

# Stage-1 Architecture Specification

Dual-Platform mRNA Therapy for X-Linked Adrenoleukodystrophy (ALD)  
(LNP + EV; Peripheral-First → CNS)

Method2Model

## Abstract

This document specifies the Stage-1 architecture for a mechanistic, scenario-capable model of a dual-platform mRNA therapy for X-linked adrenoleukodystrophy (ALD). The model integrates lipid nanoparticles (LNP) and extracellular vesicles (EV) as delivery platforms in a peripheral-first → CNS strategy. The architecture is organized into seven interacting blocks: (A) Delivery & Exposure (PBPK/Transport), (B) Expression & Targeting (mRNA → ALDP@Peroxisome), (C) Disease / Pharmacodynamics (ALDP → VLCFA → Biomarkers → Function), (D) Safety / Immunogenicity, (E) CMC / Batch Variability, (F) Uncertainty & Scenario Engine, and (G) Defensibility & Decision Logic.

The goal of this specification is to provide a complete, explicit, and implementation-ready description of the model structure at the level of states, observables, input parameters, and inter-block interfaces, without relying on simplifying assumptions such as single-compartment PK or purely linear dose-response.

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# 1 Scope and Design Intent

The architecture captures the mechanistic chain

Carrier/Route/Dosing  $\longrightarrow$  Tissue & Cellular Exposure  $\longrightarrow$  mRNA Expression  
 $\longrightarrow$  ALDP at Peroxisomal Membranes  $\longrightarrow$  VLCFA Reduction  
 $\longrightarrow$  Biomarkers & Function  $\longrightarrow$  Safety, CMC, and Decisions.

The model must support:

- Multi-route, multi-platform delivery: IV-LNP (peripheral), EV-IN, and route combinations with focused ultrasound (FUS) to modulate blood–brain barrier (BBB) permeability.
- Explicit modeling of ALDP expression and peroxisomal targeting (vs. mislocalization) in relevant cell types.
- Coupling between ALDP levels, peroxisomal  $\beta$ -oxidation capacity, and very-long-chain fatty acid (VLCFA) dynamics in plasma and CNS.
- Safety endpoints including innate cytokine responses, complement activation, hepatic toxicity (ALT/AST), and anti-carrier immunity.
- CMC-driven variability in LNP/EV potency, quality attributes, and stability.
- Uncertainty-driven scenario analysis to identify first-break mechanisms and regimen robustness.
- Generation of defensible Go/No-Go and Pivot decisions aligned with the pre-specified aims of the programme.

## 2 Indices, Phenotypes, and State Classes

### 2.1 Indices

We define the following indices:

- Species:  $s \in \{\text{mouse, human-projected}\}$ .
- Phenotype / clinical window:  $p \in \{\text{AMN, very-early cALD, early cALD}\}$ , where AMN denotes adrenomyeloneuropathy and cALD denotes cerebral ALD.
- Carrier type:  $k \in \{\text{LNP-std, LNP-brain, EV}\}$ .
- Administration route:  $r \in \{\text{IV, IN, IV+FUS, EV+FUS}\}$ .
- Compartments:

$c \in \{\text{plasma, liver, spleen, adrenal, other periphery,}$   
 $\text{CNS-blood side, CSF, CNS-parenchyma}\}.$

- CNS cell types:  $\text{cell} \in \{\text{endothelium, astrocyte, neuron, oligodendrocyte, microglia}\}.$

## 2.2 Classes of States

For each block we distinguish three categories of variables:

- **Simulated states** ( $S_{\text{sim}}$ ): latent or internal states that the model evolves over time, but that may or may not be directly measurable (e.g. per-cell mRNA counts,  $\beta$ -oxidation capacity, anti-PEG titer).
- **Observable variables** ( $S_{\text{obs}}$ ): variables that correspond to measurable quantities in experiments or clinical data (e.g. plasma VLCFA, DTI metrics, NfL, Loes score, ALT).
- **Policy/decision variables** ( $S_{\text{pol}}$ ): derived variables that encode decisions, flags, or classifications (e.g. Go/No-Go labels, first-break mechanism, safety violations).

This classification is explicit in the block-by-block specification below and is important for implementation, calibration, and reporting.

