



# International Journal of Contemporary Research In Multidisciplinary

## Review Article

# A Systematic Review on Lifestyle and Nutritional Management of Polycystic Ovary Syndrome

Dr. Anamika Dixit <sup>1\*</sup>, Durgesh Kumar Sharma <sup>2</sup>

<sup>1</sup> Assistant Professor, Department of Human Nutrition, University Institute of Health Science  
C.S.J.M University, Kanpur, Uttar Pradesh, India

<sup>2</sup> Student, Department of Human Nutrition, University Institute of Health Science,  
C.S.J.M University, Kanpur, Uttar Pradesh, India

**Corresponding Author:** \* Dr. Anamika Dixit

**DOI:** <https://doi.org/10.5281/zenodo.17678653>

Abstract	Manuscript Information
Here, we present a narrative review of the widely understood changes to the nutrition and lifestyles of women and girls with polycystic ovary syndrome (PCOS). The database was analysed, combining PCOS entries with causes, diseases, diet supplementation, lifestyle, physical activity, and use of herbs. This study explains how different biochemical routes contribute to imbalances in lipid, carbohydrate, and hormone regulation among affected individuals. It also explores links with sleep problems, physiological and psychological shifts, and stress-related inflammation. These conditions consistently lead to the occurrence of severe diseases in patients suffering from diabetes, the fatty degeneration of internal organs, infertility, atherosclerosis, cardiovascular diseases and cancer. Change in lifestyles, diet patterns and proper selection of nutrients, pharmacological and natural supplementation in the form of herbs, and physical activity have been proposed. The progress and consequences of PCOS are largely modifiable and depend on the patient's effort, although we have to take into account the genetic determinants.	<ul style="list-style-type: none"> <li>▪ ISSN No: 2583-7397</li> <li>▪ Received: 05-11-2024</li> <li>▪ Accepted: 11-12-2024</li> <li>▪ Published: 30-12-2024</li> <li>▪ IJCRM:3(6); 2024: 240-253</li> <li>▪ ©2024, All Rights Reserved</li> <li>▪ Plagiarism Checked: Yes</li> <li>▪ Peer Review Process: Yes</li> </ul>
	How to Cite this Manuscript
	Dixit A, Sharma DK. A Systematic Review on Lifestyle and Nutritional Management of Polycystic Ovary Syndrome. International Journal of Contemporary Research in Multidisciplinary.2024; 3(6): 240-253.

**KEYWORDS:** Nutrition, lifestyle, PCOS; reproduction; diet; sleep; supplementation; herbs supporting

## INTRODUCTION

Polycystic ovary syndrome (PCOS) is a prevalent hormonal disorder that affects a significant portion of women during their reproductive years, estimated at roughly one-fifth of this population <sup>[1]</sup>. In 2003, international reproductive medicine experts meeting in Rotterdam revised the diagnostic standards, leading to broader recognition of the syndrome's diverse clinical forms <sup>[2]</sup>. This diversity presents challenges for management, yet many patients show overlapping metabolic characteristics that are important for both evaluation and therapy <sup>[3]</sup>.

Many studies have shown that higher hormone levels, gut microbiome composition, and plasma metabolomics are new

parameters related to the PCOS phenotypes <sup>[4]</sup>. The clinical phenotypes can change over the life span with higher weight gain, and can found in the same patient. Individualised treatment remains the main approach, but grouping the phenotypes and following Therapeutic guidance may also have clinical value. Early adoption of well-defined management strategies is essential, particularly for females with PCOS, who face an elevated risk of developing endometrial or ovarian malignancies. <sup>[5,6]</sup>. Therefore, therapeutic strategies that incorporate anti-inflammatory agents as adjuncts to anticancer treatment are important. Such approaches can disrupt harmful signalling

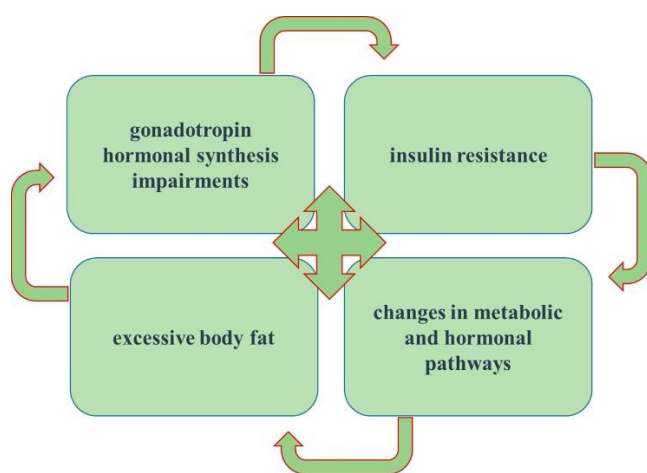
pathways, contributing to improved survival rates, quicker recovery, and enhanced quality of life for patients.

### 1.1. Physiological Basis

The four main causes of the physiological basis of PCOS include:

- disorders of gonadotropin hormonal synthesis;
- the appearance of insulin resistance;
- the influence of the present excessive body fat; and finally,
- the metabolic pathways involved in PCOS (the secretion and activity of insulin, encoding for steroidogenesis, and other metabolic and hormonal pathways) (Figure 1) [7].

**Figure 1.** Main pathophysiological basis of polycystic ovary syndrome (PCOS)-disorders of gonadotropin hormonal synthesis, the appearance of insulin resistance, the influence of the present excessive body fat and oblique metabolic pathways involved in PCOS.



Appropriate functioning of the mechanisms responsible for the maturation of the ovarian follicle and its ovulation depends on the proper physiological activity of three organs: the hypothalamus, pituitary gland, and ovaries.

Hormonal control within the hypothalamic–pituitary–ovarian (HPO) axis operates through long, short, and ultrashort negative feedback mechanisms. Neurons in the hypothalamic suprachiasmatic region synthesise gonadotropin-releasing hormone (GnRH), which enters the pituitary portal circulation via the median eminence. Its release depends on neuronal network activity and occurs in pulses that determine gonadotropin output. Slower GnRH pulses favour secretion of follicle-stimulating hormone (FSH), whereas faster pulses promote luteinizing hormone (LH) release from the anterior pituitary. LH drives corpus luteum formation and progesterone synthesis, while FSH supports follicular maturation and estrogen production by activating aromatase in granulosa cells. When LH predominates over FSH, androgen synthesis increases, a pattern often observed in polycystic ovary syndrome (PCOS) [8].

Insulin also contributes to PCOS pathophysiology by acting with LH to elevate androgen output and by reducing hepatic production of sex hormone-binding globulin (SHBG), thereby increasing free testosterone levels [8]. Excess adipose tissue further aggravates these processes because adipocytes release

hormones such as leptin and resistin and produce inflammatory mediators, including interleukin-1 $\beta$  and tumour necrosis factor- $\alpha$  [9]. The activity of leptin affects the function of the hypothalamus–pituitary gland–ovary axis by modifying the secretion of GnRH, LH, and FSH. Leptin acts on the hypothalamus to influence the release of gonadotropins, indirectly stimulating luteinizing hormone (LH) secretion and promoting gonadotropin-releasing hormone (GnRH) activity. This mechanism may enhance androgen production. Adipose tissue also releases inflammatory cytokines that sustain oxidative stress and inflammation in PCOS, conditions intensified by hyperglycemia, excess fat mass, and elevated androgens [8].

The clinical diversity of PCOS reflects the involvement of numerous metabolic pathways. These include insulin signaling and its related genes—such as those coding for the insulin receptor (IR), insulin (INS), and insulin-like growth factor (IGF) and its receptor—as well as genes linked to steroid hormone synthesis, cytochrome P450 activity (CYP17, CYP11A1), and hormone receptor function, including androgen receptor (AR), LH receptor, leptin, and follistatin [10]. Dietary habits emphasising anti-inflammatory foods and low glycemic index or reduced-fat intake appear to lower the risk of PCOS development [11,12].

### 1.2. Improvement in Metabolic Pathways

#### 1.2.1. Insulin Resistance

Weight gain mediates most of its direct medical sequelae through worsening insulin sensitivity.

Insulin resistance (IR) is central to the onset of metabolic disorders such as hypertension, impaired glucose control, and abnormal lipid profiles. Research indicates that mitochondrial impairment contributes to IR, often triggered by excess lipid accumulation in non-adipose tissues. The resulting oxidative stress in skeletal muscle increases reactive oxygen species (ROS) generation, further disrupting mitochondrial function and insulin signalling [13]. This mechanism links IR to obesity-associated conditions, including polycystic ovary syndrome (PCOS). The cellular effects of insulin occur through two main post-receptor pathways: the phosphatidylinositol 3-kinase (PI3K) and the mitogen-activated protein kinase (MAPK) pathways [14]. The PI3K pathway regulates cellular intermediary metabolism, whereas the MAPK pathway controls growth processes and mitoses [14]. AKR1C3 expression in adipocytes leads to the occurrence of insulin resistance and hyperinsulinemia, then drives a vicious circle of intra-adipose androgen activation, lipid accumulation, and hyperinsulinemia [15]. Kauffman et al. suggested that ethnicity has an additive effect on insulin resistance in PCOS. Mexican American women showed significantly higher insulin levels. Resistance compared with Caucasian American women [16].

#### 1.2.2 Oxidative Stress and Chronic Inflammation

The association between body weight and IR is mediated through inflammatory pathways [17]. Obesity causes changes in the release of key cytokines and adipokines, which in turn manifest in paracrine and endocrine effects. The increased levels of leptin

and plasminogen activator inhibitor-1 and the reduced release of adiponectin result in a generalised low-grade inflammatory response. This process is mediated by macrophages and other immune cells.

Elevated oxidative stress markers—such as reactive oxygen species (ROS), p47phox expression, and thiobarbituric acid-reactive substances (TBARS)—have been observed in women with PCOS following consumption of saturated fats, even when obesity is not present. Diets high in refined sugars and saturated fatty acids further intensify ROS formation through several pathways, including alterations in gut microbiota composition [18]. Both circulating immune cells and excess adipose tissue contribute independently to the oxidative imbalance characteristic of PCOS [19]. Lipid-driven oxidative stress appears to play a central role in the onset of insulin resistance and hyperandrogenism, with adipose tissue acting as an additional pro-oxidant source and modulator of insulin signalling [19]. Chronic androgen exposure also increases oxidative stress in pancreatic islet cells, leading to mitochondrial impairment [20,21]. Superoxide is a ROS produced when NADPH is oxidised by membrane-bound NADPH oxidase [22]. Abnormal generation of reactive oxygen species (ROS) by NADPH oxidase contributes to cardiovascular complications—such as endothelial dysfunction, atherosclerosis, and hypertension—that are frequently seen in women with PCOS [23]. Oxidative stress triggered by peroxides activates the nuclear factor  $\kappa$ B (NF- $\kappa$ B) pathway, a major regulator of inflammation that enhances the transcription of the tumour necrosis factor- $\alpha$  (TNF- $\alpha$ ) gene [24]. Intake of saturated fats can further intensify oxidative responses, promoting TNF- $\alpha$  release from leukocytes [19,25]. Our findings also indicate elevated TNF- $\alpha$  synthesis in women with PCOS [4]. Those with normal or low androgen concentrations, based on total testosterone and the free androgen index (FAI), appear particularly sensitive to TNF- $\alpha$ -driven oxidative and inflammatory changes [26].

