

## A contribution to the study of *Helotiales* and *Rhytismatales* in Turkey

MAKBULE ERDOĞDU<sup>1\*</sup>, GÖKHAN DOĞAN<sup>1</sup>,  
ELŞAD HÜSEYİN<sup>1</sup> & ZEKIYE SULUDERE<sup>2</sup>

<sup>1</sup> Ahi Evran University, Faculty of Science and Literature, Department of Biology,  
Bağbaşı, Kırşehir, Turkey

<sup>2</sup> Gazi University, Faculty of Science, Department of Biology,  
Teknikokullar, Ankara, Turkey

\* CORRESPONDENCE TO: [merdogdu@ahievran.edu.tr](mailto:merdogdu@ahievran.edu.tr)

**ABSTRACT**—*Naemacyclus fimbriatus*, *Lophodermium juniperinum*, and *Marssonina daphnes* have recently been discovered in Turkey. This is the first record of *Naemacyclus* from Turkey. Morphological data obtained by light and scanning electron microscopy of these fungi are presented.

**KEY WORDS**—acervular anamorph, *Ascomycota*, new host, new records, SEM

### Introduction

*Rhytismatales* are an order of endophytic, parasitic, or saprotrophic fungi in *Leotiomyces* (*Ascomycota*), the inoperculate discomycetes. Especially common on conifers, grasses and members of *Ericaceae*, they are also found on other vascular plants. The species disperse by ascospores and at least in temperate regions usually infect their hosts in spring/summer to develop fruiting bodies the next year on dead material (Lantz et al. 2011). The order includes plant parasitic fungi causing serious needle cast, such as *Lophodermium seditiosum* Minter et al. on *Pinus sylvestris* (Minter 1981b)., *Rhytismatalean* fungi the members of *Rhytismatales* are poorly known in Turkey and have not been yet intensively studied.

Within *Leotiomyces*, *Helotiales* represents the largest order of inoperculate discomycetes—an ecologically and morphologically highly diverse group of

ascomycetes that also includes lichen-inhabiting (lichenicolous) species (Suija et al. 2015). At present the order comprises c. 4000 species of saprophytes, mycorrhizal fungi, root endophytes, and plant and fungal (including lichens) parasites (Schoch et al. 2009).

## Materials & methods

Plant specimens infected with microfungi were collected from Erciyes Mountain in Kayseri province of Turkey. The host specimens were prepared following conventional herbarium techniques. Host plants were identified using the FLORA OF TURKEY AND EAST AEGEAN ISLANDS (Davis 1965–85). Thin fungal sections prepared from host tissue were examined under a Leica DME light microscope and measured from mounts in 5% KOH or tap water. Close-up photographs of infected host surface were done via Leica EZ4D stereomicroscope. The fungi were identified using relevant literature (for *Naemacyclus* – DiCosmo et al. 1984, Hou et al. 2006; for *Lophodermium* – Dennis 1981, Ellis & Ellis 1987; for *Marssonina* – Grove 1937, Ellis & Ellis 1987, Ignatavičiūtė & Treigienė 1998). All examined specimens were deposited in the Mycology Laboratory of Ahi Evran University, Department of Biology, Kırşehir, Turkey (C), under Gökhan Doğan (GD) collection numbers.

For scanning electron microscopy (SEM), 8–10 mm square pieces of infected leaf or cone scale were mounted on aluminium stubs with double-sided adhesive tape, coated with gold using Polaron SC 502 Sputter Coater, and examined using a Jeol JSM 6060 scanning electron microscope operated at 5–10 kV in the Electron Microscopy Unit, Gazi University (Turkey).

