

# Thermodynamically constrain a Recon3D

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**Reviewers:**

## INTRODUCTION

In flux balance analysis of genome scale stoichiometric models of metabolism, the principal constraints are uptake or secretion rates, the steady state mass conservation assumption and reaction directionality. Von Bertalanffy [1,4] is a set of methods for (i) quantitative estimation of thermochemical parameters for metabolites and reactions using the component contribution method [3], (ii) quantitative assignment of reaction directionality in a multi-compartmental genome scale model based on an application of the second law of thermodynamics to each reaction [2], (iii) analysis of thermochemical parameters in a network context, and (iv) thermodynamically constrained flux balance analysis. The theoretical basis for each of these methods is detailed within the cited papers.

## PROCEDURE

### Configure the environment

The default COBRA Toolbox paths are automatically changed here to work on the new version of vonBertalanffy

```
aPath = which('initVonBertalanffy');
basePath = strrep(aPath,['vonBertalanffy' filesep 'initVonBertalanffy.m'],'');
addpath(genpath(basePath))
folderPattern=[filesep 'old'];
method = 'remove';
editCobraToolboxPath(basePath,folderPattern,method)
```

```
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/componentContribution/old
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/directionalityReport/old
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/groupContribution/old
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/inchi/old
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/molFiles/old
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/protons/old
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/trainingModel/old
```

```
aPath = which('initVonBertalanffy');
basePath = strrep(aPath,['vonBertalanffy' filesep 'initVonBertalanffy.m'],'');
addpath(genpath(basePath))
folderPattern=[filesep 'new'];
method = 'add';
editCobraToolboxPath(basePath,folderPattern,method)
```

```
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/componentContribution/new
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/groupContribution/new
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/inchi/new
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/molFiles/new
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/protons/new
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/trainingModel/new
```

All the installation instructions are in a separate .md file named vonBertalanffy.md in docs/source/installation

With all dependencies installed correctly, we configure our environment, verify all dependencies, and add required fields and directories to the matlab path.

```
initVonBertalanffy
```

ChemAxon Marvin Beans is installed and working.

```
linux-vdso.so.1 (0x00007ffdddfcf000)
libc.so.6 => /lib/x86_64-linux-gnu/libc.so.6 (0x00007fffladeef000)
libopenbabel.so.5 => /usr/lib/libopenbabel.so.5 (0x00007fffladc9f000)
libstdc++.so.6 => /usr/lib/x86_64-linux-gnu/libstdc++.so.6 (0x00007ffflada85000)
libgcc_s.so.1 => /usr/local/bin/MATLAB/R2021a/sys/os/glnxa64/libgcc_s.so.1 (0x00007ffflad86d000)
/lib64/ld-linux-x86-64.so.2 (0x00007ffflae10f000)
libdl.so.2 => /lib/x86_64-linux-gnu/libdl.so.2 (0x00007ffflad865000)
libz.so.1 => /lib/x86_64-linux-gnu/libz.so.1 (0x00007ffflad849000)
libm.so.6 => /lib/x86_64-linux-gnu/libm.so.6 (0x00007ffflad6fa000)
libgomp.so.1 => /usr/lib/x86_64-linux-gnu/libgomp.so.1 (0x00007ffflad6b5000)
libpthread.so.0 => /lib/x86_64-linux-gnu/libpthread.so.0 (0x00007ffflad692000)
```

babel must depend on the system libstdc++.so.6 not the one from MATLAB

Trying to edit the 'LD\_LIBRARY\_PATH' to make sure that it has the correct system path before the Matlab path

The solution will be arch dependent

## Select the model

This tutorial is tested for the E. coli model iAF1260 and the human metabolic model Recon3Dmodel. However, only the data for the former is provided within the COBRA Toolbox as it is used for testing von Bertalanffy. However, the figures generated below are most suited to plotting results for Recon3D, so they may not be so useful for iAF1260. The Recon3D example uses values from literature for input variables where they are available.

```
%modelName = 'iAF1260';
modelName='Ec_iAF1260_flux1';
% uncomment this line and comment the line below if you want to use the other model- c
modelName='Recon3DModel_301';
```

## Load a model

Load a model, and save it as the original model in the workspace, unless it is already loaded into the workspace.

```
clear model
global CBTDIR
modelName = [modelName '.mat']
```

```
modelName =
'Recon3DModel_301.mat'
```

```
modelDirectory = getDistributedModelFolder(modelFileName); %Look up the folder for the
modelName= [modelDirectory filesep modelName]; % Get the full path. Necessary to
```

```

switch modelName
case 'Ec_iAF1260_flux1'
    modelFileName = [modelName '.xml'];
    model = readCbModel(modelFileName);
    if model.S(952, 350)==0
        model.S(952, 350)=1; % One reaction needing mass balancing in iAF1260
    end
    model.metCharges(strcmp('asntrna[Cytosol]', model.mets))=0; % One reaction needing ch

case 'iAF1260'
    model = readCbModel(modelFileName);
    model.mets = cellfun(@(mets) strrep(mets, '_c', '[c]'), model.mets, 'UniformOutput', false);
    model.mets = cellfun(@(mets) strrep(mets, '_e', '[e]'), model.mets, 'UniformOutput', false);
    model.mets = cellfun(@(mets) strrep(mets, '_p', '[p]'), model.mets, 'UniformOutput', false);
    bool = strcmp(model.mets, 'lipa[c]old[c]');
    model.mets{bool}='lipa_old_[c]';
    bool = strcmp(model.mets, 'lipa[c]old[e]');
    model.mets{bool}='lipa_old_[e]';
    bool = strcmp(model.mets, 'lipa[c]old[p]');
    model.mets{bool}='lipa_old_[p]';
    if model.S(952, 350)==0
        model.S(952, 350)=1; % One reaction needing mass balancing in iAF1260
    end
    model.metCharges(strcmp('asntrna[c]', model.mets))=0; % One reaction needing ch

case 'Recon3DModel_Dec2017'
    model = readCbModel(modelFileName);
    model.csense(1:size(model.S,1),1)='E';
    %Hack for thermodynamics
    model.metFormulas{strcmp(model.mets, 'h[i]')}='H';
    model.metFormulas(cellfun('isempty', model.metFormulas)) = {'R'};
    if isfield(model, 'metCharge')
        model.metCharges = double(model.metCharge);
        model=rmfield(model, 'metCharge');
    end
    modelOrig = model;
case 'Recon3DModel_301'
    model = readCbModel(modelFileName);
    %Hack for thermodynamics
    model.metFormulas(cellfun('isempty', model.metFormulas)) = {'R'};
    modelOrig = model;
otherwise
    error('setup specific parameters for your model')
end
end

```

Each model.subSystems{x} has been changed to a character array.