### 1.2.3 Anticancer Protection

Many studies have targeted the inactivation of the transcription factor (NRF2) as a therapeutic approach in various types of cancer [27]. NRF2 was first recognised in anticancer research as an inducer of several antioxidant enzymes. It can protect cells and tissues against many types of toxicants that interrupt essential biochemical processes and carcinogens by increasing the expression of cytoprotective genes [28]. The transcription factor NRF2 exhibits context-dependent behaviour, functioning either as a tumour suppressor or as a promoter of tumour progression, depending on the biological setting in which it is activated [29]. Moderate activation of NRF2 in healthy cells can limit oxidative damage and genomic instability, thereby lowering cancer risk. Conversely, persistent or constitutive NRF2 activity in malignant cells can promote survival advantages, treatment resistance, and poor clinical outcomes, often necessitating therapeutic inhibition of the pathway [29]. NRF2 stability is regulated through at least three distinct mechanisms. One involves the cytoplasmic repressor KEAP1 [30]; another relies on  $\beta$ -transducin repeat-containing protein ( $\beta$ -

TrCP) [31]; and a third is mediated by the endoplasmic reticulum-associated E3 ubiquitin ligase HRD1 [32].

The abnormal activation of the NRF2/KEAP1 pathway promotes cancer development [33], metastasis formation [34], and even resistance to ovarian cancer therapy [35]. Mutations in the KEAP1 gene induce the hyperactivation of the NRF2/KEAP1 pathway. Notably, KEAP1 missense or nonsense mutations were reported in endometrial carcinomas [36], as well as gall bladder [37], breast [38,39], cervical [40], and ovarian [41,42] cancers. MicroRNA miR-141 was the first-identified miRNA to directly repress KEAP1 levels in ovarian carcinoma cell lines [43].

### 1.3. Gut Microbiota Dysbiosis

The structural and functional dysbiosis of the gut microbiota in high-fat diet (HFD)-induced obesity was demonstrated in a mouse model [44]. Gut microorganisms and their metabolites exert broad influences on appetite regulation, lipid and glucose metabolism, and overall body weight control [44,45]. The intestinal microbiome can modulate roughly 10% of the host transcriptome, affecting immune, metabolic, and proliferative gene networks [46]. Dietary fibre, fermentation processes, and probiotic intake have therefore become major research areas in metabolic health [47]. Evidence shows that dietary fibre can restore microbial balance in individuals with type 2 diabetes [48]. Growth of Bifidobacterium species supports insulin release, enhances glucose tolerance, improves insulin sensitivity, and reduces inflammation. Beneficial microbes also generate short-chain fatty acids (SCFAs), including acetate and butyrate, which influence blood glucose via enteroendocrine hormones such as glucagon-like peptide-1 (GLP-1) and peptide YY (PYY) [45,49]. PYY, acting locally in the gut and centrally in the brain, contributes to appetite regulation. Because SCFAs are integral to lipid and carbohydrate metabolism, maintaining a healthy gut microbiota is an important therapeutic aim for reducing inflammation and preventing urogenital tract infections [50–52].

## 2. Lifestyle Changes

Lifestyle modification remains the primary therapeutic approach in the management of polycystic ovary syndrome (PCOS), though it complements rather than replaces pharmacological therapy [7]. Consistent physical activity, maintenance of healthy body weight, balanced dietary habits, and the avoidance of smoking are fundamental in the prevention and treatment of metabolic complications. These elements are incorporated into most clinical guidelines addressing metabolic and reproductive health. Attention to psychological well-being and stress reduction is also essential, as sustained behavioural change contributes to overall quality of life. Nutritional counselling has long been a cornerstone of PCOS management. However, studies indicate that severe caloric restriction rarely achieves lasting metabolic or hormonal improvement [53,54]. Even isocaloric diets, when paired with physical activity, may not significantly alter biochemical or anthropometric parameters [55].

## 2.1. Diet

Examination of macronutrient composition—relative proportions of protein, fat, and carbohydrates—has revealed no major differences in key metabolic indicators. Instead, total caloric reduction and the adoption of a diet with a low glycemic index (GI) are the most consistent predictors of clinical improvement [56,57]. Low-GI (LGI) diets have been shown to reduce insulin resistance (HOMA-IR), fasting insulin, total and LDL cholesterol, triglycerides, waist circumference, and total testosterone compared with high-GI (HGI) diets, without major changes in fasting glucose, HDL, body weight, or the free androgen index [58]. Combining an LGI diet with moderate caloric restriction, physical activity, and omega-3 fatty acid supplementation further increases HDL levels, promotes synthesis of sex hormone-binding globulin (SHBG), and reduces adiposity [8].

Dietary patterns rich in saturated fatty acids (SFA) can elevate circulating tumour necrosis factor- $\alpha$  (TNF- $\alpha$ ) and leukocytic suppressor of cytokine signalling-3 (SOCS-3) expression, suggesting that limiting SFA intake is particularly important for PCOS management [25]. Sources of  $\alpha$ -linolenic acid, such as flaxseed oil, have demonstrated favourable effects on hormonal and inflammatory markers in animal models, implying potential benefits in humans as well [59].

Soluble, fermentable dietary fibre promotes short-chain fatty acid (SCFA) formation, which supports a healthy gut microbiome and improves metabolic outcomes [60]. Diets with a low GI also modulate appetite-regulating hormones, decreasing ghrelin and increasing glucagon secretion in women with PCOS [12,61]. Conversely, excessive fructose intake may exacerbate endocrine abnormalities, worsening hormonal imbalances despite limited metabolic changes [62]. Meta-analyses confirm that LGI diets are safe, practical, and effective for improving insulin resistance, underscoring the need for individualised dietary guidance in all PCOS patients [63,64].

The ketogenic diet (KD) represents another form of carbohydrate restriction that replaces a portion of dietary carbohydrates with plant-derived fats. In women with PCOS—especially those with obesity or fatty liver disease—KD has been associated with improved menstrual regularity, reduced body mass, lower glucose and insulin levels, and better liver function [65]. A 12-week trial demonstrated significant reductions in body weight ( $\approx 9.4$  kg), BMI ( $\approx 3.3$  units), and fat mass ( $\approx 8.3$  kg), along with improved lipid profiles, reduced triglycerides and LDL cholesterol, elevated HDL cholesterol, and normalisation of hormonal ratios, including LH/FSH and androgens [66]. Estradiol, progesterone, and SHBG levels increased, while the Ferriman–Gallwey score showed a modest decline, indicating partial improvement in hirsutism. The absence of correlation between hirsutism and the visceral adiposity index (VAI) suggests that hair growth disorders are not directly driven by visceral fat dysfunction [67].

For women with obesity or metabolic syndrome, a ketogenic or low-GI diet may produce greater metabolic and hormonal benefits than standard dietary interventions. Overall, adherence to a nutrient-balanced, calorie-controlled eating plan remains key

for restoring physiological equilibrium and promoting recovery in PCOS.

### 2.1.1 Physical Activity

Exercise training in the management of PCOS is becoming more recognised and accepted among professionals in the health sector and patients. Physical training potentiates the effects caused by insulin sensitivity through the optimisation of glucose transport and metabolism [68].

Recent evidence indicates that the intensity of physical activity has a stronger impact on health outcomes than total exercise volume. Vigorous-intensity activity appears to produce the largest improvements in cardiorespiratory capacity, insulin resistance, and body composition among women with PCOS [69]. Significant reductions in insulin resistance, measured by HOMA-IR, and in BMI have been observed following moderate- and high-intensity programs (MD  $-0.57$ ; 95% CI  $-0.98$  to  $-0.16$ ,  $p = 0.01$ ; and MD  $-1.90$ ; 95% CI  $-3.37$  to  $-0.42$ ,  $p = 0.01$ , respectively) [70]. Systematic reviews recommend incorporating both aerobic and resistance exercise to optimise insulin sensitivity and androgen balance in this population [71]. A minimum of approximately 120 minutes of aerobic activity per week is generally advised [69].

### 2.1.2. Sleep

Psychological disturbances such as anxiety, depression, and sleep abnormalities occur more frequently in women with PCOS [72]. Sleep dysregulation contributes to both the onset and progression of metabolic and emotional disturbances; therefore, treating sleep issues is a critical component of PCOS management [72]. Insufficient or fragmented sleep is linked to a higher risk of insulin resistance, obesity, and type 2 diabetes [73–75]. The mechanisms appear to involve autonomic dysregulation, hormonal fluctuations in leptin and ghrelin, and inflammatory signalling. Experimental models show that chronic sleep fragmentation leads to inflammation in white adipose tissue and aggravates insulin resistance, partly through disruption of intestinal barrier function and lipopolysaccharide-mediated inflammation (“gut leakage”) [51,76]. These findings suggest that microbiota-targeted therapy may mitigate some of the metabolic effects of poor sleep [76].

Melatonin, the main hormone secreted by the pineal gland, regulates circadian rhythm and exhibits strong antioxidant properties. In PCOS, reduced melatonin concentrations have been reported in follicular fluid [77]. Melatonin receptors within ovarian tissue influence steroidogenesis during follicular maturation, and sufficient melatonin levels help protect developing oocytes from oxidative damage [77]. Overall, sleep disorders may represent an early contributor to diminished physiological resilience and worsening insulin resistance in PCOS.

### 2.1.3. Supplementation

Dietary surveys reveal that many women with PCOS consume unbalanced diets lacking fibre, omega-3 fatty acids, and key micronutrients such as calcium, magnesium, zinc, folate,



vitamins C, B12, and D, while showing excessive intake of sucrose, sodium, saturated fat, and cholesterol [8]. Calorie-controlled, low-glycemic-index diets can correct some deficiencies, especially of water-soluble vitamins [78,79]. Improved plasma concentrations of most B vitamins have been noted after dietary adjustment, though vitamin B3 responses remain suboptimal [79]. Inadequate niacin (B3) intake has been associated with inflammation and higher cardiovascular risk [80,81].

Metformin therapy, while beneficial for glycemic control, may reduce thiamine and cobalamin stores, warranting supplementation [82]. Thiamine enhances transketolase activity and supports vascular protection, potentially reducing cardiovascular complications [83,84]. Coenzyme Q10 (CoQ10) supplementation for eight weeks has been shown to improve inflammatory status and endothelial function in overweight and obese women with PCOS [85]. Vitamin D plays multiple metabolic roles, enhancing insulin synthesis, receptor expression, and response to glucose [86]. It indirectly modulates carbohydrate metabolism by maintaining calcium–parathyroid hormone balance and by suppressing pro-inflammatory cytokine expression [87]. Weekly supplementation with 20,000 IU cholecalciferol improved fasting glucose, triglycerides, estradiol levels, and menstrual regularity, although androgen levels remained unchanged [88].