## 3 Block A: Delivery and Exposure (PBPK / Transport)

### 3.1 States and Observables

For each carrier  $k$ , route  $r$ , compartment  $c$ , and cell type:

- $S_{\text{sim}}$ :
  - $C_{k,c}(t)$ : carrier concentration in compartment  $c$  at time  $t$ ,
  - $U_{k,c,\text{cell}}(t)$ : cumulative uptake of carrier  $k$  into a given cell type in  $c$ ,
  - $P_{\text{BBB}}(t)$ : effective BBB permeability at time  $t$ .
- $S_{\text{obs}}$  (if measured):
  - Plasma and tissue PK curves (e.g. LC–MS/MS, fluorescence).
  - CNS tracer uptake (e.g. radiotracers, imaging).
- $S_{\text{pol}}$ :
  - Qualitative classification of CNS exposure: “adequate” / “borderline” / “inadequate” for a given scenario.

### 3.2 Inputs

- Dose schedule:  $D_{k,r}(t_j)$  (e.g. mg mRNA/kg or particles/kg at administration times  $t_j$ ).
- FUS timing and parameters (if applicable).
- PBPK parameters: compartment volumes, blood flows, clearances  $CL_c$ , and baseline BBB permeability  $P_{\text{baseline}}$ , each with uncertainty ranges.
- Batch potency multipliers from Block E.

### 3.3 Dynamics

For each compartment  $c$ , the carrier concentration satisfies:

$$\frac{dC_{k,c}(t)}{dt} = \sum_{c'} Q_{c' \rightarrow c}(C_{k,c'}(t)) - \sum_{c''} Q_{c \rightarrow c''}(C_{k,c}(t)) - CL_c(C_{k,c}(t)) - \text{Uptake}_c(C_{k,c}(t)), \quad (1)$$

where  $Q_{c' \rightarrow c}$  are inter-compartment flows and  $\text{Uptake}_c$  represents loss into cells.

Transport across the BBB is modeled as

$$Q_{\text{plasma} \rightarrow \text{CNS}}(t) = P_{\text{BBB}}(t) A_{\text{BBB}} (C_{\text{plasma}}(t) - C_{\text{CNS}}(t)), \quad (2)$$

with

$$P_{\text{BBB}}(t) = P_{\text{baseline}} \times \begin{cases} M_{\text{FUS}}(\omega), & t \in [t_{\text{FUS}}, t_{\text{FUS}} + \Delta t], \\ 1, & \text{otherwise,} \end{cases} \quad (3)$$

where  $M_{\text{FUS}}(\omega)$  is a random multiplicative factor (e.g. in the range 5–20) capturing the magnitude of FUS-induced BBB opening, and  $\Delta t$  is the open-window duration.

Cellular uptake is described generically by

$$\frac{dU_{k,c,\text{cell}}(t)}{dt} = f_{k,r,c,\text{cell}}(C_{k,c}(t), \theta_{\text{surface}}, \text{ligands}), \quad (4)$$

where  $f_{k,r,c,\text{cell}}$  encodes route-specific and cell-type specific uptake efficiency, including effects of surface receptors and ligand decoration.

### 3.4 Outputs and Interfaces

Block A provides:

- Trajectories  $C_{k,c}(t)$  and  $U_{k,c,\text{cell}}(t)$  as inputs to Block B.
- Classification of CNS exposure quality as part of  $S_{\text{pol}}$  feeding into Block G (defensibility) and Block F (scenario outcomes).

## 4 Block B: Expression and Peroxisomal Targeting (mRNA $\rightarrow$ ALDP@Peroxisome)

### 4.1 States and Observables

For each relevant cell type (e.g. hepatocytes, oligodendrocytes):

- $S_{\text{sim}}$ :

$$\begin{aligned} m(t) &: \text{intracellular mRNA copy number for ABCD1,} \\ P_{\text{ALDP,tot}}(t) &: \text{total ALDP protein,} \\ P_{\text{ALDP,peri}}(t) &: \text{ALDP localized at peroxisomal membranes,} \\ P_{\text{ALDP,mis}}(t) &: \text{mislocalized ALDP (e.g. ER, aggregates),} \\ \theta_{\text{target}}(t) &= \frac{P_{\text{ALDP,peri}}(t)}{P_{\text{ALDP,tot}}(t)} : \text{targeting efficiency.} \end{aligned}$$

- $S_{\text{obs}}$ :

- Immunofluorescence (IF): colocalization of ALDP with peroxisomal markers (e.g. PEX14, PMP70).

