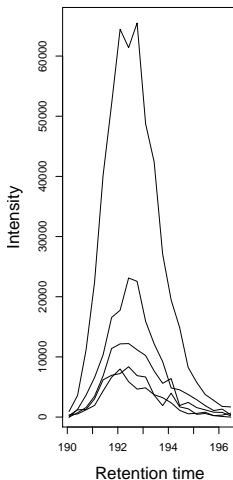
# Grouping all features of the same compound (1)

- Electrospray ionization
  - Molecule M: Formation of different ion species e.g.  $[M+H]^+$ ,  $[M+Na]^+$
  - in the same ratio
- Features belonging to the same compound
  - Same retention time
  - Feature intensities have a linear relationship



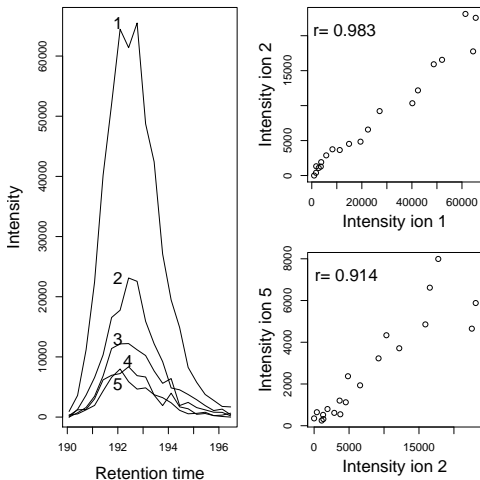
# Grouping all features of the same compound (1)

- Electrospray ionization
  - Molecule M: Formation of different ion species e.g.  $[M+H]^+$ ,  $[M+Na]^+$
  - in the same ratio
- Features belonging to the same compound
  - Same retention time
  - Feature intensities have a linear relationship



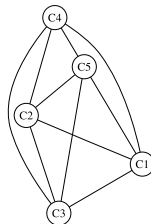
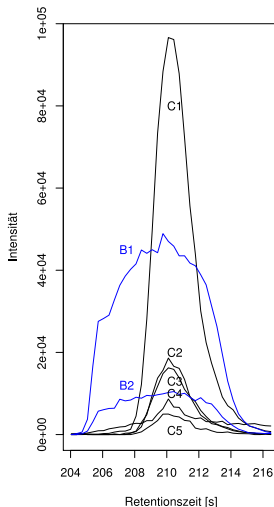
# Grouping all features of the same compound (1)

- Electrospray ionization
  - Molecule M: Formation of different ion species e.g.  $[M+H]^+$ ,  $[M+Na]^+$ 
    - in the same ratio
- Features belonging to the same compound
  - Same retention time
  - Feature intensities have a linear relationship



# Grouping all features of the same compound (2)

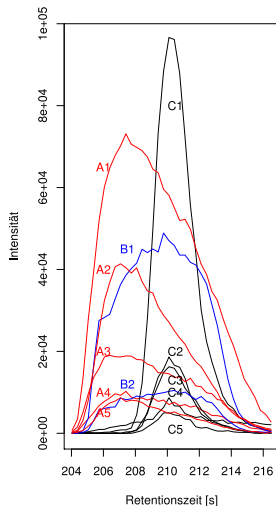
- Compute correlation coefficients
- Create graph
  - Node: Feature
  - Edge:  $r > r_{Thr}$





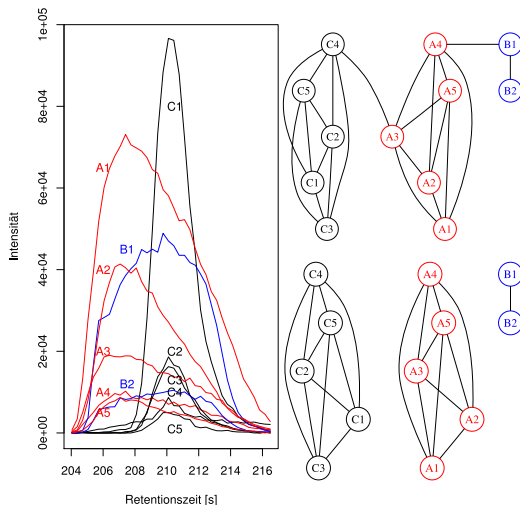
# Grouping all features of the same compound (2)

- Compute correlation coefficients
- Create graph
  - Node: Feature
  - Edge:  $r > r_{Thr}$



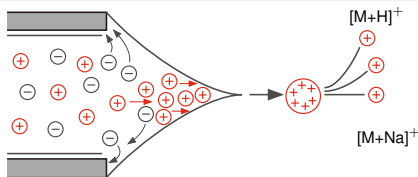
# Grouping all features of the same compound (2)

- Compute correlation coefficients
- Create graph
  - Node: Feature
  - Edge:  $r > r_{Thr}$
- HCS-Clustering [Hartuv and Shamir 2000]
- highly connected subgraphs