Combined supplementation with magnesium, zinc, calcium, and vitamin D produced reductions in hirsutism and total testosterone but did not affect SHBG or the free androgen index [89]. Similarly, vitamin D administered with fish oil reduced serum C-reactive protein, downregulated interleukin-1 expression, lowered testosterone levels, and improved mood scores in women with PCOS [90]. Overall, nutritional optimisation, targeted vitamin and mineral replacement, and antioxidant support represent important adjuncts to lifestyle and pharmacologic therapy in PCOS.

Current evidence indicates that myo-inositol provides metabolic and hormonal improvements in women with PCOS comparable to those achieved with metformin, particularly regarding insulin sensitivity and glucose regulation, but without the gastrointestinal side effects often associated with metformin therapy [91,92]. Supplementation with inositol enhances tissue responsiveness to insulin, lowers circulating androgen concentrations, improves glycemic control, and positively affects several markers of metabolic syndrome [93,94]. In PCOS, an excessive conversion of myo-inositol (MI) to D-chiro-inositol (DCI) has been observed in the ovaries under the influence of insulin, leading to a local shortage of MI and an excess of DCI. This imbalance may impair follicle-stimulating hormone (FSH) signalling and compromise oocyte maturation and quality [95]. Clinical studies suggest that using inositol isomers—either individually or in combination—can restore spontaneous ovulation, support folliculogenesis, and improve conception rates in women with PCOS. Literature reviews consistently identify inositol supplementation as a safe and effective therapeutic option that enhances ovarian function, oocyte development, and pregnancy outcomes [96].

Traditional and complementary medicine approaches have also explored the use of natural compounds such as isoquinoline alkaloids to regulate androgen synthesis and lipid and carbohydrate metabolism. Berberine, one such alkaloid, has attracted attention for its multiple beneficial effects in PCOS management [97–99]. Its metabolic action resembles that of metformin, largely through activation of adenosine monophosphate-activated protein kinase (AMPK), leading to improvements in glucose and lipid profiles, reduction in body mass, and increased insulin sensitivity [100]. Berberine additionally influences the hypothalamic–pituitary–ovarian axis by reducing the synthesis of steroid hormones and downregulating ovarian aromatase, which contributes to improved ovulatory function, menstrual regularity, and higher pregnancy and live-birth rates. Long-term use has been associated mainly with mild and transient adverse effects such as nausea or constipation, indicating a favourable safety profile [98,101,102].

Chromium, an element involved in carbohydrate and lipid metabolism, has long been included in dietary supplements marketed for metabolic health in the United States [103]. Although the essentiality of chromium remains debated, some findings suggest it can enhance insulin signalling, promote AMPK activity, and increase cellular glucose uptake, thereby benefiting patients with PCOS and type 2 diabetes [104–106]. Furthermore, alterations in the expression of steroidogenic enzymes—specifically 3 $\beta$ -hydroxysteroid dehydrogenase and 17 $\beta$ -hydroxysteroid dehydrogenase—in adipose tissue have been linked to dehydroepiandrosterone metabolism, suggesting that chromium may indirectly affect androgen balance [107].

Evidence from clinical and experimental research indicates that supplementation with trace minerals such as zinc and selenium may be beneficial for some women with PCOS. Zinc is involved in numerous intracellular processes, serving both structural and signalling functions that influence glucose and lipid metabolism as well as reproductive health [108]. Insufficient zinc intake, particularly among individuals with obesity, has been linked to hyperinsulinemia, chronic low-grade inflammation, and an adverse lipid profile. In adipose tissue, zinc ions can act similarly to insulin by promoting glucose uptake via translocation of the glucose transporter GLUT4 to the plasma membrane and by stimulating lipogenesis [109]. Several studies report that women with PCOS have lower serum zinc concentrations than healthy controls, and those with impaired glucose tolerance exhibit the lowest levels [110]. Selenium, another essential micronutrient, demonstrates potent anti-inflammatory and antioxidant actions and shows an inverse correlation with circulating C-reactive protein (CRP) levels [111]. Maintaining adequate selenium and zinc status may therefore support metabolic equilibrium and reduce the inflammatory burden characteristic of PCOS.

Omega-3 fatty acids are also frequently deficient in the diets of women with PCOS. When an overall balanced diet is followed, targeted omega-3 supplementation may be required only seasonally or during periods of dietary imbalance [112]. Polyunsaturated fatty acids (PUFAs) contribute to improved ovarian function by enhancing the expression of steroidogenic

enzymes—such as CYP51, CYP19, StAR, and 3 $\beta$ -HSD—which regulate hormone synthesis and reproductive performance [113]. Supplementation should always be individualised and monitored by a qualified dietitian to ensure patient adherence and safety. Active participation by the patient remains crucial for achieving a lasting improvement in metabolic homeostasis. A nutritionally adequate diet, combined with regular exercise and stress control, continues to represent the cornerstone of PCOS therapy.

## 2.2. Herbs Supporting Treatment

A nutritionally balanced plan that stabilises insulin response can be effectively complemented by selected medicinal herbs. Botanical preparations such as Aloe vera, cinnamon (*Cinnamomum verum*), green tea (*Camellia sinensis*), chamomile (*Matricaria chamomilla*), and white mulberry (*Morus alba*) have demonstrated beneficial effects on glucose and lipid metabolism and may also exert mild anti-inflammatory activity [114,115]. Because of these properties, such herbal agents can be applied to different PCOS phenotypes, especially where metabolic disturbances dominate. Green tea [116] and marjoram (*Majorana hortensis*) have been shown to improve insulin sensitivity, restore hormonal balance, and enhance antioxidant and anti-inflammatory parameters in both clinical and preclinical studies [117,118].

For women with elevated androgen levels, specific antiandrogenic herbs may be helpful. Spearmint (*Mentha spicata* L.) reduces circulating testosterone concentrations and supports follicular development in ovarian tissue [119,120]. Liquorice (*Glycyrrhiza glabra*), long used in traditional medicine, contains phytoactive molecules that exert both estrogen-like and antiandrogenic actions. Glycyrrhetic acid, one of its principal metabolites, inhibits 11 $\beta$ -hydroxysteroid dehydrogenase type 2 and binds to mineralocorticoid receptors, thereby influencing steroid metabolism [121,122]. Despite its therapeutic potential, liquorice can increase blood pressure and alter potassium balance; individuals with high cortisol levels or cardiovascular disease should therefore use it cautiously [123]. Other botanical extracts possess 5- $\alpha$ -reductase inhibitory activity and may counteract androgen-driven hair loss. *Serenoa repens*, *Camellia sinensis*, *Rosmarinus officinalis*, and *Glycyrrhiza glabra* have demonstrated the capacity to reduce androgen concentrations and slow the progression of androgenetic alopecia [124].

*Vitex agnus-castus* remains one of the best-studied herbal agents for restoring menstrual cyclicity and alleviating premenstrual symptoms through dopaminergic and pituitary modulation [125]. Phytoestrogen-rich sources such as flaxseed (*Linum usitatissimum*), abundant in lignans, may regulate aromatase activity and influence estrogen metabolism, thereby helping to normalise sex hormone ratios [126,127]. Turmeric (*Curcuma longa*) and its main polyphenolic constituent, curcumin, have well-documented antioxidant and anti-inflammatory effects. In PCOS, curcumin supplementation has been associated with reductions in oxidative stress markers and inhibition of NF- $\kappa$ B-dependent inflammatory signalling [128–132]. *Urtica dioica* (nettle) displays wide-ranging pharmacological activities, including antioxidant, antimutagenic, and anti-inflammatory effects that may aid metabolic regulation [133,134]. Plant-derived flavonoids, present in many of these herbs, can neutralise reactive oxygen species such as peroxides and hydroxyl radicals, protecting tissues from oxidative damage and supporting cellular resilience [135]. In more advanced PCOS accompanied by metabolic syndrome or non-alcoholic fatty liver disease, hepatoprotective plants can be useful adjuncts. Extracts from milk thistle (*Silybum marianum*), which contain silymarin, have shown antioxidant and membrane-stabilising effects that protect hepatocytes from oxidative injury [136–138]. Artichoke (*Cynara cardunculus*) provides sesquiterpene lactones and phenolic acids with similar hepatoprotective properties [139,140]. Compounds from dandelion (*Taraxacum officinale*), particularly taraxasterol, activate SIRT1-dependent pathways that safeguard liver cells [141]. *Nigella sativa* (black cumin) also exhibits antioxidant and anti-inflammatory actions, and may reduce hepatic steatosis and insulin resistance in obese women with PCOS [142].

In summary, herbal and micronutrient supplementation offer numerous complementary strategies for PCOS management. The combined actions of antioxidant, antiandrogenic, and hepatoprotective mechanisms contribute to restoring metabolic and hormonal equilibrium. Individualised selection of herbal mixtures and nutritional support—guided by clinical assessment and evidence-based practice—can enhance the efficacy of standard medical therapies and improve the overall health outcomes of women living with PCOS. Summary information has been added in Table 1.

**Table 1:** Overview of herbal interventions and their reported outcomes in PCOS

A Symptom Accompanying PCOS	Diet	Physical Activity	Sleep	Supplementation	Microbiota	Herbs
Hirsutism	Reduced Diet [26,44,45,54,58]	Daily Physical Activity [68–71]	Improving Sleep [72–77]	Magnesium, Zinc, Calcium [89,108–111], Vitamin D [86–90], Myo-Inositol [93–96]	Microbiota And Metabolites [46,47]	Gl Glycyrrhiza Glabra, Serenoa Repens, Camellia Sinensis [120,121], Rosmarinus Officinalis [122]
The androgen levels	Diet With Reduced GI And Calorie [26,44,45,54,58], Ketogenic Diet [64]	Daily Physical Activity [68–71]	Improving Sleep [72–77]	Magnesium, Zinc, Calcium [89,108–111], Vitamin D [86–90], Berberine [97–102], Chromium [105–107], Zinc [110]	SCFA [47,52], Microbiota and Metabolites [50]	Mentha Spicata [120,121], Glycyrrhiza Glabra [122], Serenoa Repens, Camellia Sinensis, Rosmarinus Officinalis [124]
Ovulation disorders	Diet With Reduced GI And Calorie [26,44,45,54,58], Ketogenic Diet [64]	Daily Physical Activity [71–74]	Improving Sleep [72–77]	Vitamin D [86–90], Myo-Inositol [97,98], Berberine [99], Zinc [108], Pufas [112,113]	Bifidobacteria [45,50]	Glycyrrhiza Glabra [121], Vitex Agnus-Castus [124], Flaxseed (Linum Usitatissimum) [59,125,126]
Fat mass reduction	High-Fibre Diet with Reduced GI And Calorie [28,46,47,56,60], Ketogenic Diet [64], Elimination Of SFA [22,58]	Daily Physical Activity [68–71]	Melatonin [77]	Vitamin B1 [82–84], Vitamin D [86–90], Myo-Inositol [91–96], Berberine [97–102], Chromium [105–107], Zinc [109]	SCFA [47,52]; Microbiota and Metabolites [50]	Aloe Vera, Cinnamomum Verum, Camellia Sinensis [115], Matricaria Chamomilla, Morus Alba [117]
Carbohydrate metabolism disorders	High-Fibre Diet With Reduced GI And Calorie [26,44,45,54,58], Ketogenic Diet [64]	—	—	Vitamin D [86–90], Myo-Inositol [91–96], Berberine [97–102]	—	Aloe Vera, Cinnamon, Green Tea [115], Chamomile, White Mulberry [117]
Insulin resistance	High-Fibre Diet With Reduced GI And Calorie [26,44,45,54,58], Elimination SFA [22,58]	Intensity Exercise [72]	—	Omega-3 [112,113], Berberine [97–102], Zinc [110]	—	Milk Thistle [137,138], Artichoke Extract [139,140], Dandelion [141], Black Cumin [142]
Lipids metabolism disorders	High-Fibre Diet With Reduced GI And Calorie [26,44,45,54,58], Elimination Of SFA [25,60]	—	—	—	Bifidobacteria [45,50]	Milk Thistle [137,138], Artichoke Extract [139,140], Dandelion [141], Black Cumin [142]
Steatosis of organs – liver profile	High-Fibre Diet With Reduced GI And Calorie [46,47,56,60]	—	—	A-Linolenic Acid [59], Vitamin B3 [80,81], Vitamin B1 [82–84], Coenzyme Q10 [85]	—	—
Cardiovascular diseases	High-Fibre Diet With Reduced GI And Calorie [46,47,56,60]	—	—	—	—	—
Intestinal dysbiosis	High-Fibre Diet [49,50]	—	—	—	—	—
A Symptom Accompanying PCOS	Diet	Physical Activity	Sleep	Supplementation	Microbiota	Herbs
Chronic inflammation	High-Fibre Diet With Reduced GI And Calorie [28,46,47,56,60]	Daily Physical Activity [71–74]	Melatonin [79]	A-Linolenic Acid [59], Vitamin B3 [80,81], Coenzyme Q10 [85], Vitamin D [88,89], Selenium [112], Flavonoids [135]	Bifidobacteria [45,50]	Marjoram [117–119], Turmeric [128–131], Nettle [133,134], Milk Thistle [137,138], Artichoke Extract [139,140], Dandelion [141], Black Cumin [142]
Limiting predisposition to cancer	Elimination SFA [25,27]; High-Fiber Diet [49,50]	—	Improving Sleep [75]	A-Linolenic Acid [59]	—	Turmeric [128–131], Nettle [133,134]