## Taxonomy

*Naemacyclus fimbriatus* (Schwein.) DiCosmo, Peredo & Minter,

Eur. J. For. Path. 13(4): 207 (1983)

PL. 1

APOTHECIA scattered, immersed to erumpent, sessile, circular to subcircular, 300–430 × 230–285 µm, dark brown to black. ASCI cylindrical, short-stalked, thin-walled, 85–95 × 8.5–9.5 µm, rostrate at the apex, without circumapical thickening, discharging spores through a small apical pore, 8-spored. ASCOSPORES fasciculate, filiform, (50–)54–80 × 2–2.2 µm, rounded at both ends, hyaline, (2–)5-septate, rarely aseptate, with mucous sheath. PARAPHYSES filiform, unbranched, septate, 90–105 × 1 µm, hyaline, covered by a thin mucous sheath.

SPECIMEN EXAMINED—TURKEY, KAYSERİ, Erciyes mountain, Turkish World Forest, 38°36'12"N 35°30'56"E, 1850–1900 m asl., on fallen female cone scales of *Pinus nigra* L. (*Pinaceae*), 31.05.2010, G. Doğan (AEUT GD1069).

NOTES: While DiCosmo et al. (1983) resolved the nomenclatural problems surrounding *N. fimbriatus*, the systematic position of this species is still