## Set the directory containing the results

```

switch modelName
case 'Ec_iAF1260_flux1'
    resultsPath=which('tutorial_vonBertalanffy.mlx');
    resultsPath=strrep(resultsPath, '/tutorial_vonBertalanffy.mlx', '');
    resultsPath=[resultsPath filesep modelName '_results'];

```

```

        resultsBaseFileName=[resultsPath filesep modelName '_results'];
    case 'iAF1260'
        resultsPath=which('tutorial_vonBertalanffy.mlx');
        resultsPath=strrep(resultsPath, '/tutorial_vonBertalanffy.mlx', '');
        resultsPath=[resultsPath filesep modelName '_results'];
        resultsBaseFileName=[resultsPath filesep modelName '_results'];
    case 'Recon3DModel_Dec2017'
        basePath=~'/work/sbgCloud';
        resultsPath=[basePath '/programReconstruction/projects/recon2models/results/the
        resultsBaseFileName=[resultsPath filesep modelName '_' datestr(now,30) '_'];
    case 'Recon3DModel_301'
        basePath=[~' filesep 'work' filesep 'sbGCloud'];
        resultsPath=which('tutorial_vonBertalanffy.mlx');
        resultsPath=strrep(resultsPath,[filesep 'tutorial_vonBertalanffy.mlx'], '');
        resultsPath=[resultsPath filesep modelName '_results'];
        resultsBaseFileName=[resultsPath filesep modelName '_results_'];
    otherwise
        error('setup specific parameters for your model')
end

```

## Set the directory containing molfiles

```

switch modelName
    case 'Ec_iAF1260_flux1'
        molFileDir = 'iAF1260Molfiles';
    case 'iAF1260'
        molFileDir = 'iAF1260Molfiles';
    case 'Recon3DModel_Dec2017'
        molFileDir = [basePath '/data/metDatabase/explicit/molFiles'];
        %molFileDir = [basePath '/programModelling/projects/atomMapping/results/molFile
        %molFileDir = [basePath '/programModelling/projects/atomMapping/results/molFile
    case 'Recon3DModel_301'
        ctfPath = [basePath filesep 'code' filesep 'fork-ctf'];
        % system(['git clone https://github.com/opencobra/ctf' ctfPath])
        molFileDir = [basePath filesep 'code' filesep 'fork-ctf' filesep 'mets' filesep
    otherwise
        molFileDir = [basePath '/code/fork-ctf/mets/molFiles'];
end

```

## Set the thermochemical parameters for the model

```

switch modelName
    case 'Ec_iAF1260_flux1'
        T = 310.15; % Temperature in Kelvin
        compartments = {'Cytosol'; 'Extra_organism'; 'Periplasm'}; % Cell compartment i
        ph = [7.7; 7.7; 7.7]; % Compartment specific pH
        is = [0.25; 0.25; 0.25]; % Compartment specific ionic strength in mol/L
        chi = [0; 90; 90]; % Compartment specific electrical potential relative to cyto
    case 'iAF1260'
        T = 310.15; % Temperature in Kelvin
        compartments = ['c'; 'e'; 'p']; % Cell compartment identifiers
        ph = [7.7; 7.7; 7.7]; % Compartment specific pH
        is = [0.25; 0.25; 0.25]; % Compartment specific ionic strength in mol/L

```

```

        chi = [0; 90; 90]; % Compartment specific electrical potential relative to cyto
case 'Recon3DModel_Dec2017'
    % Temperature in Kelvin
    T = 310.15;
    % Cell compartment identifiers
    compartments = ['c'; 'e'; 'g'; 'l'; 'm'; 'n'; 'r'; 'x'; 'i'];
    % Compartment specific pH
    ph = [7.2; 7.4; 6.35; 5.5; 8; 7.2; 7.2; 7; 7.2];
    % Compartment specific ionic strength in mol/L
    is = 0.15*ones(length(compartments),1);
    % Compartment specific electrical potential relative to cytosol in mV
    chi = [0; 30; 0; 19; -155; 0; 0; -2.303*8.3144621e-3*T*(ph(compartments == 'x')
case 'Recon3DModel_301'
    % Temperature in Kelvin
    T = 310.15;
    % Cell compartment identifiers
    compartments = ['c'; 'e'; 'g'; 'l'; 'm'; 'n'; 'r'; 'x'; 'i'];
    % Compartment specific pH
    ph = [7.2; 7.4; 6.35; 5.5; 8; 7.2; 7.2; 7; 7.2];
    % Compartment specific ionic strength in mol/L
    is = 0.15*ones(length(compartments),1);
    % Compartment specific electrical potential relative to cytosol in mV
    chi = [0; 30; 0; 19; -155; 0; 0; -2.303*8.3144621e-3*T*(ph(compartments == 'x')
otherwise
    error('setup specific parameters for your model')
end

```

## Set the default range of metabolite concentrations

```

switch modelName
case 'Ec_iAF1260_flux1'
    concMinDefault = 1e-5; % Lower bounds on metabolite concentrations in mol/L
    concMaxDefault = 0.02; % Upper bounds on metabolite concentrations in mol/L
    metBoundsFile=[];
case 'iAF1260'
    concMinDefault = 1e-5; % Lower bounds on metabolite concentrations in mol/L
    concMaxDefault = 0.02; % Upper bounds on metabolite concentrations in mol/L
    metBoundsFile=[];
case 'Recon3DModel_Dec2017'
    concMinDefault=1e-5; % Lower bounds on metabolite concentrations in mol/L
    concMaxDefault=1e-2; % Upper bounds on metabolite concentrations in mol/L
    metBoundsFile=which('HumanCofactorConcentrations.txt');%already in the COBRA to
case 'Recon3DModel_301'
    concMinDefault=1e-5; % Lower bounds on metabolite concentrations in mol/L
    concMaxDefault=1e-2; % Upper bounds on metabolite concentrations in mol/L
    metBoundsFile=which('HumanCofactorConcentrations.txt');%already in the COBRA to
otherwise
    error('setup specific parameters for your model')
end