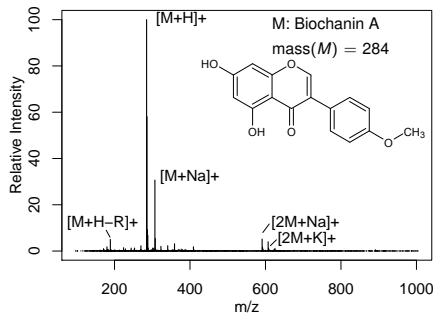
# Assignment of ion species (1)

- Ion formation depends on the compound



# Assignment of ion species (1)

- Ion formation depends on the compound
  - quasi-molecular ion, e.g.  $[M+H]^+$
  - cluster ions, e.g.  $[2M+Na]^+$
  - fragment ions, e.g.  $[M+H-R]^+$

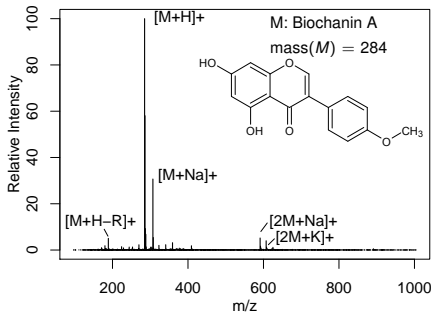


# Assignment of ion species (1)

- Ion formation depends on the compound
  - quasi-molecular ion, e.g.  $[M+H]^+$
  - cluster ions, e.g.  $[2M+Na]^+$
  - fragment ions, e.g.  $[M+H-R]^+$

Observed  $m/z$  values for a single ion species:

$$s = \frac{n \cdot \text{mass}(M) + \text{mass}(I) - \text{mass}(R)}{z}$$

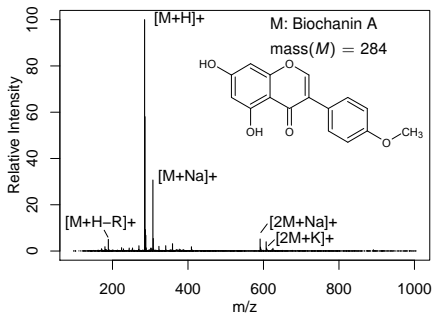


# Assignment of ion species (1)

- Ion formation depends on the compound
  - quasi-molecular ion, e.g.  $[M+H]^+$
  - cluster ions, e.g.  $[2M+Na]^+$
  - fragment ions, e.g.  $[M+H-R]^+$

Observed  $m/z$  value  $s$  for a single ion species:

$$s = \frac{n \cdot \text{mass}(M) + \text{mass}(I) - \text{mass}(R)}{z}$$



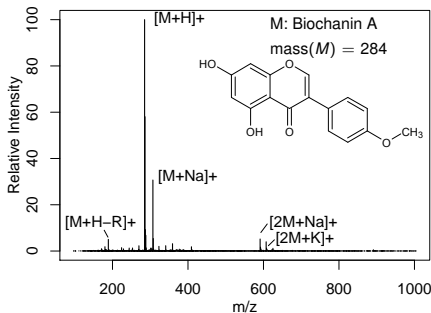
## Example

$[M+Na]^+$ ,  $\text{mass}(M)=284$ ,  
 $n=1, z=1$ ,  $\text{mass}(I)=23$

$$\begin{aligned} s &= \frac{1 \cdot 284 + 23}{1} \\ &= 307 \, m/z \end{aligned}$$

# Assignment of ion species (1)

- Ion formation depends on the compound
  - quasi-molecular ion, e.g.  $[M+H]^+$
  - cluster ions, e.g.  $[2M+Na]^+$
  - fragment ions, e.g.  $[M+H-R]^+$



Observed  $m/z$  value  $s$  for a single ion species:

$$s = \frac{n \cdot \text{mass}(M) + \text{mass}(I) - \text{mass}(R)}{z}$$

$$\text{mass}(M) = \frac{z \cdot s - \text{mass}(I) + \text{mass}(R)}{n}$$

$n, z > 0$

## Example

$[M+Na]^+$ ,  $\text{mass}(M)=284$ ,  
 $n=1, z=1$ ,  $\text{mass}(I)=23$

$$s = \frac{1 \cdot 284 + 23}{1}$$

$$= 307 \text{ } m/z$$

# Assignment of ion species (2)

feature group
$s_i [m/z]$
270.05
285.07
307.06
591.12
607.09
...

rule table ( $z, n, l, R$ )	
rule $j$	mass( $l_j$ )
$[M+H]^+$	1.01
$[M+2H]^{2+}$	2.01
$[M+Na]^+$	22.99
$[M+K]^+$	38.96
$[2M+Na]^+$	22.99
$[2M+K]^+$	38.96
...	