SCFA—short-chain fatty acids;

GI—glycemic index;

SFA—saturated fat acids;

PUFA—Polyunsaturated fatty acid.

### 3. CONCLUSIONS

Metabolic dysregulation associated with polycystic ovary syndrome (PCOS) requires a multifaceted therapeutic approach that simultaneously addresses hormonal, reproductive, and metabolic disturbances. These disturbances stem from several interacting biochemical pathways; therefore, clinical management should emphasize improving fertility outcomes, minimizing androgen-related symptoms, and restoring glucose–lipid metabolism and insulin response. Lifestyle modification remains central to therapy, with evidence supporting the benefits of a calorie-controlled, low-glycemic-index diet, adequate sleep, and consistent physical activity in reducing cardiometabolic risk and enhancing overall health in affected women.

Adjunct nutritional and phytotherapeutic strategies have gained attention as supportive interventions in PCOS. Bioactive compounds with antioxidant and hepatoprotective properties may contribute to reduced inflammation and improved metabolic status. Specific agents—such as extracts from *Curcuma longa* or *Silybum marianum* and probiotics aimed at restoring gut microflora—have shown preliminary benefits in clinical and experimental contexts. In the present study, nutrient-intake patterns and supplementation practices were evaluated to understand their influence on micronutrient balance among women with PCOS. Emerging evidence also highlights potential roles for inositols, thiamine, coenzyme Q10, vitamin D, zinc, and selenium in optimizing metabolic and reproductive outcomes.

Although these findings indicate encouraging trends, definitive conclusions cannot yet be drawn. Larger, well-designed trials are needed to validate the efficacy, safety, and long-term clinical significance of these nutritional and herbal interventions when combined with standard medical care..

### 4. Methods of Searching

This review focused on non-pharmacological approaches to the management of PCOS. A comprehensive literature search was carried out in the PubMed and Embase (Elsevier) databases, covering research published within the past twenty years. All retrieved records were screened at the abstract level. Publications unrelated to the primary topic, duplicated between the two databases, or limited to conference proceedings were excluded. Only articles written in English were included for full-text review. The discussion also draws on the authors' decade of clinical and research experience with PCOS patients. From this work, studies corresponding to each stage of the interventions discussed were selected. The literature search emphasized physiological mechanisms linking PCOS with insulin resistance, chronic inflammation, endocrine dysfunction, cancer development, and gut microbiota alterations. Lifestyle modification studies were analyzed first, followed by those investigating diet and supplementation—including inositol, berberine, vitamin D, chromium, zinc, selenium, and melatonin—along with reports evaluating herbal adjunct therapies. In cases of overlapping information across

publications, the most comprehensive and relevant studies were prioritised for inclusion.

### REFERENCES

1. Fauser, B.C.J.M.; Tarlatzis, B.C.; Rebar, R.W.; Legro, R.S.; Balen, A.H.; Lobo, R.; Carmina, E.; Chang, J.; Yildiz, B.O.; Laven, J.S.E.; et al. Consensus on Women's Health Aspects of Polycystic Ovary Syndrome (PCOS): The Amsterdam ESHRE/ASRM-Sponsored 3rd PCOS Consensus Workshop Group. *Fertil. Steril.* 2023 *97*, 28–38.e25. [[CrossRef](#)]
2. Zuo, M.; Liao, G.; Zhang, W.; Xu, D.; Lu, J.; Tang, M.; Yan, Y.; Hong, C.; Wang, Y. Effects of Exogenous Adiponectin Supplementation in Early Pregnant PCOS Mice on the Metabolic Syndrome of Adult Female Offspring. *J. Ovarian Res.* 2018, *14*, 15. [[CrossRef](#)]
3. Szczuko, M.; Zapałowska-Chwyc', M.; Maciejewska, D.; Drozd, A.; Starczewski, A.; Stachowska, E. Significant Improvement Selected Mediators of Inflammation in Phenotypes of Women with PCOS after Reduction and Low GI Diet. *Mediat. Inflamm.* 2017, *2017*, 5489523. [[CrossRef](#)]
4. Ma, L.; Cao, Y.; Ma, Y.; Zhai, J. Association between hyperandrogenism and adverse pregnancy outcomes in patients with different polycystic ovary syndrome phenotypes undergoing in vitro fertilization/intracytoplasmic sperm injection: A systematic review and meta-analysis. *Gynecol. Endocrinol.* 2017, 1–8. [[CrossRef](#)] [[PubMed](#)]
5. Martini, A.E.; Healy, M.W. Polycystic Ovarian Syndrome: Impact on Adult and Fetal Health. *Clin. Obs. Gynecol.* 2018, *64*, 26–32. [[CrossRef](#)]
6. Hong, G.; Wu, H.; Ma, S.-T.; Su, Z. Catechins from Oolong Tea Improve Uterine Defects by Inhibiting STAT3 Signaling in Polycystic Ovary Syndrome Mice. *Chin. Med.* 2018, *15*, 125. [[CrossRef](#)]
7. Del Pup, L.; Cagnacci, A. IMPROVE Lifestyle in Polycystic Ovary Syndrome: A Systematic Strategy. *Gynecol. Endocrinol.* 2018, 1–4. [[CrossRef](#)]
8. Szczuko, M.; Skowronek, M.; Zapałowska-Chwyc', M.; Starczewski, A. Quantitative Assessment of Nutrition in Patients with Polycystic Ovary Syndrome (PCOS). *Rocz. Panstw. Zakl. Hig.* 2016, *67*, 419–426. [[PubMed](#)]
9. Makki, K.; Froguel, P.; Wolowczuk, I. Adipose Tissue in Obesity-Related Inflammation and Insulin Resistance: Cells, Cytokines, and Chemokines. *ISRN Inflamm.* 2013, *2013*, 139239. [[CrossRef](#)] [[PubMed](#)]
10. Dniak-Nikolajew, A. Zespół Policystycznych Jajników Jako Przyczyna Niepłodności Kobiecej [Polycystic Ovary Syndrome as a Cause of Female Infertility]. *Położ'na Nauka I Prakt.* 2012, *17*, 14–17.
11. Panjeshahin, A.; Salehi-Abargouei, A.; Anari, A.G.; Mohammadi, M.; Hosseinzadeh, M. Association between Empirically Derived Dietary Patterns and Polycystic Ovary Syndrome: A Case-Control Study. *Nutrition* 218, 79–80, 110987. [[CrossRef](#)]