```

## Set the desired confidence level for estimation of thermochemical parameters

The confidence level for estimated standard transformed reaction Gibbs energies is used to quantitatively assign reaction directionality.

```
switch modelName
    case 'Ec_iAF1260_flux1'
        confidenceLevel = 0.95;
        DrGt0_Uncertainty_Cutoff = 20; %KJ/KMol
    case 'iAF1260'
        confidenceLevel = 0.95;
        DrGt0_Uncertainty_Cutoff = 20; %KJ/KMol
    case 'Recon3DModel_Dec2017'
        confidenceLevel = 0.95;
        DrGt0_Uncertainty_Cutoff = 20; %KJ/KMol
    otherwise
        confidenceLevel = -1;%bypass addition of uncertainty temporarily
        %confidenceLevel = 0.95;
        DrGt0_Uncertainty_Cutoff = 20; %KJ/KMol
end
```

## Prepare folder for results

```
if ~exist(resultsPath,'dir')
    mkdir(resultsPath)
end
cd(resultsPath)
```

## Set the print level and decide to record a diary or not (helpful for debugging)

```
printLevel=2;

diary([resultsPath filesep 'diary.txt'])
```

## Setup a thermodynamically constrained model

### Read in the metabolite bounds

```
setDefaultConc=1;
setDefaultFlux=0;
rxnBoundsFile=[];
model=readMetRxnBoundsFiles(model,setDefaultConc,setDefaultFlux,concMinDefault,concMaxI
```

Reading metabolite conc bounds from: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/ther

adp[c]	1e-07	0.0019
adp[m]	0.0026	0.0094
amp[c]	1e-07	0.0012
atp[c]	0.00129	0.0049
atp[m]	0.0028	0.0204
coa[c]	2.92e-05	0.0001168
coa[m]	0.0022	0.0039
nal[c]	1e-07	0.025
nal[e]	0.1326	0.1554
nad[c]	0.00010546	0.0007572
nad[m]	0.0005	0.0075

nadh[c]	9.2574e-07	0.00038294
nadh[m]	1e-07	0.0011
nadp[c]	1e-07	5.8284e-06
nadp[m]	1e-07	0.0015
nadph[c]	1e-07	0.00037523
nadph[m]	1e-07	0.0042
nh4[c]	0.0007	0.0009
pi[c]	0.001	0.0063
ppi[c]	0.0021	0.0076
udp[g]	1.4e-06	0.00014

## Check inputs

```
model = configureSetupThermoModelInputs(model,T,compartments,ph,is,chi,concMinDefault,c
```

```
Field metCompartments is missing from model structure. Attempting to create it.
Attempt to create field metCompartments successful.
```

```
Warning: Setting temperature to a value other than 298.15 K may introduce error, since enthalpies and heat
```

## Add InChI to model

```
%[model, pKaErrorMets] = setupComponentContribution(model,molFileDir);
model = addInchiToModel(model, molFileDir, 'sdf', printLevel);
```

```
Creating MetStructures.sdf from molfiles.
Percentage of metabolites without mol files: 9.1%
Converting SDF to InChI strings.
5835 = number of model metabolites
5835 ... with mol files
0 ... without mol files
4949 ... with nonstandard inchi
886 ... without nonstandard inchi
108 ... composite inchi removed
```

## Add pseudoisomers to model

```
[model, nonphysicalMetBool, pKaErrorMetBool] = addPseudoisomersToModel(model, printLevel)
```

```
Estimating metabolite pKa values.
```

```
ChemAxon's pKa calculator plugin returned an error for 2 metabolites:
```

```
{'CE6252'      }      {'InChI=1/C5H3N4O3/c10-3-1-2(7-4(11)6-1)8-5(12)9-3/h(H3,6,7...'}
{'pchol2ste_hs'}      {'InChI=1/C26H55NO7P/c1-5-6-7-8-9-10-11-12-13-14-15-16-17-1...'}
{'pchol2ste_hs'}
```

```
Assuming that metabolite species in model.metFormulas are representative for metabolites where pKa could not
```

```
5835 = number of model metabolites
217 = number of nonphysical model metabolites
3 = number of model metabolites with pKa error
```

## Check elemental balancing of metabolic reactions

```
ignoreBalancingOfSpecifiedInternalReactions=1;
if ~exist('massImbalance','var')
    if isfield(model,'Srecon')
        model.S=model.Srecon;
    end
    % Check for imbalanced reactions
```

```

fprintf('\nChecking mass and charge balance.\n');
%Heuristically identify exchange reactions and metabolites exclusively involved in
if ~isfield(model,'SIntMetBool') || ~isfield(model,'SIntRxnBool') || ignoreBalancing
    %finds the reactions in the model which export/import from the model
    %boundary i.e. mass unbalanced reactions
    %e.g. Exchange reactions
    %     Demand reactions
    %     Sink reactions
    model = findSExRxnInd(model,[],printLevel);
end

if ignoreBalancingOfSpecifiedInternalReactions
    [nMet,nRxn]=size(model.S);
    ignoreBalancingMetBool=false(nMet,1);
    for m=1:nMet
        %         if strcmp(model.mets{m},'Rtotal3coa[m]')
        %             pause(0.1);
        %         end
        if ~isempty(model.metFormulas{m})
            ignoreBalancingMetBool(m,1)=numAtomsOfElementInFormula(model.metFormulas{m});
        end
    end
    ignoreBalancingRxnBool=getCorrespondingCols(model.S,ignoreBalancingMetBool,model.SIntRxnBool);
    model.SIntRxnBool=model.SIntRxnBool & ~ignoreBalancingRxnBool;
end

printLevelcheckMassChargeBalance=-1; % -1; % print problem reactions to a file
%mass and charge balance can be checked by looking at formulas
[massImbalance,imBalancedMass,imBalancedCharge,imBalancedRxnBool,Elements,missingFormulae]=
    checkMassChargeBalance(model,printLevelcheckMassChargeBalance,resultsBaseFile);
model.balancedRxnBool=~imBalancedRxnBool;
model.balancedMetBool=balancedMetBool;
model.Elements=Elements;
model.missingFormulaeBool=missingFormulaeBool;

%reset original boolean vector
if ignoreBalancingOfSpecifiedInternalReactions
    model.SIntRxnBool=SIntRxnBool;
end
end

```

Checking mass and charge balance.