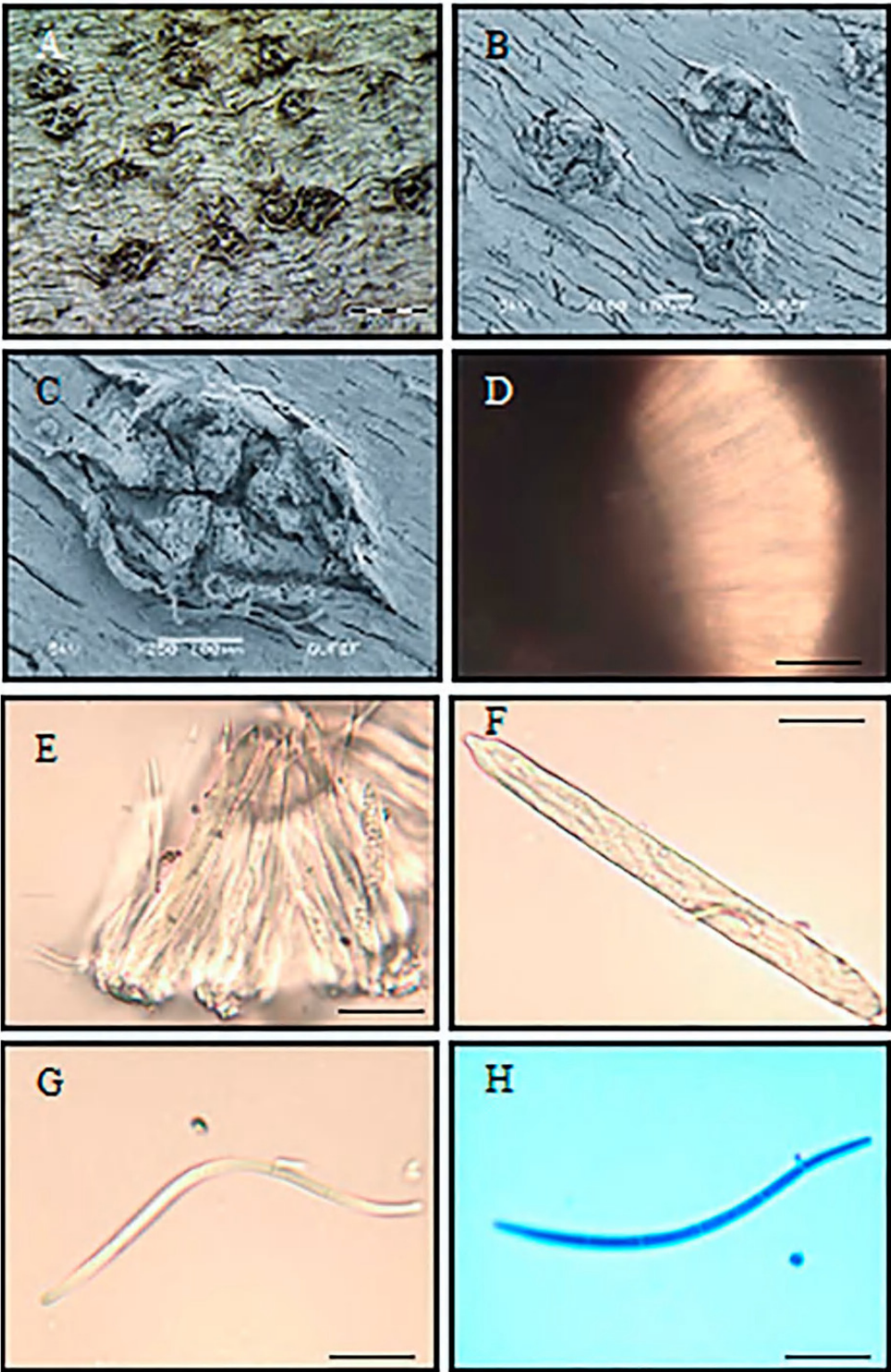


PLATE 1. Microscopic characters of *Naemacyclus fimbriatus*. A: apothecia on cone scales; B, C: apothecia on cone scales (SEM); D: vertical section of an apothecium; E: asci and ascospores; F: ascus and ascospores; G: ascospore; H: lacto phenol cotton blue mount showing ascospore. Scale bars = D: 100  $\mu$ m; E: 20  $\mu$ m; F–H: 15  $\mu$ m.

controversial. *Naemacyclus fimbriatus* was once placed in the *Phacidiaceae*, but DiCosmo (1979) suggested that it is more closely related to *Rhytismataceae* because of J- apical ring and ascospores covered by a mucous sheath (Kirk et al. 2001, Cannon & Kirk 2007). The recent analysis of the partial small subunit rDNA (Hou et al. 2006) indicates that *N. fimbriatus* is closely related to taxa in the *Helotiales*.

Although our Turkish specimen of *Naemacyclus fimbriatus* is morphologically similar to specimens described in literature (DiCosmo et al. 1984, Vujanovic et al. 1998, Hou et al. 2006), it does differ slightly in some aspects, particularly with respect to ascospore size and number of septa: 75–90 × 2–2.5 µm and 7-septate (DiCosmo et al. 1984); 78–91 × 2–2.5 µm and 7-septate (Vujanovic et al. 1998); 65–95 × 1–1.5 µm and (2–)4–6-septate (rarely aseptate) (Hou et al. 2006). The ascospores in the specimen from Turkey are shorter and (2–)5-septate (rarely aseptate).

*Naemacyclus fimbriatus* is widely distributed on *Pinus* spp. in Asia (China), Europe, and North America (Cannon et al. 1985; Dennis 1981; Dudka et al. 2004; Eriksson 2014; Gremmen 1960, as *Lasiostictis fimbriata*; Hanlin 1963, as *Stictis fimbriata*; Lin 2012; Minter 1981a; Sherwood 1979, as *Lasiostictis fimbriata*; Vujanovic et al. 1998). The genus *Naemacyclus* and *N. fimbriatus* are reported for the first time from Turkey.

***Lophodermium juniperinum* (Fr.) De Not., G. Bot. Ital. 2(7–8): 46 (1847) PL. 2**

APOTHECIA hysterioid, elliptical, blister-like, strongly raised above the surface of needles, 600–650 × 200–220 µm, blackish; disc soft, whitish. ASCI cylindrical-clavate, short-stalked, thin-walled, 95–110 × 13.5–15.5 µm, 8-spored; ASCOSPORES fasciculate, filiform, attenuated at both ends, 65–87 × 1.5–3 µm, guttulate, hyaline, with a thin hyaline gelatinous coating. PARAPHYSES slender, filiform, hyaline, swollen and curled at the tip.

SPECIMEN EXAMINED—TURKEY, KAYSERİ, Erciyes mountain, Develi, 2000–2050 m asl., conifer plantation area, on dead leaves of *Juniperus communis* L. (*Cupressaceae*), 25.07.2011, G. Doğan (AEUT GD1106).