12. Szczuko, M.; Zapalowska-Chwyc', M.; Drozd, R. A Low Glycemic Index Decreases Inflammation by Increasing the Concentration of Uric Acid and the Activity of Glutathione Peroxidase (GPx3) in Patients with Polycystic Ovary Syndrome (PCOS). *Molecules* 2018, *24*, 1508. [[CrossRef](#)]
13. Di Meo, S.; Iossa, S.; Venditti, P. Skeletal Muscle Insulin Resistance: Role of Mitochondria and Other ROS Sources. *J. Endocrinol.* 2017, *233*, R15–R42. [[CrossRef](#)]
14. Barber, T.M.; Kyrou, I.; Randeva, H.S.; Weickert, M.O. Mechanisms of Insulin Resistance at the Crossroad of Obesity with Associated Metabolic Abnormalities and Cognitive Dysfunction. *Int. J. Mol. Sci.* 2021, *22*, 546. [[CrossRef](#)]
15. Kempegowda, P.; Melson, E.; Manolopoulos, K.N.; Arlt, W.; O'Reilly, M.W. Implicating Androgen Excess in Propagating Metabolic Disease in Polycystic Ovary Syndrome. *Adv. Endocrinol. Metab.* 2022, *11*. [[CrossRef](#)]
16. Kauffman, R.P.; Baker, V.M.; Dimarino, P.; Gimpel, T.; Castracane, V.D. Polycystic Ovarian Syndrome and Insulin Resistance in White and Mexican American Women: A Comparison of Two Distinct Populations. *Am. J. Obs. Gynecol.* 2002, *187*, 1362–1369. [[CrossRef](#)] [[PubMed](#)]
17. Shoelson, S.E.; Herrero, L.; Naaz, A. Obesity, Inflammation, and Insulin Resistance. *Gastroenterology* 2007, *132*, 2169–2180. [[CrossRef](#)] [[PubMed](#)]
18. Fajstova, A.; Galanova, N.; Coufal, S.; Malkova, J.; Kostovcik, M.; Cermakova, M.; Pelantova, H.; Kuzma, M.; Sediva, B.; Hudcovic, T.; et al. Diet Rich in Simple Sugars Promotes Pro-Inflammatory Response via Gut Microbiota Alteration and TLR4 Signaling. *Cells* 2018, *9*, 2701. [[CrossRef](#)]
19. González, F.; Considine, R.V.; Abdelhadi, O.A.; Acton, A.J. Oxidative Stress in Response to Saturated Fat Ingestion Is Linked to Insulin Resistance and Hyperandrogenism in Polycystic Ovary Syndrome. *J. Clin. Endocrinol. Metab.* 2018, *104*, 5360–5371. [[CrossRef](#)]
20. Wang, H.; Wang, X.; Zhu, Y.; Chen, F.; Sun, Y.; Han, X. Increased Androgen Levels in Rats Impair Glucose-Stimulated Insulin Secretion through Disruption of Pancreatic Beta Cell Mitochondrial Function. *J. Steroid Biochem. Mol. Biol.* 2015, *154*, 254–266. [[CrossRef](#)] [[PubMed](#)]
21. Liu, S.; Navarro, G.; Mauvais-Jarvis, F. Androgen Excess Produces Systemic Oxidative Stress and Predisposes to Beta-Cell Failure in Female Mice. *PLoS ONE* 2010, *5*, e11302. [[CrossRef](#)]
22. Jiang, F.; Zhang, Y.; Dusting, G.J. NADPH Oxidase-Mediated Redox Signaling: Roles in Cellular Stress Response, Stress Tolerance, and Tissue Repair. *Pharm. Rev.* 2011, *63*, 218–242. [[CrossRef](#)] [[PubMed](#)]
23. Bedard, K.; Krause, K.-H. The NOX Family of ROS-Generating NADPH Oxidases: Physiology and Pathophysiology. *Physiol. Rev.* 2007, *87*, 245–313. [[CrossRef](#)] [[PubMed](#)]
24. Evans, J.L.; Goldfine, I.D.; Maddux, B.A.; Grodsky, G.M. Oxidative Stress and Stress-Activated Signaling Pathways: A Unifying Hypothesis of Type 2 Diabetes. *Endocr. Rev.* 2002, *23*, 599–622. [[CrossRef](#)] [[PubMed](#)]
25. González, F.; Considine, R.V.; Abdelhadi, O.A.; Acton, A.J. Saturated Fat Ingestion Promotes Lipopolysaccharide-Mediated Inflammation and Insulin Resistance in Polycystic Ovary Syndrome. *J. Clin. Endocrinol. Metab.* 2018, *104*, 934–946. [[CrossRef](#)] [[PubMed](#)]
26. Szczuko, M.; Zapalowska-Chwyc', M.; Maciejewska, D.; Drozd, A.; Starczewski, A.; Stachowska, E. High Glycemic Index Diet in PCOS Patients. The Analysis of IGF I and TNF- $\alpha$  Pathways in Metabolic Disorders. *Med. Hypotheses* 2016, *96*, 42–47. [[CrossRef](#)]
27. Panieri, E.; Saso, L. Potential Applications of NRF2 Inhibitors in Cancer Therapy. *Oxidative Med. Cell. Longev.* 2018, *2019*, e8592348. [[CrossRef](#)]
28. Panieri, E.; Buha, A.; Telkoparan-Akillilar, P.; Cevik, D.; Kouretas, D.; Veskokis, A.; Skaperda, Z.; Tsatsakis, A.; Wallace, D.; Suzen, S.; et al. Potential Applications of NRF2 Modulators in Cancer Therapy. *Antioxidants* 2018, *9*, 193. [[CrossRef](#)]
29. Panieri, E.; Telkoparan-Akillilar, P.; Suzen, S.; Saso, L. The NRF2/KEAP1 Axis in the Regulation of Tumor Metabolism: Mechanisms and Therapeutic Perspectives. *Biomolecules* 2018, *10*, 791. [[CrossRef](#)] [[PubMed](#)]
30. Suzuki, T.; Yamamoto, M. Stress-Sensing Mechanisms and the Physiological Roles of the Keap1-Nrf2 System during Cellular Stress. *J. Biol. Chem.* 2017, *292*, 16817–16824. [[CrossRef](#)]
31. Chowdhry, S.; Zhang, Y.; McMahon, M.; Sutherland, C.; Cuadrado, A.; Hayes, J.D. Nrf2 Is Controlled by Two Distinct  $\beta$ -TrCP Recognition Motifs in Its Neh6 Domain, One of Which Can Be Modulated by GSK-3 Activity. *Oncogene* 2013, *32*, 3765–3781. [[CrossRef](#)]
32. Wu, T.; Zhao, F.; Gao, B.; Tan, C.; Yagishita, N.; Nakajima, T.; Wong, P.K.; Chapman, E.; Fang, D.; Zhang, D.D. Hrd1 Suppresses Nrf2-Mediated Cellular Protection during Liver Cirrhosis. *Genes Dev.* 2014, *28*, 708–722. [[CrossRef](#)]
33. Rojo, A.I.; Rada, P.; Mendiola, M.; Ortega-Molina, A.; Wojdyla, K.; Rogowska-Wrzesinska, A.; Hardisson, D.; Serrano, M.; Cuadrado, A. The PTEN/NRF2 Axis Promotes Human Carcinogenesis. *Antioxid. Redox. Signal.* 2014, *21*, 2498–2514. [[CrossRef](#)] [[PubMed](#)]
34. Zhang, C.; Wang, H.-J.; Bao, Q.-C.; Wang, L.; Guo, T.-K.; Chen, W.-L.; Xu, L.-L.; Zhou, H.-S.; Bian, J.-L.; Yang, Y.-R.; et al. NRF2 Promotes Breast Cancer Cell Proliferation and Metastasis by Increasing RhoA/ROCK Pathway Signal Transduction. *Oncotarget* 2016, *7*, 73593–73606. [[CrossRef](#)] [[PubMed](#)]
35. Bao, L.; Wu, J.; Dodson, M.; Rojo de la Vega, E.M.; Ning, Y.; Zhang, Z.; Yao, M.; Zhang, D.D.; Xu, C.; Yi, X.

- ABCF2, an Nrf2 Target Gene, Contributes to Cisplatin Resistance in Ovarian Cancer Cells. *Mol. Carcinog.* 2017, 56, 1543–1553. [[CrossRef](#)]
36. Wong, T.F.; Yoshinaga, K.; Monma, Y.; Ito, K.; Niikura, H.; Nagase, S.; Yamamoto, M.; Yaegashi, N. Association of Keap1 and Nrf2 Genetic Mutations and Polymorphisms with Endometrioid Endometrial Adenocarcinoma Survival. *Int. J. Gynecol. Cancer* 2011, 21, 1428–1435. [[CrossRef](#)] [[PubMed](#)]
  37. Shibata, T.; Kokubu, A.; Gotoh, M.; Ojima, H.; Ohta, T.; Yamamoto, M.; Hirohashi, S. Genetic Alteration of Keap1 Confers Constitutive Nrf2 Activation and Resistance to Chemotherapy in Gallbladder Cancer. *Gastroenterology* 2008, 135, 1358–1368. e4. [[CrossRef](#)]
  38. Nioi, P.; Nguyen, T. A Mutation of Keap1 Found in Breast Cancer Impairs Its Ability to Repress Nrf2 Activity. *Biochem. Biophys. Res. Commun.* 2007, 362, 816–821. [[CrossRef](#)]
  39. Sjöblom, T.; Jones, S.; Wood, L.D.; Parsons, D.W.; Lin, J.; Barber, T.D.; Mandelker, D.; Leary, R.J.; Ptak, J.; Silliman, N.; et al. The Consensus Coding Sequences of Human Breast and Colorectal Cancers. *Science* 2006, 314, 268–274. [[CrossRef](#)] [[PubMed](#)]
  40. Chu, X.-Y.; Li, Z.-J.; Zheng, Z.-W.; Tao, Y.-L.; Zou, F.-X.; Yang, X.-F. KEAP1/NRF2 Signaling Pathway Mutations in Cervical Cancer. *Eur. Rev. Med. Pharm. Sci.* 2018, 22, 4458–4466. [[CrossRef](#)]
  41. Konstantinopoulos, P.A.; Spentzos, D.; Fountzilas, E.; Francoeur, N.; Sanisetty, S.; Grammatikos, A.P.; Hecht, J.L.; Cannistra, S.A. Keap1 Mutations and Nrf2 Pathway Activation in Epithelial Ovarian Cancer. *Cancer Res.* 2011, 71, 5081–5089. [[CrossRef](#)]
  42. Martinez, V.D.; Vucic, E.A.; Thu, K.L.; Hubaux, R.; Enfield, K.S.S.; Pikor, L.A.; Becker-Santos, D.D.; Brown, C.J.; Lam, S.; Lam, W.L. Unique Somatic and Malignant Expression Patterns Implicate PIWI-Interacting RNAs in Cancer-Type Specific Biology. *Sci. Rep.* 2015, 5. [[CrossRef](#)]
  43. Yamamoto, S.; Inoue, J.; Kawano, T.; Kozaki, K.; Omura, K.; Inazawa, J. The Impact of MiRNA-Based Molecular Diagnostics and Treatment of NRF2-Stabilized Tumors. *Mol. Cancer Res.* 2014, 12, 58–68. [[CrossRef](#)] [[PubMed](#)]
  44. Jiao, N.; Baker, S.S.; Nugent, C.A.; Tsompana, M.; Cai, L.; Wang, Y.; Buck, M.J.; Genco, R.J.; Baker, R.D.; Zhu, R.; et al. Gut Microbiome May Contribute to Insulin Resistance and Systemic Inflammation in Obese Rodents: A Meta-Analysis. *Physiol. Genom.* 2018, 50, 244–254. [[CrossRef](#)]
  45. Zhang, Z.; Bai, L.; Guan, M.; Zhou, X.; Liang, X.; Lv, Y.; Yi, H.; Zhou, H.; Liu, T.; Gong, P.; et al. Potential probiotics *Lactobacillus casei* K11 combined with plant extracts reduce markers of type 2 diabetes mellitus in mice. *J. Appl. Microbiol.* 2021. [[CrossRef](#)]
  46. Bamberger, C.; Rossmeier, A.; Lechner, K.; Wu, L.; Waldmann, E.; Fischer, S.; Stark, R.G.; Altenhofer, J.; Henze, K.; Parhofer, K.G. A Walnut-Enriched Diet Affects Gut Microbiome in Healthy Caucasian Subjects: A Randomized, Controlled Trial. *Nutrients* 2018, 10, 244. [[CrossRef](#)] [[PubMed](#)]
  47. Gomez-Arango, L.F.; Barrett, H.L.; Wilkinson, S.A.; Callaway, L.K.; McIntyre, H.D.; Morrison, M.; Dekker Nitert, M. Low Dietary Fiber Intake Increases Collinsella Abundance in the Gut Microbiota of Overweight and Obese Pregnant Women. *Gut Microbes* 2018, 9, 189–201. [[CrossRef](#)] [[PubMed](#)]
  48. Ojo, O.; Feng, Q.-Q.; Ojo, O.O.; Wang, X.-H. The Role of Dietary Fibre in Modulating Gut Microbiota Dysbiosis in Patients with Type 2 Diabetes: A Systematic Review and Meta-Analysis of Randomised Controlled Trials. *Nutrients* 2017, 12, 3239. [[CrossRef](#)] [[PubMed](#)]
  49. den Besten, G.; van Eunen, K.; Groen, A.K.; Venema, K.; Reijngoud, D.-J.; Bakker, B.M. The Role of Short-Chain Fatty Acids in the Interplay between Diet, Gut Microbiota, and Host Energy Metabolism. *J. Lipid Res.* 2013, 54, 2325–2340. [[CrossRef](#)]
  50. Heimann, E.; Nyman, M.; Pålbrink, A.-K.; Lindkvist-Petersson, K.; Degerman, E. Branched Short-Chain Fatty Acids Modulate Glucose and Lipid Metabolism in Primary Adipocytes. *Adipocyte* 2016, 5, 359–368. [[CrossRef](#)]
  51. Matijašić, M.; Meštrović, T.; Perić, M.; Čipčić, Paljetak, H.; Panek, M.; Vranešić, Bender, D.; Ljubas Kelečić, D.; Krznarić, Ž.; Verbanac, D. Modulating Composition and Metabolic Activity of the Gut Microbiota in IBD Patients. *Int. J. Mol. Sci.* 2016, 17, 578. [[CrossRef](#)]
  52. Meštrović, T.; Matijašić, M.; Perić, M.; Čipčić, Paljetak, H.; Barešić, A.; Verbanac, D. The Role of Gut, Vaginal, and Urinary Microbiome in Urinary Tract Infections: From Bench to Bedside. *Diagnostics* 2017, 11, 7. [[CrossRef](#)]
  53. Franks, S.; Kiddy, D.S.; Hamilton-Fairley, D.; Bush, A.; Sharp, P.S.; Reed, M.J. The Role of Nutrition and Insulin in the Regulation of Sex Hormone Binding Globulin. *J. Steroid Biochem. Mol. Biol.* 1991, 39, 835–838. [[CrossRef](#)]
  54. Tymchuk, C.N.; Tessler, S.B.; Barnard, R.J. Changes in Sex Hormone-Binding Globulin, Insulin, and Serum Lipids in Post-menopausal Women on a Low-Fat, High-Fiber Diet Combined with Exercise. *Nutr. Cancer* 2000, 38, 158–162. [[CrossRef](#)] [[PubMed](#)]
  55. Gann, P.H.; Chatterton, R.T.; Gapstur, S.M.; Liu, K.; Garside, D.; Giovanazzi, S.; Thedford, K.; Van Horn, L. The Effects of a Low-Fat/High-Fiber Diet on Sex Hormone Levels and Menstrual Cycling in Premenopausal Women: A 12-Month Randomized Trial (the Diet and Hormone Study). *Cancer* 2003, 98, 1870–1879. [[CrossRef](#)]
  56. Moran, L.J.; Noakes, M.; Clifton, P.M.; Tomlinson, L.; Galletly, C.; Norman, R.J. Dietary Composition in Restoring Reproductive and Metabolic Physiology in