- Protease-protection assays on peroxisomal fractions.
- PTS1/PTS2 import reporter readouts.
- Western blots: peroxisomal vs total ALDP.
- $S_{\text{pol}}$ :
  - Regime label: “sub-therapeutic”, “within therapeutic window”, “overexpression-risk”.

## 4.2 Inputs

- Uptake trajectories  $U_{k,c,\text{cell}}(t)$  from Block A (converted to intracellular mRNA input rate).
- mRNA features: cap structure, nucleoside modifications, UTRs, codon optimization, which influence translation rate  $k_{\text{tr}}$  and innate sensing.
- Peroxisomal biogenesis capacity: PEX19/PEX3 levels and saturation characteristics.

## 4.3 Dynamics

**mRNA kinetics.**

$$\frac{dm(t)}{dt} = k_{\text{in}}(t) - k_{\text{deg,m}} m(t), \quad (5)$$

where  $k_{\text{in}}(t)$  is determined by cellular uptake and endosomal escape, and  $k_{\text{deg,m}}$  is the effective degradation rate of mRNA.

**Translation.**

$$\frac{dP_{\text{ALDP,tot}}(t)}{dt} = k_{\text{tr}} m(t) - k_{\text{deg,p}} P_{\text{ALDP,tot}}(t), \quad (6)$$

where  $k_{\text{tr}}$  is the translation rate and  $k_{\text{deg,p}}$  is the protein degradation rate.

**Peroxisomal targeting vs mislocalization.**

$$\frac{dP_{\text{ALDP,peri}}(t)}{dt} = k_{\text{ins}}(\text{PEX19/PEX3}, P_{\text{ALDP,tot}}(t), \text{crowding}) - k_{\text{turnover,peri}} P_{\text{ALDP,peri}}(t), \quad (7)$$

with

$$P_{\text{ALDP,mis}}(t) = P_{\text{ALDP,tot}}(t) - P_{\text{ALDP,peri}}(t). \quad (8)$$

The insertion rate  $k_{\text{ins}}$  is modeled as a saturating function of total ALDP and the available PEX19/PEX3 capacity; at high overexpression levels, saturation leads to an increased fraction of  $P_{\text{ALDP,mis}}$  and potential peroxisome/ER stress.

## 4.4 Therapeutic Window and Stress Zone

Let  $P_{\text{ALDP,WT}}$  denote the peroxisomal ALDP level in a wild-type cell.

- **Therapeutic region:**  $P_{\text{ALDP,peri}} \approx (1-3.5) \times P_{\text{ALDP,WT}}$ , sufficient to restore a large fraction of peroxisomal  $\beta$ -oxidation capacity and to drive  $\geq 50\%$  reduction in VLCFA in vitro.
- **Sub-therapeutic region:**  $P_{\text{ALDP,peri}} < 0.5 \times P_{\text{ALDP,WT}}$ .
- **Overexpression-risk region:**  $P_{\text{ALDP,tot}} > 3-4 \times P_{\text{ALDP,WT}}$ , associated with increased risk of mislocalization and peroxisomal/ER stress.

These regions are encoded as sets in the  $(P_{\text{ALDP,peri}}, P_{\text{ALDP,mis}})$  plane rather than as single thresholds.

## 4.5 Targeting Score

We define a composite targeting score:

$$S_{\text{target}} = f_{\text{IF}} \cdot f_{\text{PTS}} \cdot f_{\text{protease}}, \quad (9)$$

where each factor is normalized to  $[0, 1]$  based on:

- $f_{\text{IF}}$ : quality and extent of ALDP colocalization with peroxisomal markers in IF.
- $f_{\text{PTS}}$ : efficiency of PTS1/PTS2 cargo import.
- $f_{\text{protease}}$ : degree of protection of ALDP in protease-protection assays of peroxisomal fractions.

## 4.6 Outputs and Interfaces

Block B outputs:

- Trajectories  $P_{\text{ALDP,peri}}(t)$ ,  $\theta_{\text{target}}(t)$ , and  $S_{\text{target}}(t)$ .
- State classifications (therapeutic vs sub-therapeutic vs overexpression-risk).
- An effective peroxisomal capacity signal for Block C (via  $\beta$ -oxidation capacity  $V_{\beta}(t)$ ).