Assuming biomass reaction is: biomass\_maintenance

ATP demand reaction is not considered an exchange reaction by default. It should be mass balanced:

DM\_atp\_c\_ h2o[c] + atp[c] -> h[c] + adp[c] + pi[c]

There are mass imbalanced reactions, see /home/rfleming/work/sbgCloud/code/fork-COBRA.tutorials/analysis/v

There are mass balanced, but charge imbalanced reactions, see /home/rfleming/work/sbgCloud/code/fork-COBRA

## Create the thermodynamic training model

```
if 0
```



```

%use previously generated training model
aPath = which('driver_createTrainingModel.mlx');
aPath = strrep(aPath,['new' filesep 'driver_createTrainingModel.mlx'],['cache' filesep 'driver_createTrainingModel.mlx']);
load([aPath 'trainingModel.mat'])
else
%recreate the trainingModel
driver_createTrainingModel
end

```

```

removing: /home/rfleming/work/sbgCloud/code/fork-cobratoobox/src/analysis/thermo/componentContribution/old
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoobox/src/analysis/thermo/directionalityReport/old
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoobox/src/analysis/thermo/groupContribution/old
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoobox/src/analysis/thermo/inchi/old
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoobox/src/analysis/thermo/molFiles/old
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoobox/src/analysis/thermo/protons/old
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoobox/src/analysis/thermo/trainingModel/old
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoobox/src/analysis/thermo/componentContribution/new
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoobox/src/analysis/thermo/groupContribution/new
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoobox/src/analysis/thermo/inchi/new
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoobox/src/analysis/thermo/molFiles/new
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoobox/src/analysis/thermo/protons/new
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoobox/src/analysis/thermo/trainingModel/new
ChemAxon Marvin Beans is installed and working.

```

```

linux-vdso.so.1 (0x00007ffe937e7000)
libc.so.6 => /lib/x86_64-linux-gnu/libc.so.6 (0x00007f25ad526000)
libopenbabel.so.5 => /usr/lib/libopenbabel.so.5 (0x00007f25ad2d6000)
libstdc++.so.6 => /usr/lib/x86_64-linux-gnu/libstdc++.so.6 (0x00007f25ad0bc000)
libgcc_s.so.1 => /usr/local/bin/MATLAB/R2021a/sys/os/glnxa64/libgcc_s.so.1 (0x00007f25acea4000)
/lib64/ld-linux-x86-64.so.2 (0x00007f25ad746000)
libdl.so.2 => /lib/x86_64-linux-gnu/libdl.so.2 (0x00007f25ace9c000)
libz.so.1 => /lib/x86_64-linux-gnu/libz.so.1 (0x00007f25ace80000)
libm.so.6 => /lib/x86_64-linux-gnu/libm.so.6 (0x00007f25acd31000)
libgomp.so.1 => /usr/lib/x86_64-linux-gnu/libgomp.so.1 (0x00007f25acce0000)
libpthread.so.0 => /lib/x86_64-linux-gnu/libpthread.so.0 (0x00007f25accc9000)

```

babel must depend on the system libstdc++.so.6 not the one from MATLAB  
Trying to edit the 'LD\_LIBRARY\_PATH' to make sure that it has the correct system path before the Matlab path  
The solution will be arch dependent

```

Successfully added 3914 values from TECRDB
Successfully added 223 formation energies
Successfully added 13 redox potentials
mol2inchi: could not generate inchi for C00080
0 molecules converted
2 audit log messages

```

```

createInChIStruct: no molecule identifier in C00080
mol2inchi: could not generate inchi for C00080
0 molecules converted
2 audit log messages

```

```

mol2inchi: could not generate inchi for C00080
0 molecules converted
2 audit log messages

```

```

mol2inchi: could not generate inchi for C00080
0 molecules converted
2 audit log messages

```

```

mol2inchi: could not generate inchi for C00125
babel: Alias R was not chemically interpreted
createInChIStruct: no molecule identifier in C00125
mol2inchi: could not generate inchi for C00125
babel: Alias R was not chemically interpreted

```





createInChIStruct: no molecule identifier in C02554  
mol2inchi: could not generate inchi for C02554  
babel: Alias R was not chemically interpreted  
mol2inchi: could not generate inchi for C02554  
babel: Alias R was not chemically interpreted  
mol2inchi: could not generate inchi for C02554  
babel: Alias R was not chemically interpreted  
mol2inchi: could not generate inchi for C02780  
0 molecules converted  
2 audit log messages

createInChIStruct: no molecule identifier in C02780  
mol2inchi: could not generate inchi for C02780  
0 molecules converted  
2 audit log messages

mol2inchi: could not generate inchi for C02780  
0 molecules converted  
2 audit log messages

mol2inchi: could not generate inchi for C02780  
0 molecules converted  
2 audit log messages

mol2inchi: could not generate inchi for C02839  
babel: Alias R was not chemically interpreted  
createInChIStruct: no molecule identifier in C02839  
mol2inchi: could not generate inchi for C02839  
babel: Alias R was not chemically interpreted  
mol2inchi: could not generate inchi for C02839  
babel: Alias R was not chemically interpreted  
mol2inchi: could not generate inchi for C02839  
babel: Alias R was not chemically interpreted  
mol2inchi: could not generate inchi for C02988  
babel: Alias R was not chemically interpreted  
createInChIStruct: no molecule identifier in C02988  
mol2inchi: could not generate inchi for C02988  
babel: Alias R was not chemically interpreted  
mol2inchi: could not generate inchi for C02988  
babel: Alias R was not chemically interpreted  
mol2inchi: could not generate inchi for C02988  
babel: Alias R was not chemically interpreted  
mol2inchi: could not generate inchi for C02992  
babel: Alias R was not chemically interpreted  
createInChIStruct: no molecule identifier in C02992  
mol2inchi: could not generate inchi for C02992  
babel: Alias R was not chemically interpreted  
mol2inchi: could not generate inchi for C02992  
babel: Alias R was not chemically interpreted  
mol2inchi: could not generate inchi for C02992  
babel: Alias R was not chemically interpreted  
mol2inchi: could not generate inchi for C03127  
babel: Alias R was not chemically interpreted  
createInChIStruct: no molecule identifier in C03127  
mol2inchi: could not generate inchi for C03127  
babel: Alias R was not chemically interpreted  
mol2inchi: could not generate inchi for C03127  
babel: Alias R was not chemically interpreted  
mol2inchi: could not generate inchi for C03127  
babel: Alias R was not chemically interpreted  
mol2inchi: could not generate inchi for C03127  
babel: Alias R was not chemically interpreted  
mol2inchi: could not generate inchi for C03511  
babel: Alias R was not chemically interpreted  
createInChIStruct: no molecule identifier in C03511  
mol2inchi: could not generate inchi for C03511



```

mol2inchi: could not generate inchi for C06021
babel: Alias R was not chemically interpreted
mol2inchi: could not generate inchi for C06021
babel: Alias R was not chemically interpreted
mol2inchi: could not generate inchi for C06567
babel: Alias R was not chemically interpreted
createInChIstruct: no molecule identifier in C06567
mol2inchi: could not generate inchi for C06567
babel: Alias R was not chemically interpreted
mol2inchi: could not generate inchi for C06567
babel: Alias R was not chemically interpreted
mol2inchi: could not generate inchi for C06567
babel: Alias R was not chemically interpreted
672 = number of model metabolites
657 ... with mol files
15 ... without mol files
627 ... with nonstandard inchi
45 ... without nonstandard inchi
0 ... composite inchi removed

```

Estimating metabolite pKa values for training trainingModel...