- Overweight Women with Polycystic Ovary Syndrome. *J. Clin. Endocrinol. Metab.* 2003, 88, 812–819. [[CrossRef](#)]
57. Szczuko, M.; Zapałowska-Chwyc', M.; Drozd, A.; Maciejewska, D.; Starczewski, A.; Wysokiński, P.; Stachowska, E. Changes in the IGF-1 and TNF- $\alpha$  Synthesis Pathways before and after Three-Month Reduction Diet with Low Glycemic Index in Women with PCOS. *Ginekol. Pol.* 2018, 89, 295–303. [[CrossRef](#)]
  58. Kazemi, M.; Hadi, A.; Pierson, R.A.; Lujan, M.E.; Zello, G.A.; Chilibeck, P.D. Effects of Dietary Glycemic Index and Glycemic Load on Cardiometabolic and Reproductive Profiles in Women with Polycystic Ovary Syndrome: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Adv. Nutr.* 2018, 12, 161–178. [[CrossRef](#)]
  59. Wang, T.; Sha, L.; Li, Y.; Zhu, L.; Wang, Z.; Li, K.; Lu, H.; Bao, T.; Guo, L.; Zhang, X.; et al. Dietary  $\alpha$ -Linolenic Acid-Rich Flaxseed Oil Exerts Beneficial Effects on Polycystic Ovary Syndrome Through Sex Steroid Hormones-Microbiota-Inflammation Axis in Rats. *Front. Endocrinol.* 2018, 11, 284. [[CrossRef](#)]
  60. Barber, T.M.; Kabisch, S.; Pfeiffer, A.F.H.; Weickert, M.O. The Health Benefits of Dietary Fibre. *Nutrients* 2018, 12, 3209. [[CrossRef](#)]
  61. Hoover, S.E.; Gower, B.A.; Cedillo, Y.E.; Chandler-Laney, P.C.; Deemer, S.E.; Goss, A.M. Changes in Ghrelin and Glucagon Following a Low Glycemic Load Diet in Women with PCOS. *J. Clin. Endocrinol. Metab.* 2018. [[CrossRef](#)]
  62. Akintayo, C.O.; Johnson, A.D.; Badejogbin, O.C.; Olaniyi, K.S.; Oniyide, A.A.; Ajadi, I.O.; Ojewale, A.O.; Adeyomoye, O.I.; Kayode, A.B. High Fructose-Enriched Diet Synergistically Exacerbates Endocrine but Not Metabolic Changes in Letrozole-Induced Polycystic Ovarian Syndrome in Wistar Rats. *Heliyon* 2018, 7, e05890. [[CrossRef](#)]
  63. Shang, Y.; Zhou, H.; Hu, M.; Feng, H. Effect of Diet on Insulin Resistance in Polycystic Ovary Syndrome. *J. Clin. Endocrinol. Metab.* 2018, 105. [[CrossRef](#)]
  64. Porchia, L.M.; Hernandez-Garcia, S.C.; Gonzalez-Mejia, M.E.; López-Bayghen, E. Diets with Lower Carbohydrate Concentrations Improve Insulin Sensitivity in Women with Polycystic Ovary Syndrome: A Meta-Analysis. *Eur. J. Obs. Gynecol. Reprod. Biol.* 2018, 248, 110–117. [[CrossRef](#)] [[PubMed](#)]
  65. Shisheghar, F.; Mirmiran, P.; Rahmati, M.; Tohidi, M.; Ramezani Tehrani, F. Does a Restricted Energy Low Glycemic Index Diet Have a Different Effect on Overweight Women with or without Polycystic Ovary Syndrome? *BMC Endocr. Disord.* 2018, 19, 93. [[CrossRef](#)] [[PubMed](#)]
  66. Paoli, A.; Mancin, L.; Giacona, M.C.; Bianco, A.; Caprio, M. Effects of a Ketogenic Diet in Overweight Women with Polycystic Ovary Syndrome. *J. Transl. Med.* 2018, 18, 104. [[CrossRef](#)] [[PubMed](#)]
  67. Fonseka, S.; Subhani, B.; Wijeyaratne, C.N.; Gawarammana, I.B.; Kalupahana, N.S.; Ratnatunga, N.; Rosairo, S.; Vithane, K.P. Association between Visceral Adiposity Index, Hirsutism and Cardiometabolic Risk Factors in Women with Polycystic Ovarian Syndrome: A Cross-Sectional Study. *Ceylon Med. J.* 2018, 64, 111–117. [[CrossRef](#)]
  68. Marson, E.C.; Delevatti, R.S.; Prado, A.K.G.; Netto, N.; Krue, L.F.M. Effects of Aerobic, Resistance, and Combined Exercise Training on Insulin Resistance Markers in Overweight or Obese Children and Adolescents: A Systematic Review and Meta-Analysis. *Prev. Med.* 2016, 93, 211–218. [[CrossRef](#)]
  69. Patten, R.K.; Boyle, R.A.; Moholdt, T.; Kiel, I.; Hopkins, W.G.; Harrison, C.L.; Stepto, N.K. Exercise Interventions in Polycystic Ovary Syndrome: A Systematic Review and Meta-Analysis. *Front. Physiol.* 2018, 11, 606. [[CrossRef](#)]
  70. Santos, I.K.D.; Nunes, F.A.S.d.S.; Queiros, V.S.; Cobucci, R.N.; Dantas, P.B.; Soares, G.M.; Cabral, B.G.d.A.T.; Maranhão, T.M.d.O.; Dantas, P.M.S. Effect of High-Intensity Interval Training on Metabolic Parameters in Women with Polycystic Ovary Syndrome: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *PLoS ONE* 2018, 16, e0245023. [[CrossRef](#)]
  71. Shele, G.; Genkil, J.; Speelman, D. A Systematic Review of the Effects of Exercise on Hormones in Women with Polycystic Ovary Syndrome. *J. Funct. Morphol. Kinesiol.* 2018, 5, 35. [[CrossRef](#)]
  72. Yang, Y.; Deng, H.; Li, T.; Xia, M.; Liu, C.; Bu, X.-Q.; Li, H.; Fu, L.-J.; Zhong, Z.-H. The Mental Health of Chinese Women with Polycystic Ovary Syndrome Is Related to Sleep Disorders, Not Disease Status. *J. Affect. Disord.* 2018, 282, 51–57. [[CrossRef](#)]
  73. Leproult, R.; Van Cauter, E. Role of Sleep and Sleep Loss in Hormonal Release and Metabolism. *Endocr. Dev.* 2010, 17, 11–21. [[CrossRef](#)]
  74. Donga, E.; Romijn, J.A. Sleep Characteristics and Insulin Sensitivity in Humans. *Handb. Clin. Neurol.* 2014, 124, 107–114. [[CrossRef](#)]
  75. Reutrakul, S.; Van Cauter, E. Sleep Influences on Obesity, Insulin Resistance, and Risk of Type 2 Diabetes. *Metabolism* 2018, 84, 56–66. [[CrossRef](#)]
  76. Poroyko, V.A.; Carreras, A.; Khalyfa, A.; Khalyfa, A.A.; Leone, V.; Peris, E.; Almendros, I.; Gileles-Hillel, A.; Qiao, Z.; Hubert, N.; et al. Chronic Sleep Disruption Alters Gut Microbiota, Induces Systemic and Adipose Tissue Inflammation and Insulin Resistance in Mice. *Sci. Rep.* 2016, 6, 35405. [[CrossRef](#)]
  77. Mojaverrostami, S.; Asghari, N.; Khamisabadi, M.; Heidari Khoei, H. The Role of Melatonin in Polycystic Ovary Syndrome: A Review. *Int. J. Reprod. Biomed.* 2018, 17, 865–882. [[CrossRef](#)] [[PubMed](#)]
  78. Szczuko, M.; Hawryłkiewicz, V.; Kikut, J.; Drozd, A. The Implications of Vitamin Content in the Plasma in Reference to the Parameters of Carbohydrate Metabolism