## 5 Block C: Disease / Pharmacodynamics (ALDP $\rightarrow$ VLCFA $\rightarrow$ Biomarkers $\rightarrow$ Function)

Block C has two branches: a mouse branch  $C_{\text{mouse}}$  and a human-projected branch  $C_{\text{human}}$ . The former is used for design and interpretation of preclinical studies; the latter is used for translational reasoning and risk mapping in humans.

### 5.1 Mouse Branch $C_{\text{mouse}}$

#### 5.1.1 States and Observables

- $S_{\text{sim}}$ :

$V_{\beta}(t)$ : peroxisomal  $\beta$ -oxidation capacity,  
 $C_{26}^{\text{plasma}}(t)$ ,  $C_{26}^{\text{CNS}}(t)$ : VLCFA concentrations,  
 $\text{Inflamm}_{\text{CNS}}(t)$ : CNS inflammatory tone,  
 $\text{AxonInjury}(t)$ : latent axonal injury index.

- $S_{\text{obs}}$ :
  - Plasma and CNS VLCFA levels (e.g. C26:0 and C26:0/C22:0).
  - DTI metrics (FA, MD, RD) in relevant brain regions.
  - Rotarod performance.
  - Histological measures (e.g. demyelination, microgliosis).
- $S_{\text{pol}}$ :
  - Aim-2 success/failure classification.
  - Progression vs stabilization labels based on combined VLCFA and functional readouts.



### 5.1.2 Inputs

- Peroxisomal ALDP signal from Block B:  $P_{\text{ALDP,peri}}(t)$ .
- Phenotype  $p$  (e.g. AMN-like vs early cALD-like), which influences VLCFA production and baseline inflammation.

### 5.1.3 Dynamics

**ALDP to  $\beta$ -oxidation capacity.**

$$V_{\beta}(t) = V_{\beta}^{\text{WT}} \cdot g\left(\frac{P_{\text{ALDP,peri}}(t)}{P_{\text{ALDP,WT}}}\right), \quad (10)$$

where  $g$  is a saturating or threshold-like function with steep response at low ALDP levels (partial rescue can yield substantial functional restoration) and a plateau near wild-type.

**VLCFA dynamics.** For each relevant compartment (plasma or CNS), we consider:

$$\frac{dC_{26}(t)}{dt} = P_{\text{prod}}(p, s, t) - V_{\beta}(t) h(C_{26}(t)), \quad (11)$$

where  $P_{\text{prod}}(p, s, t)$  is the VLCFA production rate (phenotype- and age-dependent) and  $h$  is a substrate-dependent function (e.g. of Michaelis–Menten type).

**Axonal injury and imaging/behavior.** A latent axonal injury index evolves according to

$$\frac{d\text{AxonInjury}(t)}{dt} = F_{\text{axon}}\left(C_{26}^{\text{CNS}}(t), \text{Inflamm}_{\text{CNS}}(t)\right), \quad (12)$$

and DTI metrics and Rotarod performance are modeled as functions of  $\text{AxonInjury}(t)$  plus measurement noise and sensitivity characteristics specific to each assay.

### 5.1.4 Endpoints and Aim-2 Criteria

Example Aim-2 criteria (preclinical):

- Plasma C26:0 reduction  $\geq 35\%$  at week 4 versus untreated control.
- CNS C26:0 reduction  $\geq 25\%$  at week 4.
- Functional improvement:
  - Rotarod performance improvement  $\geq 20\%$ , and/or
  - DTI fractional anisotropy (FA) increase  $\geq 10\%$  (or RD decrease) in predefined regions.

Block  $C_{\text{mouse}}$  aggregates these into  $S_{\text{pol}}$  (Aim-2 Go / No-Go / borderline).

## 5.2 Human-Projected Branch $C_{\text{human}}$

### 5.2.1 States and Observables

- $S_{\text{sim}}$ :

$V_{\beta}^{\text{human}}(t)$ : projected human peroxisomal capacity,  
 $C_{26,\text{human}}^{\text{plasma}}(t), C_{26,\text{human}}^{\text{CNS}}(t)$ : human VLCFA states,  
 $\text{AxonInjury}_{\text{human}}(t)$ : latent human axonal injury,  
 $\text{MicroglialAct}(t)$ : microglial activation level.