NOTES: Ten rhytismataceous ascomycetes from the genera *Coccomyces*, *Colpoma*, *Hypoderma*, *Lophodermium*, *Pseudophacidium*, *Solella*, and *Tryblidiopsis* have been reported on species of *Juniperus*; those on juniper needles are mostly pathogenic, causing needle cast (Hou et al. 2005). *Lophodermium juniperinum* is probably weakly parasitic, capable of endophytic growth in needles for part of its life cycle, like most species of *Lophodermium* (Minter 1981b).

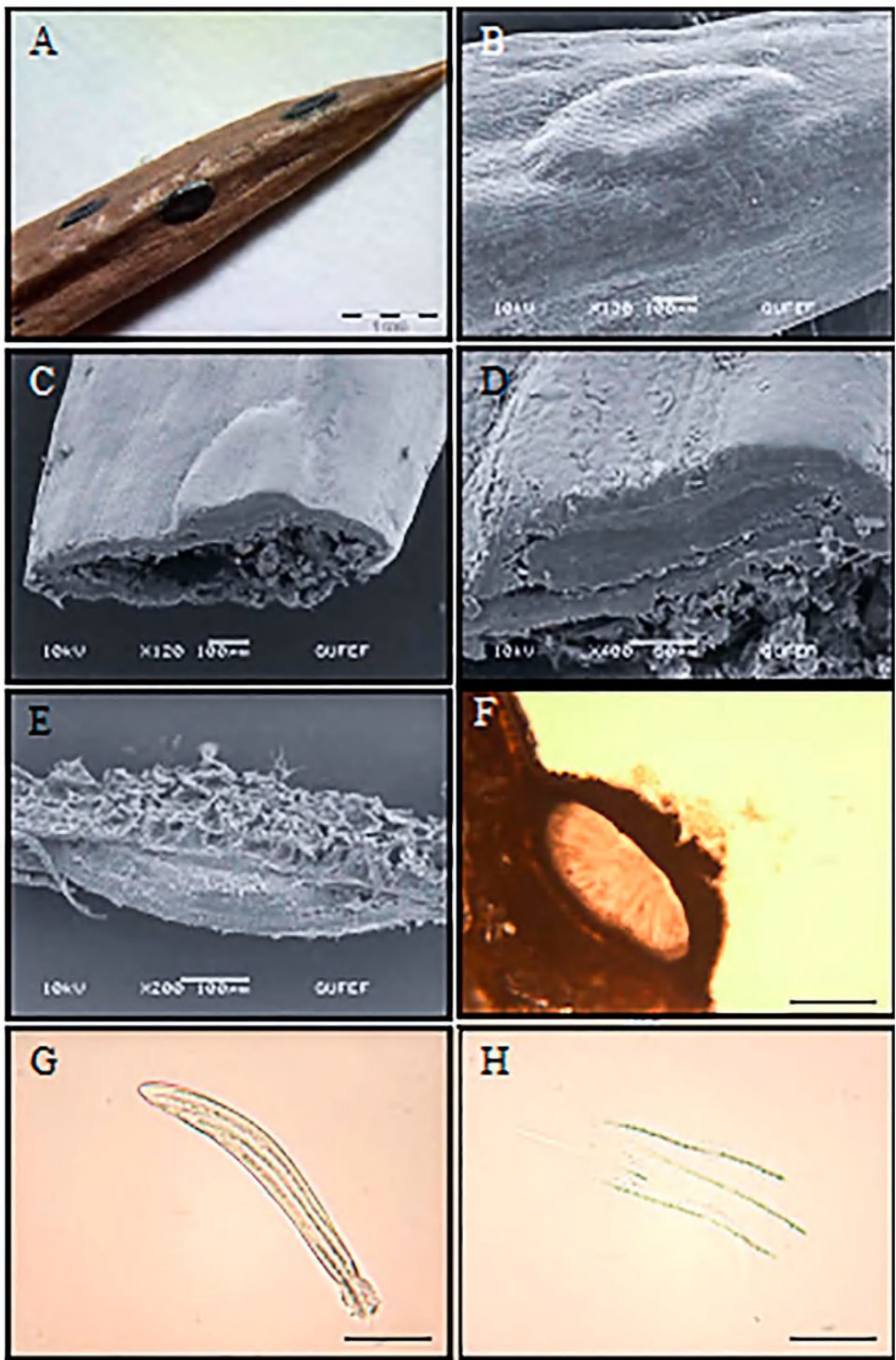


PLATE 2. Microscopic characters of *Lophodermium juniperinum*. A: apothecia on leaf; B: apothecium on leaf (SEM); C-E: vertical section of an apothecium (SEM); F: vertical section of an apothecium; G: ascus and ascospores; H: ascospores. Scale bars = F: 200 µm; G,H: 25 µm.

The Turkish specimen of *L. juniperinum* agrees with other reports of the species in ascomata, asci and ascospores morphology; the only observable difference being the smaller dimensions of ascomata, asci, and ascospores. Dennis (1981) describes  $1 \times 0.4$  mm apothecia,  $130 \times 17$   $\mu\text{m}$  asci, and  $60\text{--}100 \times 2$   $\mu\text{m}$  ascospores; Ellis & Ellis (1987) cite 1 mm long apothecia and  $60\text{--}90 \times 2$   $\mu\text{m}$  ascospores.

*Lophodermium juniperinum* is known from Asia (Dudka et al. 2004), Europe, and North America (Hou et al. 2005). This species is reported for the first time from Turkey.

*Marssonina daphnes* (Roberge ex Desm.) Magnus, Hedwigia 45: 89 (1906) Pl. 3