- and Hormone and Lipid Profiles in PCOS. *J. Steroid Biochem. Mol. Biol.* 2018, 198, 105570. [[CrossRef](#)]
79. Szczuko, M.; Szydłowska, I.; Nawrocka-Rutkowska, J. A Properly Balanced Reduction Diet and/or Supplementation Solve the Problem with the Deficiency of These Vitamins Soluble in Water in Patients with PCOS. *Nutrients* 2018, 13, 746. [[CrossRef](#)] [[PubMed](#)]
  80. Suzuki, H.; Kunisawa, J. Vitamin-Mediated Immune Regulation in the Development of Inflammatory Diseases. *Endocr. Metab. Immune Disord. Drug Targets* 2015, 15, 212–215. [[CrossRef](#)] [[PubMed](#)]
  81. Wanders, D.; Graff, E.C.; White, B.D.; Judd, R.L. Niacin Increases Adiponectin and Decreases Adipose Tissue Inflammation in High Fat Diet-Fed Mice. *PLoS ONE* 2013, 8, e71285. [[CrossRef](#)]
  82. Esmailzadeh, S.; Gholinezhad-Chari, M.; Ghadimi, R. The Effect of Metformin Treatment on the Serum Levels of Homocysteine, Folic Acid, and Vitamin B12 in Patients with Polycystic Ovary Syndrome. *J. Hum. Reprod. Sci.* 2017, 10, 95–101. [[CrossRef](#)] [[PubMed](#)]
  83. DiNicolantonio, J.J.; Liu, J.; O'Keefe, J.H. Thiamine and Cardiovascular Disease: A Literature Review. *Prog. Cardiovasc. Dis.* 2018, 61, 27–32. [[CrossRef](#)] [[PubMed](#)]
  84. Eshak, E.S.; Arafa, A.E. Thiamine Deficiency and Cardiovascular Disorders. *Nutr. Metab. Cardiovasc. Dis.* 2018, 28, 965–972. [[CrossRef](#)] [[PubMed](#)]
  85. Taghizadeh, S.; Izadi, A.; Shirazi, S.; Parizad, M.; Pourghassem Gargari, B. The Effect of Coenzyme Q10 Supplementation on Inflammatory and Endothelial Dysfunction Markers in Overweight/Obese Polycystic Ovary Syndrome Patients. *Gynecol. Endocrinol.* 2018, 37, 26–30. [[CrossRef](#)]
  86. Teegarden, D.; Donkin, S.S. Vitamin D: Emerging New Roles in Insulin Sensitivity. *Nutr. Res. Rev.* 2009, 22, 82–92. [[CrossRef](#)] [[PubMed](#)]
  87. He, C.; Lin, Z.; Robb, S.W.; Ezeamama, A.E. Serum Vitamin D Levels and Polycystic Ovary Syndrome: A Systematic Review and Meta-Analysis. *Nutrients* 2015, 7, 4555–4577. [[CrossRef](#)]
  88. Wehr, E.; Pieber, T.R.; Obermayer-Pietsch, B. Effect of Vitamin D3 Treatment on Glucose Metabolism and Menstrual Frequency in Polycystic Ovary Syndrome Women: A Pilot Study. *J. Endocrinol. Investig.* 2011, 34, 757–763. [[CrossRef](#)]
  89. Maktabi, M.; Jamilian, M.; Asemi, Z. Magnesium-Zinc-Calcium-Vitamin D Co-Supplementation Improves Hormonal Profiles, Biomarkers of Inflammation and Oxidative Stress in Women with Polycystic Ovary Syndrome: A Randomized, Double-Blind, Placebo-Controlled Trial. *Biol. Trace Elem. Res.* 2018, 182, 21–28. [[CrossRef](#)]
  90. Jamilian, M.; Samimi, M.; Mirhosseini, N.; Afshar Ebrahimi, F.; Aghadavod, E.; Talaei, R.; Jafarnejad, S.; Hashemi Dizaji, S.; Asemi, Z. The Influences of Vitamin D and Omega-3 Co-Supplementation on Clinical, Metabolic and Genetic Parameters in Women with Polycystic Ovary Syndrome. *J. Affect. Disord.* 2018, 238, 32–38. [[CrossRef](#)]
  91. Formoso, C.; Stracquadanio, M.; Ciotta, L. Myo-Inositol vs. D-Chiro Inositol in PCOS Treatment. *Minerva Ginecol.* 2015, 67, 321–325. [[PubMed](#)]
  92. Fruzzetti, F.; Perini, D.; Russo, M.; Bucci, F.; Gadducci, A. Comparison of Two Insulin Sensitizers, Metformin and Myo-Inositol, in Women with Polycystic Ovary Syndrome (PCOS). *Gynecol. Endocrinol.* 2017, 33, 39–42. [[CrossRef](#)] [[PubMed](#)]
  93. Saleem, F.; Rizvi, S.W. New Therapeutic Approaches in Obesity and Metabolic Syndrome Associated with Polycystic Ovary Syndrome. *Cureus* 2017, 9, e1844. [[CrossRef](#)] [[PubMed](#)]
  94. Genazzani, A.D.; Santagni, S.; Ricchieri, F.; Campedelli, A.; Rattighieri, E.; Chierchia, E.; Marini, G.; Despini, G.; Prati, A.; Simoncini, T. Myo-Inositol Modulates Insulin and Luteinizing Hormone Secretion in Normal Weight Patients with Polycystic Ovary Syndrome. *J. Obs. Gynaecol. Res.* 2014, 40, 1353–1360. [[CrossRef](#)] [[PubMed](#)]
  95. Facchinetti, F.; Bizzarri, M.; Benvenega, S.; D'Anna, R.; Lanzone, A.; Soulage, C.; Di Renzo, G.C.; Hod, M.; Cavalli, P.; Chiu, T.T.; et al. Results from the International Consensus Conference on Myo-Inositol and d-Chiro-Inositol in Obstetrics and Gynecology: The Link between Metabolic Syndrome and PCOS. *Eur. J. Obs. Gynecol. Reprod. Biol.* 2015, 195, 72–76. [[CrossRef](#)] [[PubMed](#)]
  96. Unfer, V.; Nestler, J.E.; Kamenov, Z.A.; Prapas, N.; Facchinetti, F. Effects of Inositol(s) in Women with PCOS: A Systematic Review of Randomized Controlled Trials. *Int. J. Endocrinol.* 2016, 2016, 1849162. [[CrossRef](#)]
  97. Li, Y.; Ma, H.; Zhang, Y.; Kuang, H.; Ng, E.H.Y.; Hou, L.; Wu, X. Effect of Berberine on Insulin Resistance in Women with Polycystic Ovary Syndrome: Study Protocol for a Randomized Multicenter Controlled Trial. *Trials* 2013, 14, 226. [[CrossRef](#)]
  98. Rondanelli, M.; Infantino, V.; Riva, A.; Petrangolini, G.; Faliva, M.A.; Peroni, G.; Naso, M.; Nichetti, M.; Spadaccini, D.; Gasparri, C.; et al. Polycystic Ovary Syndrome Management: A Review of the Possible Amazing Role of Berberine. *Arch. Gynecol. Obs.* 2018, 301, 53–60. [[CrossRef](#)]
  99. Xiang, D.; Lu, J.; Wei, C.; Cai, X.; Wang, Y.; Liang, Y.; Xu, M.; Wang, Z.; Liu, M.; Wang, M.; et al. Berberine Ameliorates Prenatal Dihydrotestosterone Exposure-Induced Autism-Like Behavior by Suppression of Androgen Receptor. *Front. Cell Neurosci.* 2018, 14. [[CrossRef](#)] [[PubMed](#)]
  100. Bertuccioli, A.; Moricoli, S.; Amatori, S.; Rocchi, M.B.L.; Vici, G.; Sisti, D. Berberine and Dyslipidemia: Different Applications and Biopharmaceutical Formulations



- Without Statin-Like Molecules-A Meta-Analysis. *J. Med. Food* 2018 23, 101–113. [[CrossRef](#)] [[PubMed](#)]
101. Wei, W.; Zhao, H.; Wang, A.; Sui, M.; Liang, K.; Deng, H.; Ma, Y.; Zhang, Y.; Zhang, H.; Guan, Y. A Clinical Study on the Short-Term Effect of Berberine in Comparison to Metformin on the Metabolic Characteristics of Women with Polycystic Ovary Syndrome. *Eur. J. Endocrinol.* 2012, 166, 99–105. [[CrossRef](#)] [[PubMed](#)]
  102. Kuang, H.; Duan, Y.; Li, D.; Xu, Y.; Ai, W.; Li, W.; Wang, Y.; Liu, S.; Li, M.; Liu, X.; et al. The Role of Serum Inflammatory Cytokines and Berberine in the Insulin Signaling Pathway among Women with Polycystic Ovary Syndrome. *PLoS ONE* 2018 15. [[CrossRef](#)] [[PubMed](#)]
  103. Lucidi, R.S.; Thyer, A.C.; Easton, C.A.; Holden, A.E.C.; Schenken, R.S.; Brzyski, R.G. Effect of Chromium Supplementation on Insulin Resistance and Ovarian and Menstrual Cyclicity in Women with Polycystic Ovary Syndrome. *Fertil. Steril.* 2005, 84, 1755–1757. [[CrossRef](#)] [[PubMed](#)]
  104. Tang, X.-L.; Sun, Z.; Gong, L. Chromium Supplementation in Women with Polycystic Ovary Syndrome: Systematic Review and Meta-Analysis. *J. Obs. Gynaecol. Res.* 2018, 44, 134–143. [[CrossRef](#)]
  105. Fazelian, S.; Rouhani, M.H.; Bank, S.S.; Amani, R. Chromium Supplementation and Polycystic Ovary Syndrome: A Systematic Review and Meta-Analysis. *J. Trace Elem. Med. Biol.* 2017, 42, 92–96. [[CrossRef](#)] [[PubMed](#)]
  106. Ashoush, S.; Abou-Gamrah, A.; Bayoumy, H.; Othman, N. Chromium Picolinate Reduces Insulin Resistance in Polycystic Ovary Syndrome: Randomized Controlled Trial. *J. Obs. Gynaecol. Res.* 2016, 42, 279–285. [[CrossRef](#)]
  107. Piotrowska, A.; Pilch, W.; Czerwin'ska-Ledwig, O.; Zuziak, R.; Siwek, A.; Wolak, M.; Nowak, G. The Possibilities of Using Chromium Salts as an Agent Supporting Treatment of Polycystic Ovary Syndrome. *Biol. Trace Elem. Res.* 2018, 192, 91–97. [[CrossRef](#)]
  108. Maxel, T.; Svendsen, P.F.; Smidt, K.; Lauridsen, J.K.; Brock, B.; Pedersen, S.B.; Rungby, J.; Larsen, A. Expression Patterns and Correlations with Metabolic Markers of Zinc Transporters ZIP14 and ZNT1 in Obesity and Polycystic Ovary Syndrome. *Front. Endocrinol.* 2017, 8. [[CrossRef](#)]
  109. Nasiadek, M.; Stragierowicz, J.; Klimczak, M.; Kilanowicz, A. The Role of Zinc in Selected Female Reproductive System Disorders. *Nutrients* 2018, 12, 2464. [[CrossRef](#)]
  110. Guler, I.; Himmetoglu, O.; Turp, A.; Erdem, A.; Erdem, M.; Onan, M.A.; Taskiran, C.; Taslipinar, M.Y.; Guner, H. Zinc and Homocysteine Levels in Polycystic Ovarian Syndrome Patients with Insulin Resistance. *Biol. Trace Elem. Res.* 2014, 158, 297–304. [[CrossRef](#)]
  111. Coskun, A.; Arikan, T.; Kilinc, M.; Arikan, D.C.; Ekerbiçer, H.Ç. Plasma Selenium Levels in Turkish Women with Polycystic Ovary Syndrome. *Eur. J. Obs. Gynecol. Reprod. Biol.* 2013, 168, 183–186. [[CrossRef](#)] [[PubMed](#)]
  112. Michael, P.J.; Stepanic', V.; Nadja, T.; Panek, M.; Verbanac, D. Mild Plant and Dietary Immunomodulators. *Nijkamp Parnham's Princ. Immunopharmacol.* 2018, 561–587. [[CrossRef](#)]
  113. Ma, X.; Weng, X.; Hu, X.; Wang, Q.; Tian, Y.; Ding, Y.; Zhang, C. Roles of Different N-3/n-6 PUFA Ratios in Ovarian Cell Development and Steroidogenesis in PCOS Rats. *Food. Funct.* 2018, 10, 7397–7406. [[CrossRef](#)]
  114. Popova, A.; Mihaylova, D. A Review of the Medicinal Plants in Bulgaria: Collection, Storage, And Extraction Techniques. *Asian J. Pharm. Clin. Res.* 2018, 28–35. [[CrossRef](#)]
  115. Ashkar, F.; Rezaei, S.; Salahshornezhad, S.; Vahid, F.; Gholamalizadeh, M.; Dahka, S.M.; Doaei, S. The Role of Medicinal Herbs in Treatment of Insulin Resistance in Patients with Polycystic Ovary Syndrome: A Literature Review. *Biomol. Concepts* 2018 11, 57–75. [[CrossRef](#)]
  116. Tehrani, H.G.; Allahdadian, M.; Zarre, F.; Ranjbar, H.; Allahdadian, F. Effect of Green Tea on Metabolic and Hormonal Aspect of Polycystic Ovarian Syndrome in Overweight and Obese Women Suffering from Polycystic Ovarian Syndrome: A Clinical Trial. *J. Educ. Health Promot.* 2017, 6, 36. [[CrossRef](#)]
  117. Haj-Husein, I.; Tukan, S.; Alkazaleh, F. The Effect of Marjoram (*Origanum Majorana*) Tea on the Hormonal Profile of Women with Polycystic Ovary Syndrome: A Randomised Controlled Pilot Study. *J. Hum. Nutr. Diet.* 2016, 29, 105–111. [[CrossRef](#)]
  118. Rababa'h, A.M.; Matani, B.R.; Ababneh, M.A. The Ameliorative Effects of Marjoram in Dehydroepiandrosterone Induced Polycystic Ovary Syndrome in Rats. *Life Sci.* 2018, 261, 118353. [[CrossRef](#)] [[PubMed](#)]
  119. Grant, P. Spearmint Herbal Tea Has Significant Anti-Androgen Effects in Polycystic Ovarian Syndrome. A Randomized Controlled Trial. *Phytother. Res.* 2010, 24, 186–188. [[CrossRef](#)]
  120. Sadeghi Ataabadi, M.; Alaei, S.; Bagheri, M.J.; Bahmanpoor, S. Role of Essential Oil of *Mentha Spicata* (Spearmint) in Addressing Reverse Hormonal and Folliculogenesis Disturbances in a Polycystic Ovarian Syndrome in a Rat Model. *Adv. Pharm. Bull.* 2017, 7, 651–654. [[CrossRef](#)]
  121. Sabbadin, C.; Bordin, L.; Donà, G.; Manso, J.; Avruscio, G.; Armanini, D. Licorice: From Pseudohyperaldosteronism to Therapeutic Uses. *Front. Endocrinol.* 2018, 10, 484. [[CrossRef](#)] [[PubMed](#)]
  122. Arentz, S.; Abbott, J.A.; Smith, C.A.; Bensoussan, A. Herbal Medicine for the Management of Polycystic Ovary Syndrome (PCOS) and Associated Oligo/Amenorrhoea and Hyperandrogenism; a Review of the Laboratory Evidence for Effects with Corroborative Clinical

