**Methods**

Hela cells were seeded in 12 well plates at density 1e5 cells/ml, respectively, and next day co-transfected with 0.1 µg GFP-tagged histone and: empty vector (0.9 µg) or empty vector (0.5 and 0.1 µg) + Hdac11 (WT or Y304H and H142,143A catalytic mutant) -FLAG (0.4 or 0.8 µg) using  XtremeGene HP transfection reagent, following manufacturer instructions. After 4 h media was removed and cells were treated with or without HDAC11 inhibitors SIS17, SIS7 (control) and FT895. Cell lysates were analysed in western blot for H3K18Ac levels normalized to GFP.

**Western blot**

Total cell lysates were resolved in 4-12% Bis-Tris Protein Gels (Invitrogen) and transferred in for 1.5 h (80 V) onto PVDF membrane (Millipore) in Tris-Glycine transfer buffer containing 20% MeOH and 0.05% SDS. Blots were blocked for 1 h in blocking buffer (5% milk in PBS) and incubated with primary antibodies anti-H3K18Ac (CST,#9675)and anti-GFP (Clontech, NC9777966) in blocking buffer (5% BSA in PBST: 0.1% Tween 20 PBS) overnight at 4 °C. After five washes with PBST the blots were incubated with goat-anti rabbit (IR800) and donkey anti-mouse (IR 680) antibodies in Odyssey Blocking Buffer (LiCor) for 1 h at RT and washed five times with PBST. The signal was read on an Odyssey scanner (LiCor) at 800 nm and 700 nm.