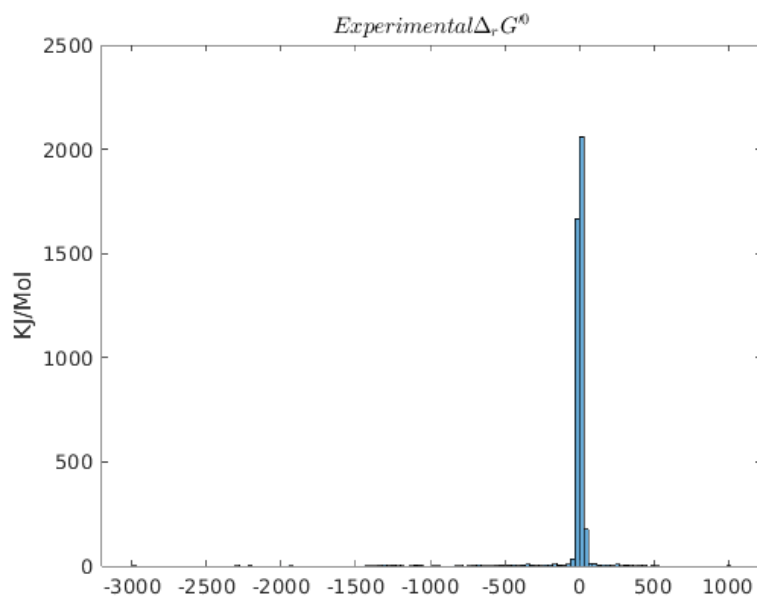
...done.

There are mass imbalanced reactions, see /home/rfleming/work/sbgCloud/code/fork-COBRA.tutorials/analysis/v  
 Performing reverse Legendre transform

```

figure
histogram(trainingModel.DrGt0)
title('$Experimental \smallskip \Delta_{\rm r} G^{\prime 0}$','Interpreter','latex')
ylabel('KJ/Mol')

```



```

fprintf('%u%s\n',nnz(trainingModel.DrGt0==0),' = number of zero DrGt0, i.e. experimental

```

35 = number of zero DrGt0, i.e. experimental apparent equilibrium constant equal to one

```

formulas = printRxnFormula(trainingModel,trainingModel.rxns(trainingModel.DrGt0==0));

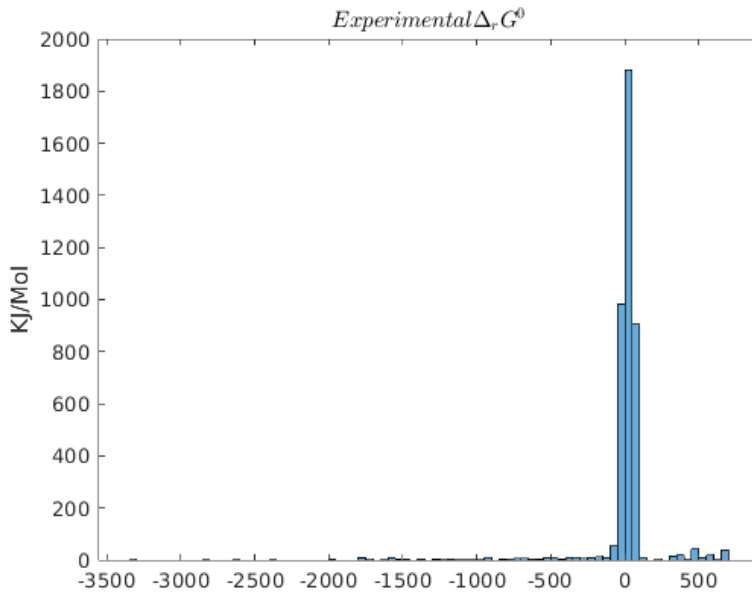
```

TECRDB_79	C01101	<=>	C00231
TECRDB_244	C00041	<=>	C00133
TECRDB_580	C00002 + C00065 + C01650	<=>	C00013 + C00020 + C02553
TECRDB_698	C00003 + C00644	<=>	C00004 + C00085
TECRDB_733	C01101	<=>	C00231
TECRDB_815	C00002 + C00055	<=>	C00008 + C00112
TECRDB_1090	C00001 + C00006 + C00311	<=>	C00005 + C00026 + C00288
TECRDB_1272	C00001 + C06322	<=>	C06749
TECRDB_2030	C00041	<=>	C00133
TECRDB_2202	C00003 + C00579	<=>	C00004 + C00248
TECRDB_2339	C00063 + C00103	<=>	C00013 + C00501
TECRDB_2392	C01213	<=>	C00683
TECRDB_2584	C00041	<=>	C00133
TECRDB_2620	C00026 + C00041	<=>	C00022 + C00025
TECRDB_2621	C00002 + C01107	<=>	C00008 + C01143
TECRDB_2629	C00025	<=>	C00217
TECRDB_2639	C00002 + C00104	<=>	C00008 + C00081
TECRDB_2746	C00031	<=>	C00095
TECRDB_2791	C00010 + C00042 + C00044	<=>	C00009 + C00035 + C00091
TECRDB_2841	2 C00008	<=>	C00002 + C00020
TECRDB_2894	C00041	<=>	C00133
TECRDB_3608	C00031	<=>	C00095
TECRDB_3640	C00047	<=>	C00739
TECRDB_3803	C00636	<=>	C00275
TECRDB_3808	C00075 + C00103	<=>	C00013 + C00029
TECRDB_4052	C00041	<=>	C00133
TECRDB_4271	C00935	<=>	C00190
TECRDB_4375	C00123	<=>	C01570
TECRDB_4377	C00009 + C00299	<=>	C00106 + C00620
TECRDB_4398	C00026 + C00041	<=>	C00022 + C00025
TECRDB_4536	C00031	<=>	C00095
TECRDB_4537	C00031	<=>	C00095
FORM_C00023		<=>	C00023
FORM_C00034		<=>	C00034
FORM_C00080		<=>	C00080

```

figure
histogram(trainingModel.DrG0)
title('$Experimental \medskip \Delta_{r} G^{0}$','Interpreter','latex')
ylabel('KJ/Mol')