- $S_{\text{obs}}$ :
  - Plasma VLCFA (e.g. C26:0, ratios).
  - Neurofilament light chain (NfL).
  - **Loes score** (MRI-based severity index; *only present in this human-projected branch, not in the mouse branch*).
  - Clinical functional scales (e.g. neurologic or disability scores).
- $S_{\text{pol}}$ :
  - Projected stabilization vs progression vs improvement with respect to natural history or standard-of-care.

### 5.2.2 Dynamics

The structure mirrors the mouse branch but with human-specific time scales and scaling relations. For example:

$$\frac{dC_{26,\text{human}}^{\text{CNS}}(t)}{dt} = P_{\text{prod}}^{\text{human}}(p, t) - V_{\beta}^{\text{human}}(t) h_{\text{human}}(C_{26,\text{human}}^{\text{CNS}}(t)), \quad (13)$$

$$\frac{d\text{NfL}(t)}{dt} = G_{\text{NfL}}(C_{26,\text{human}}^{\text{CNS}}(t), \text{AxonInjury}_{\text{human}}(t)), \quad (14)$$

$$\frac{d\text{Loes}(t)}{dt} = G_{\text{Loes}}(C_{26,\text{human}}^{\text{CNS}}(t), \text{MicroglialAct}(t)). \quad (15)$$

The functions  $G_{\text{NfL}}$  and  $G_{\text{Loes}}$  encode relationships between VLCFA burden, inflammatory/axonal injury, and biomarkers. They are parameterized using human data from natural history and relevant interventions (e.g. HSCT, gene therapy), subject to uncertainty quantification in Block F.

## 5.3 Outputs and Interfaces

Block C exports:

- Simulated trajectories of VLCFA, NfL, DTI/Rotarod (mouse), and NfL/Loes (human-projected).
- Aim-1 and Aim-2 success probabilities (preclinical) and qualitative translational implications (stabilization vs progression).
- Inputs to Block G for defensibility and decision-making.

## 6 Block D: Safety and Immunogenicity

### 6.1 States and Observables

- $S_{\text{sim}}$ :
  - Cytokine<sub>*i*</sub>(*t*): innate cytokines (e.g. IL-6, TNF- $\alpha$ ),
  - sC5b9(*t*): complement activation,
  - ALT(*t*), AST(*t*): liver enzyme states,
  - Ab<sub>PEG</sub>(*t*): anti-PEG (or anti-lipid) antibody titer.
- $S_{\text{obs}}$ :
  - Measured cytokine profiles, complement markers.

- ALT/AST laboratory values.
- CRS grades, clinical signs.
- Imaging evidence of FUS-related events (e.g. microbleeds).
- $S_{\text{pol}}$ :
  - Safety violation flags: liver, CRS, FUS-related, etc.

## 6.2 Inputs

- Dose history for each carrier/route.
- mRNA and carrier features (e.g. dsRNA content, endotoxin, nucleoside modifications, lipid composition).
- Individual or strain-level baseline immune sensitivity.

## 6.3 Dynamics

**Innate cytokine and complement response.**

$$\text{Cytokine}_{i,\text{max}} = f_{\text{innate},i}(D_{\text{effective}}, \text{dsRNA}, \text{modifications}, s), \quad (16)$$

$$\text{sC5b9}_{\text{max}} = f_{\text{comp}}(D_{\text{effective}}, \text{lipid composition}, s), \quad (17)$$

where  $f_{\text{innate},i}$  and  $f_{\text{comp}}$  are carrier- and species-specific response functions, modulated by nucleoside chemistry and formulation.

**Liver enzymes.**

$$\frac{d\text{ALT}(t)}{dt} = f_{\text{hepatotox}}(C_{\text{LNP,liver}}(t), \text{repeat-dose history}), \quad (18)$$

with a similar expression for AST.

**Anti-PEG and effect on PK.**

$$\frac{dAb_{\text{PEG}}(t)}{dt} = f_{\text{B-cell}}(D_{\text{LNP}}(t)) - k_{\text{decay}} Ab_{\text{PEG}}(t). \quad (19)$$

The level  $Ab_{\text{PEG}}(t)$  feeds back into Block A by effectively increasing clearance of LNPs and potentially modifying uptake patterns.