# Assignment of ion species (2)

- Creation of mass-hypotheses

$$M_{i,j} = \frac{z_j \cdot s_i - l_j + R_j}{n_j}$$

feature group

$s_i$ [ $m/z$ ]
270.05
285.07
307.06
591.12
607.09
...

rule table ( $z, n, l, R$ )	
rule $j$	mass( $l_j$ )
$[M+H]^+$	1.01
$[M+2H]^{2+}$	2.01
$[M+Na]^+$	22.99
$[M+K]^+$	38.96
$[2M+Na]^+$	22.99
$[2M+K]^+$	38.96
...	

# Assignment of ion species (2)

- Creation of mass-hypotheses

$$M_{i,j} = \frac{z_j \cdot s_i - l_j + R_j}{n_j}$$

feature group
$s_i$ [ $m/z$ ]
270.05
285.07
307.06
591.12
607.09
...

rule table ( $z, n, l, R$ )	
rule $j$	mass( $l_j$ )
$[M+H]^+$	1.01
$[M+2H]^{2+}$	2.01
$[M+Na]^+$	22.99
$[M+K]^+$	38.96
$[2M+Na]^+$	22.99
$[2M+K]^+$	38.96
...	

$$M_{i,j} =$$

269	538.1	247.1	231.1	123.5	115.5
284.1	568.1	262.1	246.1	131	123.1
306.1	612.1	284.1	268.1	142	134.1
590.1	1180.2	568.1	552.2	284.1	276.1
606.1	1212.2	584.1	568.1	292.1	284.1

# Assignment of ion species (2)

## ● Creation of mass-hypotheses

$$M_{i,j} = \frac{z_j \cdot s_i - l_j + R_j}{n_j}$$

$$\begin{aligned} \text{e.g. } M_{5,6} &= \frac{1 \cdot 607.09 - 38.96 + 0}{2} \\ &= 284.1 \end{aligned}$$

feature group

$s_i$ [m/z]
270.05
285.07
307.06
591.12
607.09
...

rule table ( $z, n, l, R$ )

rule $j$	mass( $l_j$ )
[M+H] <sup>+</sup>	1.01
[M+2H] <sup>2+</sup>	2.01
[M+Na] <sup>+</sup>	22.99
[M+K] <sup>+</sup>	38.96
[2M+Na] <sup>+</sup>	22.99
[2M+K] <sup>+</sup>	38.96
...	

$$M_{i,j} =$$

269	538.1	247.1	231.1	123.5	115.5
284.1	568.1	262.1	246.1	131	123.1
306.1	612.1	284.1	268.1	142	134.1
590.1	1180.2	568.1	552.2	284.1	276.1
606.1	1212.2	584.1	568.1	292.1	284.1

# Assignment of ion species (2)

## ● Creation of mass-hypotheses

$$M_{i,j} = \frac{z_j \cdot s_i - l_j + R_j}{n_j}$$

$$\begin{aligned} \text{e.g. } M_{5,6} &= \frac{1 \cdot 607.09 - 38.96 + 0}{2} \\ &= 284.1 \end{aligned}$$

feature group		
$s_i$ [ $m/z$ ]	Ion	Mass
270.05		
285.07	[M+H] <sup>+</sup>	} M=284.1
307.06	[M+Na] <sup>+</sup>	
591.12	[2M+Na] <sup>+</sup>	
607.09	[2M+K] <sup>+</sup>	
...		

rule table ( $z, n, l, R$ )	
rule $j$	mass( $l_j$ )
[M+H] <sup>+</sup>	1.01
[M+2H] <sup>2+</sup>	2.01
[M+Na] <sup>+</sup>	22.99
[M+K] <sup>+</sup>	38.96
[2M+Na] <sup>+</sup>	22.99
[2M+K] <sup>+</sup>	38.96
...	

$$M_{i,j} =$$

269	538.1	247.1	231.1	123.5	115.5
284.1	568.1	262.1	246.1	131	123.1
306.1	612.1	284.1	268.1	142	134.1
590.1	1180.2	568.1	552.2	284.1	276.1
606.1	1212.2	584.1	568.1	292.1	284.1

# Usage

- <http://bioconductor.org/packages/devel/bioc/html/CAMERA.html>

- or (within R) type:

```
source("http://bioconductor.org/biocLite.R")
biocLite("CAMERA")
```

- example using the faahKO dataset

```
library(CAMERA)
xs <- xcmsSet(...)
## group, retcor, etc.

an <- annotate(xs)
peaklist <- getPeaklist(an)
write.csv(peaklist, file='xsannotated.csv')
```

# Interpretation of the Results

id	mz	rt	isotopes	adduct	pc
65	176.04	280.09			
76	136.05	280.43	[14][M+1]1+ [14][M]1+		5
77	135.05	280.43			5
74	153.06	280.43		[M+H]+ 152.05437	5
75	175.04	280.43		[M+Na]+ 152.05437	5
73	197.02	280.76		[M+2Na-H]+ 152.05437	5
78	377.74	286.15			
79	732.5	286.49			
83	488.32	286.82		[M+Na]+ 465.33205	7
82	466.34	286.82		[M+H]+ 465.33205	7
...					