```



```
fprintf('%u%s\n',nnz(trainingModel.DrG0==0),' = number of zero DrG0. i.e. equilibrium constant equal to one and same number of hydrogens on both sides')
```

16 = number of zero DrG0. i.e. equilibrium constant equal to one and same number of hydrogens on both sides

```
formulas = printRxnFormula(trainingModel,trainingModel.rxns(trainingModel.DrG0==0));
```

```
TECRDB_79      C01101      <=>      C00231
TECRDB_244     C00041      <=>      C00133
TECRDB_733     C01101      <=>      C00231
TECRDB_1272    C00001 + C06322      <=>      C06749
TECRDB_2030    C00041      <=>      C00133
TECRDB_2392    C01213      <=>      C00683
TECRDB_2584    C00041      <=>      C00133
TECRDB_2629    C00025      <=>      C00217
TECRDB_2894    C00041      <=>      C00133
TECRDB_3640    C00047      <=>      C00739
TECRDB_4052    C00041      <=>      C00133
TECRDB_4271    C00935      <=>      C00190
TECRDB_4375    C00123      <=>      C01570
FORM_C00023     <=>      C00023
FORM_C00034     <=>      C00034
FORM_C00080     <=>      C00080
```

## Create Group Incidence Matrix

Create the group incidence matrix (G) for the combined set of all metabolites.

```
save('data_prior_to_createGroupIncidenceMatrix')
```

```
%param.fragmentationMethod='manual';
param.fragmentationMethod='abinitio';
param.printLevel=0;
param.modelCache=['autoFragment_' modelName];
param.debug=1;
```



```
param.radius=2;
```

```
combinedModel = createGroupIncidenceMatrix(model, trainingModel, param);
```

```
Creating group incidence matrix
```

```
There are 574 fragments unique to the training model.
```

```
There are 914 fragments in common between the training and test models.
```

```
There are 2659 fragments unique to the test model.
```

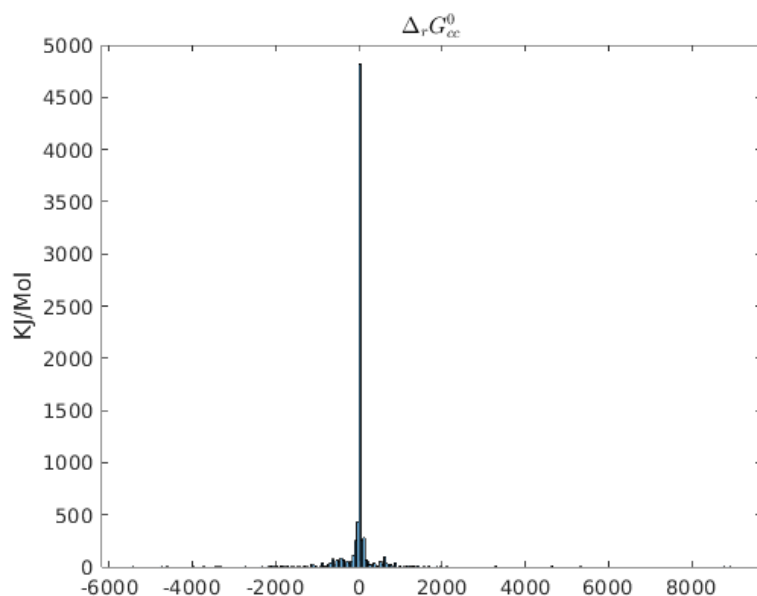
```
save('data_prior_to_componentContribution','model','combinedModel')
```

## Apply component contribution method

```
if ~isfield(model,'DrG0')  
    [model,solution] = componentContribution(model,combinedModel);  
end
```

```
Running Component Contribution method
```

```
figure  
histogram(model.DrG0(~model.unconstrainedDrG0_cc))  
title('$\Delta_r G^0_{cc}$','Interpreter','latex')  
ylabel('KJ/Mol')
```



```
fprintf('%u%s\n',length(model.DrG0),' model reactions')
```

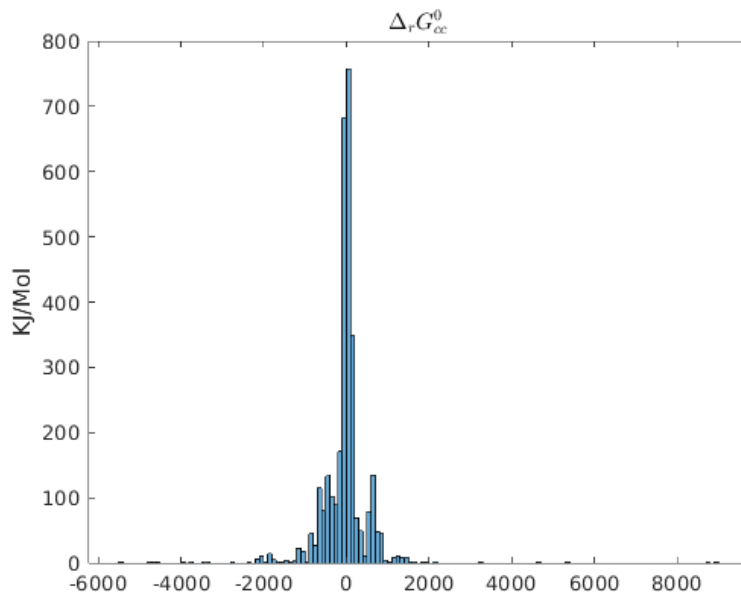
```
10600 model reactions
```

```
fprintf('%u%s\n',nnz(model.unconstrainedDrG0_cc),' of which have partially unconstrained groups in DrG0_cc')
```

```
3147 of which have partially unconstrained groups in DrG0_cc
```

```
figure  
model.transportRxnBool = transportReactionBool(model);
```

```
bool = model.SIntRxnBool & ~model.transportRxnBool & ~model.unconstrainedDrG0_cc;
histogram(model.DrG0(bool))
title('$\Delta_r G^0_{cc}$', 'Interpreter', 'latex')
ylabel('KJ/Mol')
```



```
fprintf('%u%s\n',length(model.DrG0),' model reactions')
```

10600 model reactions

```
fprintf('%u%s\n',nnz(model.unconstrainedDrG0_cc),' of which have partially unconstrained')
```