LEAF SPOTS amphigenous, irregularly rounded, 1–3 mm diam., greenish and then brownish. CONIDIOMATA acervular, amphigenous, at first immersed in host tissue, later erumpent, small, pale brown. CONIDIA ovoid to pyriform, slightly curved,  $15\text{--}20 \times 6.5\text{--}8$   $\mu\text{m}$ , when mature with transverse septum near the base, not constricted, obtuse at the apex, truncate at the base, with numerous guttulations, hyaline.

SPECIMEN EXAMINED—TURKEY, KAYSERİ, Erciyes mountain, Kayseri memorial forest,  $38^{\circ}36'13''\text{N}$   $35^{\circ}30'58''\text{E}$ , 1870 m asl., on living leaves of *Daphne oleoides* Schreb. subsp. *oleoides* (Thymelaeaceae), 25.07.2011, G. Doğan (AEUT GD1080).

NOTES: *Marssonina daphnes* is the causal agent of the daphne anthracnose, which usually causes little damage. The Turkish specimen of *Marssonina daphnes* is morphologically similar to specimens described in literature (Grove 1937, Ellis & Ellis 1987, Ignatavičiūtė & Treigienė 1998). However, the conidia of the Turkish samples are wider than the British collection ( $12\text{--}20 \times 4\text{--}5$   $\mu\text{m}$ ; Grove 1937), while those reported by Ignatavičiūtė & Treigienė (1998) are slightly narrower and shorter than our specimen.

*Marssonina daphnes* is distributed on *Daphne* spp. in Asia (Japan), Australasia, Europe, and North America (Conners 1967; Ginns 1986; Cook & Dubé 1989; Kobayashi 2007; Pennycook & Galloway 2004; Piątek & Wołczańska 2004; Shaw 1973). The species is reported for the first time from Turkey, and *Daphne oleoides* Schreb. subsp. *oleoides* represents a new host record.

#### Acknowledgments

We are grateful to Dr. Eugene Yurchenko, and Sevda Kırbağ for serving as pre-submission reviewers, to Dr. Pennycook for nomenclatural review.