## 6.4 Policy and Safety Rules

Typical policy rules encoded in  $S_{\text{pol}}$  include:

- If ALT or AST exceeds  $3\times$  the upper limit of normal (ULN), then raise a “liver safety violation” flag.
- If CRS grade  $\geq 2$ , then raise a “CRS violation” flag.
- If FUS-induced imaging abnormalities (e.g. microbleeds, edema) are detected, then raise a “FUS safety violation” flag and recommend stopping or de-escalating FUS in that arm.

## 7 Block E: CMC and Batch Variability

### 7.1 States and Observables

For each batch of LNP:

- $S_{\text{sim}}$ :

Size, PDI, EE%, RNAInt, Pot<sub>in vitro</sub>, Endotoxin, pH, Osm

Pot( $t$ ): effective potency over time,

where Pot( $t$ ) encodes the fraction of nominal dose that remains functionally active given storage time and handling.

- $S_{\text{obs}}$ :

- Measured size/PDI/EE% (e.g. DLS, HPLC).
- RNA integrity (e.g. RIN-like indices).
- In vitro potency assays (e.g. reporter expression, VLCFA rescue in cells).

- $S_{\text{pol}}$ :

- Batch classification: “in-spec”, “borderline”, or “out-of-spec”.

For EV-based products, analogous variables include particle count, size distribution, mRNA copies per  $10^{10}$  EV, translation competency, and relevant markers.

### 7.2 Role in the Model

**Hard constraints.** If critical quality attributes (CQA) fall outside pre-defined specification limits (e.g. PDI > 0.2, excessive endotoxin), the batch is considered unusable.

**Potency as a random variable.** For in-spec batches, we model a batch-specific potency multiplier Pot <sub>$k$ ,batch</sub> as a random variable with a distribution informed by empirical data. The effective dose entering Block A and Block B is:

$$D_{\text{effective}} = D_{\text{nominal}} \cdot \text{Pot}_{k,\text{batch}}. \quad (20)$$

**Stability and freeze-thaw.** Time- and handling-dependent decay of potency is captured by:

$$\text{Pot}(t) = \text{Pot}_0 \cdot e^{-k_{\text{decay}} t} \cdot f_{\text{FT}}(\#\text{freeze-thaw}), \quad (21)$$

where  $k_{\text{decay}}$  is a storage-dependent decay constant and  $f_{\text{FT}}$  accounts for additional potency loss per freeze-thaw cycle.

### 7.3 Outputs and Interfaces

Block E provides:

- Effective potency multipliers and quality states per batch to Block A and Block B.
- Batch classification for Block G (defensibility) and Block F (scenario generation).

## 8 Block F: Uncertainty and Scenario Engine

### 8.1 Uncertain Parameters

Block F treats the following as random parameters with prior distributions:

- Block A: CNS uptake fractions per route (IV-LNP, EV-IN, EV+FUS), blood flows, clearances  $CL_c$ , FUS multiplier  $M_{FUS}$ .
- Block B: Translation rates  $k_{tr}$ , insertion rates  $k_{ins}$ , mRNA and protein half-lives ( $k_{deg,m}$ ,  $k_{deg,p}$ ), PEX19/PEX3 capacity.
- Block C: VLCFA production rates  $P_{prod}$ , functional forms and parameters of  $g$  and  $h$ , sensitivities mapping VLCFA/injury to NfL/DTI/Loes.
- Block D: Innate response sensitivities, dose–response of cytokines and complement, hepatotoxicity response, anti-PEG kinetics.
- Block E: Batch potency distributions, decay  $k_{decay}$ , and freeze–thaw sensitivity.

### 8.2 Scenario Dimensions

Scenarios combine:

- Phenotype: AMN vs very-early cALD vs early cALD.
- Route/platform: IV-LNP (peripheral), EV-IN, IV+FUS, EV+FUS, and combinations.
- Dosing regimens: weekly vs every-two-weeks (Q2W), duration 4–8 weeks, dose levels.
- Batch qualities: high-, medium-, low-potency batches consistent with Block E.
- Immune backgrounds: innate “sensitive” vs “non-sensitive”, prior exposure effects.