- Findings. *BMC Complement. Altern Med.* 2014, 14, 511. [[CrossRef](#)] [[PubMed](#)]
123. Adamczak, M.; Wiecek, A. Food Products That May Cause an Increase in Blood Pressure. *Curr. Hypertens. Rep.* 2018, 22, 2. [[CrossRef](#)] [[PubMed](#)]
  124. Dhariwala, M.Y.; Ravikumar, P. An Overview of Herbal Alternatives in Androgenetic Alopecia. *J. Cosmet Derm.* 2018 18, 966–975. [[CrossRef](#)] [[PubMed](#)]
  125. Kakadia, N.; Patel, P.; Deshpande, S.; Shah, G. Effect of Vitex Negundo L. Seeds in Letrozole Induced Polycystic Ovarian Syndrome. *J. Tradit Complement. Med.* 2018, 9, 336–345. [[CrossRef](#)]
  126. Mehraban, M.; Jelodar, G.; Rahmanifar, F. A Combination of Spearmint and Flaxseed Extract Improved Endocrine and Histomorphology of Ovary in Experimental PCOS. *J. Ovarian Res.* 2018, 13, 32. [[CrossRef](#)]
  127. Brooks, J.D.; Thompson, L.U. Mammalian Lignans and Genistein Decrease the Activities of Aromatase and 17 $\beta$ -Hydroxysteroid Dehydrogenase in MCF-7 Cells. *J. Steroid Biochem. Mol. Biol.* 2005, 94, 461–467. [[CrossRef](#)]
  128. Heshmati, J.; Moini, A.; Sepidarkish, M.; Morvaridzadeh, M.; Salehi, M.; Palmowski, A.; Mojtaheidi, M.F.; Shidfar, F. Effects of Curcumin Supplementation on Blood Glucose, Insulin Resistance and Androgens in Patients with Polycystic Ovary Syndrome: A Randomized Double-Blind Placebo-Controlled Clinical Trial. *Phytomedicine* 2018, 80, 153395. [[CrossRef](#)]
  129. Marmitt, D.J.; Shahrajabian, M.H.; Goettert, M.I.; Rempel, C. Clinical Trials with Plants in Diabetes Mellitus Therapy: A Systematic Review. *Expert Rev. Clin. Pharm.* 2018, 1–13. [[CrossRef](#)]
  130. Heshmati, J.; Golab, F.; Morvaridzadeh, M.; Potter, E.; Akbari-Fakhrabadi, M.; Farsi, F.; Tanbakooei, S.; Shidfar, F. The Effects of Curcumin Supplementation on Oxidative Stress, Sirtuin-1 and Peroxisome Proliferator Activated Receptor  $\gamma$  Coactivator 1 $\alpha$  Gene Expression in Polycystic Ovarian Syndrome (PCOS) Patients: A Randomized Placebo-Controlled Clinical Trial. *Diabetes Metab. Syndr.* 2018 14, 77–82. [[CrossRef](#)]
  131. Yuandani, I.J.; Rohani, A.S.; Sumantri, I.B. Immunomodulatory Effects and Mechanisms of Curcuma Species and Their Bioactive Compounds: A Review. *Front. Pharm.* 2018, 12, 643119. [[CrossRef](#)]
  132. Chowdhury, I.; Banerjee, S.; Driss, A.; Xu, W.; Mehrabi, S.; Nezhat, C.; Sidell, N.; Taylor, R.N.; Thompson, W.E. Curcumin Attenuates Proangiogenic and Proinflammatory Factors in Human Eutopic Endometrial Stromal Cells through the NF-KB Signaling Pathway. *J. Cell Physiol* 2018, 234, 6298–6312. [[CrossRef](#)]
  133. Işler, S.C.; Demircan, S.; Çakar, S.; Çebi, Z.; Keskin, C.; Soluk, M.; Yüzbaşıoğlu, E. Effects of Folk Medicinal Plant Extract Ankaferd Blood Stopper® on Early Bone Healing. *J. Appl. Oral Sci.* 2010, 18, 409–414. [[CrossRef](#)] [[PubMed](#)]
  134. Ziaei, R.; Foshati, S.; Hadi, A.; Kermani, M.A.H.; Ghavami, A.; Clark, C.C.T.; Tarrahi, M.J. The Effect of Nettle (*Urtica Dioica*) Supplementation on the Glycemic Control of Patients with Type 2 Diabetes Mellitus: A Systematic Review and Meta-Analysis. *Phytother. Res.* 2018, 34, 282–294. [[CrossRef](#)] [[PubMed](#)]
  135. Sarma Katak, M.; Murugamani, V.; Rajkumari, A.; Singh Mehra, P.; Awasthi, D.; Shankar Yadav, R. Antioxidant, Hepatoprotective, and Anthelmintic Activities of Methanol Extract of *Urtica Dioica* L. Leaves. *Pharm. Crop.* 2012, 3, 38–46. [[CrossRef](#)]
  136. Ferro, D.; Baratta, F.; Pastori, D.; Cocomello, N.; Colantoni, A.; Angelico, F.; Del Ben, M. New Insights into the Pathogenesis of Non-Alcoholic Fatty Liver Disease: Gut-Derived Lipopolysaccharides and Oxidative Stress. *Nutrients* 2018, 12, 2762. [[CrossRef](#)]
  137. Wat, E.; Wang, Y.; Chan, K.; Law, H.W.; Koon, C.M.; Lau, K.M.; Leung, P.C.; Yan, C.; Lau, C.B.S. An in Vitro and in Vivo Study of a 4-Herb Formula on the Management of Diet-Induced Metabolic Syndrome. *Phytomedicine* 2018, 42, 112–125. [[CrossRef](#)]
  138. MacDonald-Ramos, K.; Michán, L.; Martínez-Ibarra, A.; Cerbón, M. Silymarin Is an Ally against Insulin Resistance: A Review. *Ann. Hepatol.* 2018, 23, 100255. [[CrossRef](#)]
  139. Oppedisano, F.; Muscoli, C.; Musolino, V.; Carresi, C.; Macri, R.; Giancotta, C.; Bosco, F.; Maiuolo, J.; Scarano, F.; Paone, S.; et al. The Protective Effect of *Cynara Cardunculus* Extract in Diet-Induced NAFLD: Involvement of OCTN1 and OCTN2 Transporter Subfamily. *Nutrients* 2018, 12, 1435. [[CrossRef](#)]
  140. Zhao, Y.-M.; Wang, C.; Zhang, R.; Hou, X.-J.; Zhao, F.; Zhang, J.-J.; Wang, C. [Study on literature of artichoke and properties of traditional Chinese medicine]. *Zhongguo Zhong Yao Za Zhi* 2018, 45, 3481–3488. [[CrossRef](#)] [[PubMed](#)]
  141. Park, S.; Kim, D.S.; Wu, X.; J Yi, Q. Mulberry and Dandelion Water Extracts Prevent Alcohol-Induced Steatosis with Alleviating Gut Microbiome Dysbiosis. *Exp. Biol. Med.* 2018, 243, 882–894. [[CrossRef](#)] [[PubMed](#)]
  142. Azizi, N.; Amini, M.R.; Djafarian, K.; Shab-Bidar, S. The Effects of *Nigella Sativa* Supplementation on Liver Enzyme Levels: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Clin. Nutr. Res.* 2018, 10, 72–82. [[CrossRef](#)] [[PubMed](#)]

**Creative Commons (CC) License**

This article is an open-access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY 4.0) license. This license permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.