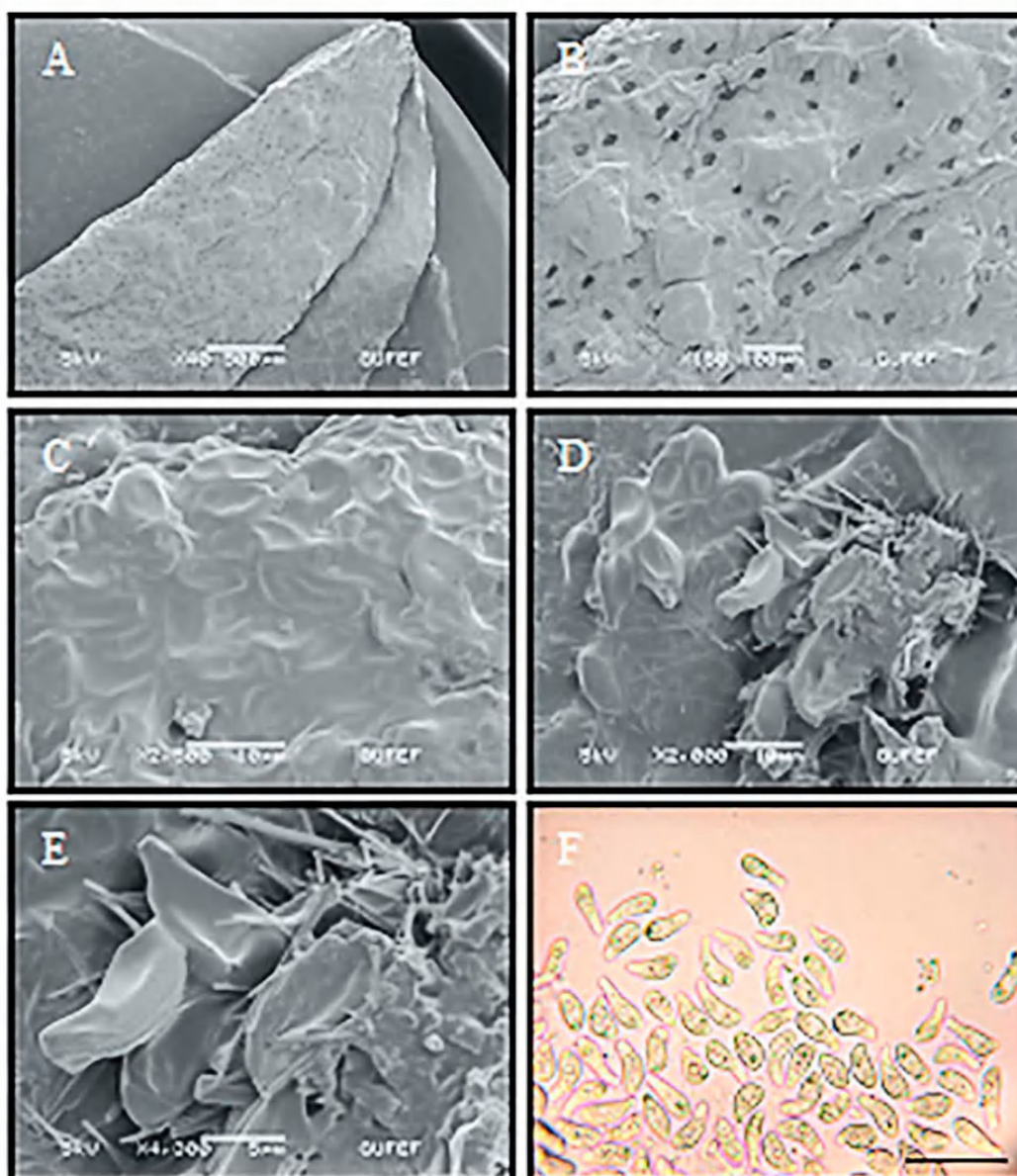


PLATE 3. Microscopic characters of *Marssonina daphnes*. A,B: acervulus on leaf (SEM); C–E: Conidia (SEM); F: Conidia. Scale bar = 10  $\mu$ m.