3147 of which have partially unconstrained groups in DrG0\_cc

```
ind=find(model.unconstrainedDrG0_cc);
formulas = printRxnFormula(model,model.rxns(ind(1:10)));
```

```
2AMACsULT    2amac[c] + nadph[c] + paps[c]    ->    nadp[c] + Lcyst[c] + pap[c]
2DR1PP       h2o[c] + 2drlp[c]    ->    pi[c] + drib[c]
34DHPLACOX   h2o[c] + nad[c] + 34dhpac[c]    ->    2 h[c] + nadh[c] + 34dhpha[c]
34DHPLACOX_NADP_ h2o[c] + nadp[c] + 34dhpac[c]    <=>    2 h[c] + nadph[c] + 34dhpha[c]
34DHXMANDACOX h2o[c] + nad[c] + 34dhmal[d]    ->    2 h[c] + nadh[c] + 34dhoxmand[c]
34DHXMANDACOX_NADP_ h2o[c] + nadp[c] + 34dhmal[d]    <=>    2 h[c] + nadph[c] + 34dhoxmand[c]
3AIBTm       2mop[m] + glu_L[m]    <=>    akg[m] + 3aib[m]
3HAO         o2[c] + 3hanthrn[c]    ->    h[c] + cmusa[c]
3HBcDm       h2o[m] + b2coa[m]    <=>    3hbcoa_R[m]
3HLYTCL      h[c] + 34dhphe[c]    ->    co2[c] + dopa[c]
```

## Setup a thermodynamically constrained model

```
save('debug_prior_to_setupThermoModel')
```

```
if ~isfield(model,'DfGt0')
    model = setupThermoModel(model,confidenceLevel);
end
```

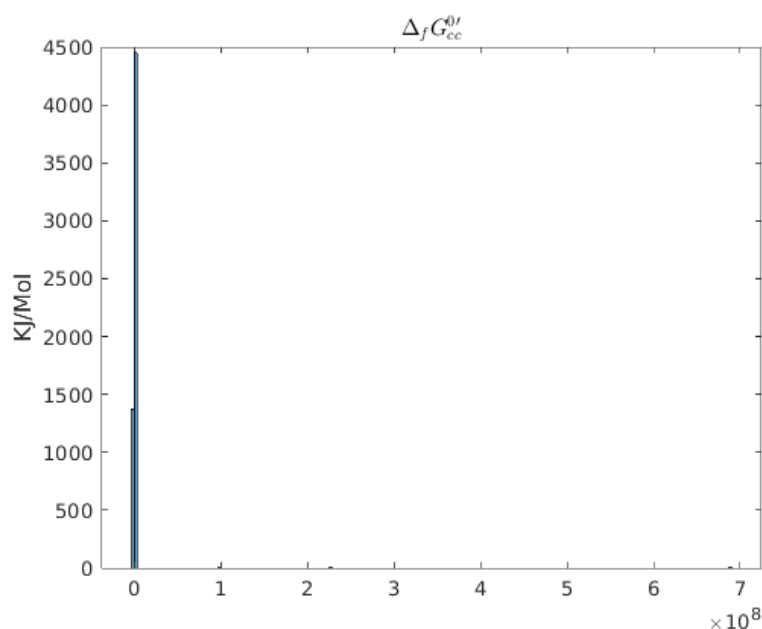
Estimating standard transformed Gibbs energies of formation.

Estimating bounds on transformed Gibbs energies.

Additional effect due to possible change in chemical potential of Hydrogen ions for transport reactions.

Additional effect due to possible change in electrical potential for transport reactions.

```
figure
histogram(model.DfGt0)
title('$\Delta_{f} G^{0\prime}_{cc}$','Interpreter','latex')
ylabel('KJ/Mol')
```



## Generate a model with reactants instead of major microspecies

```
if ~isfield(model,'Srecon')
    printLevel_pHbalanceProtons=-1;
    model=pHbalanceProtons(model,massImbalance,printLevel_pHbalanceProtons,resultsBaseE
end
```

Warning: vonBertalanffy:pHbalanceProtons 'Hydrogen unbalanced reconstruction reactions exist!

## Determine quantitative directionality assignments

```
if ~exist('directions','var') | 1
    fprintf('Quantitatively assigning reaction directionality.\n');
    [model, directions] = thermoConstrainFluxBounds(model,confidenceLevel,DrGt0_Uncerta
end
```

Quantitatively assigning reaction directionality.

9/10600 reactions with DrGtMin=DrGtMax~=0

4/10600 reactions with DrGtMin=DrGtMax=0

The following reactions have DrGtMax=DrGtMin=0:

H2Oter    h2o[c]    <=>    h2o[r]

```

H2Otn    h2o[n]    <=>    h2o[c]
Htr      h[c]      <=>    h[r]
HMR_1095 h[c]      <=>    h[n]
ACYP

```

## Analyse thermodynamically constrained model

Choose the cutoff for probability that reaction is reversible

```
cumNormProbCutoff=0.2;
```

Build Boolean vectors with reaction directionality statistics

```
[model,directions]=directionalityStats(model,directions,cumNormProbCutoff,printLevel);
```

```
9/10600 reactions with DrGtMin=DrGtMax~=0
```

```
4/10600 reactions with DrGtMin=DrGtMax=0
```

```
Qualitative internal reaction directionality:
```

```

8791    internal reconstruction reaction directions.
5208    forward reconstruction assignment.
4        reverse reconstruction assignment.
3579    reversible reconstruction assignment.

```

```
Quantitative internal reaction directionality:
```

```

8791    internal reconstruction reaction directions.
8036    of which have a thermodynamic assignment.
751     of which have no thermodynamic assignment.
1636    forward thermodynamic only assignment.
1512    reverse thermodynamic only assignment.
4888    reversible thermodynamic only assignment.