### 8.3 Outputs

Using Monte Carlo or Latin hypercube sampling across the parameter and scenario space, Block F yields:

- **First-break maps:** for each scenario, the distribution of which constraint or mechanism fails first (e.g. CNS exposure, peroxisomal targeting, PD effect, safety, CMC).
- **Robustness curves:** probability of satisfying Aim-1 and Aim-2 criteria as a function of uncertainty and regimen design.
- **Generalization matrices:** cross-scenario comparisons such as AMN vs cALD, LNP vs EV, and route-level performance.

These outputs feed directly into Block G for decision-making and into Method2Model Use Cases 1, 4, 5, and 6.

## 9 Block G: Defensibility and Decision Logic

### 9.1 Chain-of-Evidence Construction

Block G collects and organizes outputs from Blocks A–F into a defensible chain of evidence:

1. **Delivery & exposure:** CNS exposure trajectories, tissue distribution, and cell-type uptake for each route and platform.

2. **Targeting:** ALDP peroxisomal levels  $P_{\text{ALDP,peri}}(t)$ , targeting efficiency  $\theta_{\text{target}}(t)$ , and composite score  $S_{\text{target}}$ .
3. **Biochemistry:** VLCFA trajectories in plasma and CNS in mouse and human-projected branches.
4. **Function:**
  - Mouse: Rotarod performance, DTI metrics, histology.
  - Human-projected: NfL and *Loes score* (the latter present only in the human-projected branch).
5. **Safety:** Risks and violation probabilities for ALT/AST, CRS, and FUS safety.
6. **CMC:** Batch quality and stability, in-spec vs borderline vs out-of-spec status, and comparability to reference lots.

## 9.2 Decision Surfaces

Based on the chain-of-evidence and Block F outputs, Block G defines multi-dimensional decision surfaces:

- **Go:** high probability of meeting Aim-1 and Aim-2 thresholds with acceptable safety and CMC performance; first-break mechanisms manageable via tuning.
- **No-Go:** structural failures, such as:
  - CNS delivery remains inadequate even under optimistic parameter combinations, or
  - safety/CMC constraints are violated in ways that are not addressable by realistic adjustments.
- **Pivot:** intermediate cases where:
  - platform mix should shift (e.g. more EV, less LNP),
  - route should change (e.g. from IN to FUS),
  - phenotype priority should be adjusted (e.g. focus on AMN if CNS effect is insufficient for cALD),
  - endpoint strategy should refocus on more sensitive or robust biomarkers.

Block G also formalizes endpoint hierarchy (primary/secondary/exploratory) consistent with regulatory and reviewer expectations, and prepares standard outputs for IRB, grant, and sponsor review, including robustness plots, first-break maps, and generalization matrices.

## 10 Alignment with Method2Model Stage-1 Use Cases

The architecture directly supports Method2Model Stage-1 Use Cases:

- *Use Case 1 – Scan for protocol blind spots:* via Block F first-break maps and cross-block constraints (A–E).
- *Use Case 2 – Validate power against real-world noise:* by providing variance and uncertainty structures for key endpoints in Block C (mouse and human-projected branches).
- *Use Case 3 – Stop paying for low-information data:* by evaluating, via Block F, which endpoints actually change Go/No-Go decisions.

- *Use Case 4 – Stress-test regimen robustness:* by exploring dosing and scheduling scenarios in Block A and Block F.
- *Use Case 5 – Prevent “works here, fails there”:* through the mouse vs human-projected dual branch in Block C and scenario generalization in Block F.
- *Use Case 6 – Simulate bottlenecks and drop-offs:* via CMC-driven constraints in Block E and their impact on exposure and efficacy.
- *Use Case 7 – Before you submit:* via Block G, which assembles the chain-of-evidence and decision surfaces into a reviewer-ready defensibility pack.

## 11 Conclusion

This Stage-1 architecture (Version 2) defines a mechanistic, multi-block model that:

1. Represents delivery, expression, disease dynamics, safety, and CMC as coupled but distinct subsystems.
2. Distinguishes simulated states, observable variables, and policy/decision variables for clarity and implementability.
3. Explicitly separates mouse and human-projected branches in Block C, with the Loes score confined to the human-projected branch.
4. Supports uncertainty-aware scenario analysis and structured Go/No-Go/Pivot decisions.

This specification is intended to be implementation-ready: in the next stage, each block can be mapped to software modules (e.g. Python classes) with parameter dictionaries and solver interfaces, while preserving the structure and semantics defined herein.