#### Literature cited

- Cannon PF, Kirk PM. 2007. Fungal families of the world. CAB International Wallingford.  
<https://doi.org/10.1079/9780851998275.0000>
- Cannon PF, Hawksworth DL, Sherwood-Pike MA. 1985. The British *Ascomycotina*. An annotated checklist. Commonwealth Mycological Institute, Kew.
- Connors IL. 1967. An annotated index of plant diseases in Canada and fungi recorded on plants in Alaska, Canada and Greenland. Canada Department of Agriculture, Ottawa.
- Cook RP, Dubé AJ. 1989. Host-pathogen index of plant diseases in South Australia. Field Crops Pathology Group, South Australian Department of Agriculture, Adelaide.

- Davis PH (ed.). 1965–85. Flora of Turkey and East Aegean Islands. Vols 1–9. Edinburgh University Press, Edinburgh.
- Dennis RWG. 1981. British ascomycetes. J. Cramer, Vaduz.
- DiCosmo F. 1979. *Lasiostictis* reassessed. Canadian Journal of Botany 57: 1838–1840. <https://doi.org/10.1139/b79-229>
- DiCosmo F, Peredo H, Minter DW. 1983. *Cyclaneusma* gen. nov., *Naemacyclus* and *Lasiostictis*, a nomenclatural problem solved. European Journal of Forest Pathology 13(4): 206–212. <https://doi.org/10.1111/j.1439-0329.1983.tb00119.x>
- DiCosmo F, Nag Raj TR, Kendrick WB. 1984. A revision of the *Phacidiaceae* and related anamorphs. Mycotaxon 21: 1–234.
- Dudka IO, Heluta VP, Tykhonenko YY, Andrianova TV, Hayova VP, Prydiuk MP, Dzhagan VV, Isikov VP. 2004. Fungi of the Crimean Peninsula. M.G. Kholodny Institute of Botany, Ukraine.
- Ellis MB, Ellis JP. 1987. Microfungi on land plants: an identification handbook. Croom Helm, London–Sydney.
- Eriksson OE. 2014. Checklist of the non-lichenized ascomycetes of Sweden. Symbolae Botanicae Upsalienses 36(2). 501 p.
- Ginns JH. 1986. Compendium of plant disease and decay fungi in Canada 1960–1980. Canadian Government Publishing Centre, Ottawa. <https://doi.org/10.5962/bhl.title.58888>
- Gremmen J. 1960 [“1959”]. A contribution to the mycoflora of pine forests in the Netherlands. Nova Hedwigia 1(3–4): 251–288.
- Grove WB. 1937. British stem- and leaf-fungi (*Coelomycetes*), vol. 1. *Sphaeropsidales*. Cambridge University Press, Cambridge.
- Hanlin RT. 1963. A revision of the ascomycetes of Georgia. Georgia Agricultural Experiment Station, Mimeo Series n.s. 175. 65 p.
- Hou CL, Gao J, Piepenbring M. 2006. Four rhytismataceous ascomycetes on needles of pine from China. Nova Hedwigia 83: 511–522. <https://doi.org/10.1127/0029-5035/2006/0083-0511>
- Hou CL, Lin YR, Piepenbring M. 2005. Species of *Rhytismataceae* on needles of *Juniperus* spp. from China. Canadian Journal of Botany 83: 37–46. <https://doi.org/10.1139/b04-149>
- Ignatavičiūtė M, Treigienė A. 1998. Mycota Lithuaniae, vol. 9. *Melanconiales*. UAB Vaslstiečių Laikraštis, Vilnius.
- Lantz H, Johnston PR, Park D, Minter DW. 2011. Molecular phylogeny reveals a core clade of *Rhytismatales*. Mycologia 103: 57–74. <https://doi.org/10.3852/10-060>
- Kirk PM, Cannon PF, David JC, Stalpers JA. 2001. Ainsworth & Bisby’s dictionary of the fungi. 9th edn. CAB International, Kew.
- Kobayashi T. 2007. Index of fungi inhabiting woody plants in Japan. Host, distribution and literature. Zenkoku-Noson-Kyoiku Kyokai Publishing Co., Tokyo.
- Lin YR. 2012. *Rhytismatales*. Flora Fungorum Sinicorum 40. 261 p.
- Minter DW. 1981a. Microfungi on needles, twigs and cones of pines in Czechoslovakia. Česká Mykologie 35(2): 90–101.
- Minter DW. 1981b. *Lophodermium* on pines. Mycological Papers 147. 54 p.
- Piątek M, Wołczańska A. 2004. Some phytopathogenic fungi rare or new to Poland. Polish Botanical Studies 49(1): 67–72.
- Pennycook SR, Galloway DJ. 2004. Checklist of New Zealand “fungi”. 401–488, in: EHC McKenzie (ed.). Introduction to fungi of New Zealand. Fungi of New Zealand vol. 1. Fungal Diversity Research Series 14.
- Schoch CL, Sung GH, López-Giráldez F, Townsend JP, Miadlikowska J, Hofstetter V, Robbertse B, Matheny PB, Kauff F, Wang Z, Gueidan C, Andrieu RM, Trippel K, Ciufetti