```

```
Qualitative vs Quantitative:
```

```

2525    Reversible -> Reversible
347     Reversible -> Forward
583     Reversible -> Reverse
120     Reversible -> Uncertain
1286    Forward -> Forward
929     Forward -> Reverse
2362    Forward -> Reversible
631     Forward -> Uncertain
1       Reverse -> Reverse
3       Reverse -> Forward
1       Reverse -> Reversible
0       Reversible -> Uncertain

```

```
Breakdown of relaxation of reaction directionality, Qualitative vs Quantitative:
```

```

2362    qualitatively forward reactions that are quantitatively reversible (total).
1183    of which are quantitatively reversible by range of dGt0.  $P(\Delta_r G^{\prime} < 0) > 0.7$ 
0       of which are quantitatively reversible by range of dGt0.  $0.3 < P(\Delta_r G^{\prime} < 0) < 0.7$ 
1179    of which are quantitatively reversible by range of dGt0.  $P(\Delta_r G^{\prime} < 0) < 0.3$ 
56      of which are quantitatively forward by fixed dGr0t, but reversible by concentration alone (negative fix)
0       of which are quantitatively reverse by dGr0t, but reversible by concentration (negative fix)
0       of which are quantitatively forward by dGr0t, but reversible by concentration (positive fix)
0       of which are quantitatively reverse by dGr0t, but reversible by concentration (uncertain ne)
0       of which are quantitatively forward by dGr0t, but reversible by concentration (uncertain po)

```

```

% directions    a structue of boolean vectors with different directionality
%               assignments where some vectors contain subsets of others
%
% qualtiative -> quantiative changed reaction directions
% .forward2Forward

```

```

% .forward2Reverse
% .forward2Reversible
% .forward2Uncertain
% .reversible2Forward
% .reversible2Reverse
% .reversible2Reversible
% .reversible2Uncertain
% .reverse2Forward
% .reverse2Reverse
% .reverse2Reversible
% .reverse2Uncertain
% .tightened
%
% subsets of qualitatively forward -> quantitatively reversible
% .forward2Reversible_bydGt0
% .forward2Reversible_bydGt0LHS
% .forward2Reversible_bydGt0Mid
% .forward2Reversible_bydGt0RHS
%
% .forward2Reversible_byConc_zero_fixed_DrG0
% .forward2Reversible_byConc_negative_fixed_DrG0
% .forward2Reversible_byConc_positive_fixed_DrG0
% .forward2Reversible_byConc_negative_uncertain_DrG0
% .forward2Reversible_byConc_positive_uncertain_DrG0

```

Write out reports on directionality changes for individual reactions to the results folder.

```
fprintf('%s\n','directionalityChangeReport...');
```

```
directionalityChangeReport...
```

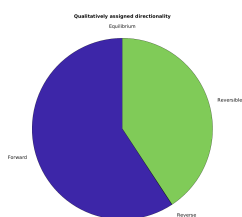
```
directionalityChangeReport(model,directions,cumNormProbCutoff,printLevel,resultsBaseFile
```

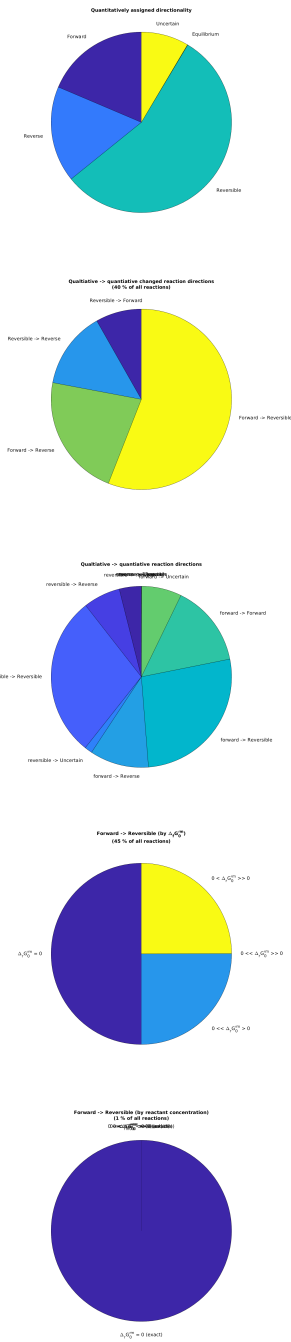
Generate pie charts with proportions of reaction directionalities and changes in directionality

```
fprintf('%s\n','directionalityStatFigures...');
```

```
directionalityStatFigures...
```

```
directionalityStatsFigures(directions,resultsBaseFileName)
```

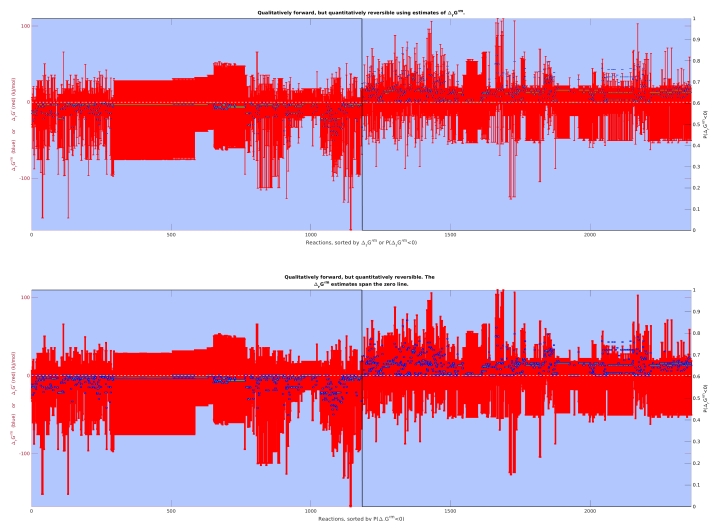




Generate figures to interpret the overall reasons for reaction directionality changes for the qualitatively forward now quantitatively reversible reactions

```
if any(directions.forward2Reversible)
    fprintf('%s\n', 'forwardReversibleFigures...');
    forwardReversibleFigures(model,directions,confidenceLevel)
end
```

forwardReversibleFigures...



Write out tables of experimental and estimated thermochemical parameters for the model

```
generateThermodynamicTables(model,resultsBaseFileName);
save([datestr(now,30) '_' modelName '_thermo'],'model')
save([datestr(now,30) '_vonB_tutorial_complete'])
```

## REFERENCES

- [1] Fleming, R. M. T. & Thiele, I. von Bertalanffy 1.0: a COBRA toolbox extension to thermodynamically constrain metabolic models. *Bioinformatics* 27, 142–143 (2011).
- [2] Haraldsdóttir, H. S., Thiele, I. & Fleming, R. M. T. Quantitative assignment of reaction directionality in a multicompartmental human metabolic reconstruction. *Biophysical Journal* 102, 1703–1711 (2012).
- [3] Noor, E., Haraldsdóttir, H. S., Milo, R. & Fleming, R. M. T. Consistent Estimation of Gibbs Energy Using Component Contributions. *PLoS Comput Biol* 9, e1003098 (2013).
- [4] Fleming, R. M. T. , Predicat, G., Haraldsdóttir, H. S., Thiele, I. von Bertalanffy 2.0 (in preparation).