- LM, Wynns A, Fraker E, Hodkinson BP, Bonito G, Groenewald JZ, Arzanlou M, de Hoog GS, Crous PW, Hewitt D, Pfister DH, Peterson K, Gryzenhout M, Wingfield MJ, Aptroot A, Suh SO, Blackwell M, Hillis DM, Griffith GW, Castlebury LA, Rossman AY, Lumbsch HT, Lücking R, Büdel B, Rauhut A, Diederich P, Ertz D, Geiser DM, Hosaka K, Inderbitzin P, Kohlmeyer J, Volkmann-Kohlmeyer B, Mostert L, O'Donnell K, Sipman H, Rogers JD, Shoemaker RA, Sugiyama J, Summerbell RC, Untereiner W, Johnston PR, Stenroos S, Zuccaro A, Dyer PS, Crittenden PD, Cole MS, Hansen K, Trappe JM, Yahr R, Lutzoni F, Spatafora JW. 2009. The *Ascomycota* tree of life: a phylum-wide phylogeny clarifies the origin and evolution of fundamental reproductive and ecological traits. *Systematic Biology* 58: 224–239. <https://doi.org/10.1093/sysbio/syp020>
- Shaw CG. 1973. Host fungus index for the Pacific Northwest – I. Hosts. Washington Agricultural Experimental Station Bulletin 765. 121 p.
- Sherwood MA. 1979. *Phacidiales* Exsiccati. Decades I–III. *Mycotaxon* 10: 241–245.
- Suija A, Ertz D, Lawrey JD, Diederich P. 2015. Multiple origin of the lichenicolous life habit in *Helotiales*, based on nuclear ribosomal sequences. *Fungal Diversity* 70: 55–72. <https://doi.org/10.1007/s13225-014-0287-4>
- Vujanovic V, St.-Arnaud M, Neumann P. 1998. First report of *Naemacyclus fimbriatus* infecting pitch pine (*Pinus rigida*). *Plant Disease* 82: 959. <https://doi.org/10.1094/PDIS.1998.82.8.959A>



Erdoğdu, Makbule et al. 2018. "A contribution to the study of Helotiales and Rhytismatales in Turkey." *Mycotaxon* 132(4), 885–893.

<https://doi.org/10.5248/132.885>.

**View This Item Online:** <https://www.biodiversitylibrary.org/item/335881>

**DOI:** <https://doi.org/10.5248/132.885>

**Permalink:** <https://www.biodiversitylibrary.org/partpdf/393034>

#### **Holding Institution**

Noni Korf

#### **Sponsored by**

Cornell University Library

#### **Copyright & Reuse**

Copyright Status: In copyright. Digitized with the permission of the rights holder.

Rights Holder: Noni Korf

License: <https://creativecommons.org/licenses/by-nc-sa/4.0/>

Rights: <http://biodiversitylibrary.org/permissions>

This document was created from content at the **Biodiversity Heritage Library**, the world's largest open access digital library for biodiversity literature and archives. Visit BHL at <https://www.biodiversitylibrary.org>.